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Review Article

A Mini Systematic Review: Eucheuma cottonii, a Red Algae, as a Radiosensitizer?

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ABSTRACT

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Radiosensitizers assist radiotherapy in providing greater tumor inactivation. Currently there is a search for natural radiosensitizer components which are expected to provide lesser side effects than chemical radiosensitizers. *Eucheuma cottonii* is a plant with antioxidant and anti-tumor effects. This review aims to search for the potential use of *Eucheuma cottonii* as a radiosensitizer. This is a mixed review study, where the main component is a systematic review and then followed by a narrative review. This review suggests that *Eucheuma cottonii* has the potential to become a radiosensitizer, by interfering with the cell cycle control mechanisms and reactivation of p53. Further research is needed to explore the synergistic effect of the combined use of radiotherapy and *Eucheuma cottonii*.

Keywords: Eucheuma cottonii, Anti-tumour, Radiosensitizer, Immunomodulator

Introduction

Cancer is among the world's top causes of death. According to the data of GLOBOCAN, in 2018, there were 18.1 million new cancer cases, 9.6 million of which resulted in death. Globally, 1 out of 5 men and 1 out of 6 women suffer from cancer. The data also shows that 1 out of 8 men and 1 out of 11 women died from cancer.¹

Cancer treatment modalities consist of radiotherapy, surgery, chemotherapy, immunotherapy, and hormone therapy.² Either for curative or palliative purpose, 80% of cancer patients require radiotherapy treatment. An effort to optimize radiation therapy is through the use of radiosensitizer. Radiosensitizer is a compound which, when combined with radiation, will provide greater tumour inactivation than the additive effect of each modality.^{3,4} The majority of radiosensitizers are synthetic chemical compounds, which have been proven to be too toxic in effective clinical doses. Radiosensitizer made of natural materials is believed to be safer than synthetic materials.5 Eucheuma cottonii, is commonly found in the Indonesian ocean. Eucheuma cottonii (Kappaphycus alvarezi) is used to produce carrageenan that functions as a stabilizer, a gellant, binder, thickening agent, and supplements in the pharmaceutical and food industries.⁶ It has high number of proteins, dietary fibers, antioxidants, vitamins, polyphenols, phytochemicals, minerals, and polyunsaturated fatty acids. It also has many medical uses.8 Many studies have reported the anticancer effect of Eucheuma cottonii, either in vitro or in vivo. However, there have been no studies that observe the combination of radiation and Eucheuma cottonii in cancer cell lines. Therefore, the present study aims to investigate the radiosensitization effect of Eucheuma cottonii.

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Method

Literature search was conducted on the PubMed, Cochrane, EBSCO, and SCOPUS databases for articles published from 2010 to 2020 on studies on the effects of Eucheuma cottonii on cancer cells. Search was performed using search terms (Kappaphycus alvarezii OR Eucheuma cottonii OR Red seaweed) and (cytotoxic OR antiproliferative OR anticancer OR celldeath) and (tumor OR cancer). Selected references were assessed based on relevance, and suitability for writing purpose. The literature search inclusion criteria included experimental studies that discussed the effects of Eucheuma cottonii/ Kappaphycus alvarezii on cancer cells in vitro and in vivo. The exclusion criteria used were publication with the type of review, used different algae species, or in combination with other substances. The risk of biased in vivo studies was conducted using SYRCLE. In vitro studies were analyzed using SciRAP. Literature searches and critical reviews were conducted independently by two reviewers. We carried out a narrative review to analyze the role of Eucheuma cottonii on the effects of radiation. We made a flow diagram in agreement with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement on systematic review reporting (see Figure 1).

Results and Discussion

Thirteen articles were involved in this systematic review. Eleven articles were *in vitro* studies, and four articles were *in vivo* studies. We made a summary about the type of research, type of cell line, and the results from all studies are presented in Table 1.

Anticancer/antiproliferative/cytotoxic activity

Eleven *in vitro* studies were conducted using 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) test to observe the cytotoxic effect. They are listed in Table 2. Table 2 contains cell types, solvents and IC₅₀ values from *in vitro* studies. The inhibitor activity is expressed in Inhibitory Concentration 50% (IC₅₀). IC₅₀ is the extract concentration that can inhibit cell growth by 50%. The results of which proved that *Eucheuma cottonii* extract inhibit the growth of cancer cells. However, the IC₅₀ values obtained were different with wide variations (IC₅₀ = 20 µg/mL to 4 mg/mL).

This can occur because the procedures, reagents, and passage cells used in each study are different.

Proapoptotic and anti-apoptotic protein regulation

Several main target proteins for radiosensitization were seen in cancer cells treated with *Eucheuma cottonii*. *Eucheuma cottonii* extract was able to restore the apoptotic response of cancer cells by increasing the expression of p53 and Chk1 (Figure 2). This extract inhibited the expression of the antiapoptotic gene BIRC5 (Baculoviral IAP repeat-containing protein) and Bag1 and reduced the expression of MDM2 (mouse double minute 2 homologous).¹⁸ The inhibition of MDM2 which normally binds to p53 will lead to the stabilization of p53 and CDK6 activity resulting in a termination of the cell cycle at the G1 phase.^{21,22} In addition to being able to phosphorylate p53, an increase in Chk1 will also phosphorylate CDC25 (cell division cycle 25), thus inhibiting CDK2-cyclin E and CDK1-cyclin B which results in a cessation of the cell cycle at the G1 and G2 phases.²¹

Cell cycle profiles and cell development were seen using FUCCI (fluorescence ubiquitination-based cell-cycle indicator).¹⁶ Cells treated with k-carrageenan (k.CO) showed a rise in the number of cells that stopped in the G2/M phase. Giving k-CO to cells results in a G2 / M phase twice as long as the G1 phase. The cessation of the cell cycle is likely a continuation of the p53 activation response which activates p21 as a cyclin-dependent kinase (CDK) inhibitor that induces the cessation of the cell cycle.²¹ This greatly helps the role of radiation, where the G2/M phase is known to be sensitive to radiation. BIRC5 or survivin is an inhibitor of apoptosis (IAP) member which works to inhibit caspase and inhibits cell death. BIRC5 is found in most cancer cells and has been associated with poor clinical outcomes. Survivin expression is very high in the G2/M phase and decreases swiftly in the G1 phase of the cell cycle. Survivin has been proven to inhibit apoptosis via a caspase-dependent and independent pathway. Survivin inhibits caspases 3, 7 and 9 and inhibits apoptosis-inducing factor (AIF) which is released from the mitochondrial intermembrane space into the cytoplasm. Surviving expression can be suppressed by the p53-E2F complex, by directly binding to the surviving promoter. In other words, survivin can also influence p53 activity through regulation of MDM2 and proteosome.²³

Bag1 (BCL2 associated Athano Gene-1) inhibits cell death by synergistic action with the antiapoptotic gene BCL2. Bag 1 works to regulate the integrity of the mitochondrial membrane and prevent caspase activation.²⁴ The potential pathways for the radiosensitizer effect of *Eucheuma cottonii* can be seen in Figure 3.

Immunomodulator

The electron microscope imaging shows the presence of macrophage activity, which suggests that the extract modulates the immune response. From the TEM (Transmission Electron Microscopy) examination after administration of the extract, monocytes and macrophages were seen. The macrophages responsible for phagocytosis are the main immune response associated with antigens on the surface of tumor cells. The presence of monocytes activates dendritic cells (DCs), which together with macrophages present CD4 and CD8 T cell antigens, so that CD4 and CD8 are activated. When CD4 and CD8 enter the circulation, it destroys tumour cells.^{25,26,27} The extract was also shown to reduce IL4 and increase IFNy.18 Cytokines IL4 and IFNy have a major contribution in the regulation and formation of immune responses. Macrophage activity will increase due to the influence of IFNy and inhibited by IL4.28 The immunomodulator pathway of Eucheuma cottonii is shown in Figure 4.

Survival pathway

Eucheuma cottonii extract can increase NF-κB in cells.¹⁸ NF-κB can increase anti-apoptotic protein expression. NF-κB also induces expression of IAP and several members of the anti-apoptotic family Bcl-2.²⁹ This raises the question of how *Eucheuma cottonii* can cause cancer cell death, as has been proven from the MTT test results and the visible characteristics of apoptotic cells after administration of *Eucheuma cottonii*. Another thing to keep in mind is that increasing Chk1 and IFNγ results in the upregulation of PDL-1 (Programmed Death Ligand 1).³⁰ PDL-1 is a PD-1 (Programmed Cell Death 1) ligand. PD1/PDL-1 is an immune checkpoint that causes tumour cells to avoid CD8.³¹ It is not known how *Eucheuma cottonii* affects PD-1/PDL-1.



Figure 1: Searching strategy using PRISMA flow diagram



Figure 2: Biomolecular pathway effect of *Eucheuma cottoni*. *Eucheuma cottonii* can decrease the anti-apoptotic gene (Birc5, Bag 1 and MDM2), and increase the proapoptotic gene (Chk1 and p53)¹⁸

Studies	Type of studies	Cell line	Result	
Studies Namvar,2012 ⁷	Type of studies In vitro/non randomized	Cell line MCF-7 (breast cancer cell estrogen dependent) MB-MDA-231(breast cancer cell estrogen independent) 	Result In vitro test found that ECME (<i>Eucheuma cottonii</i> polyphenol-rich extract) was anti-proliferative against estrogen-dependent MCF-7 and estrogen-independent MB-MDA-231 human breast-cancer cells (IC ₅₀ values of 20 and 42 µg/mL, respectively) but was non-toxic to normal cell lines.	
	In vivo/	 A normal African green monkey kidney Vero cell line Rat mammary tumour 	Eucheuma cottonii hindered tumor growth and erythrocyte lipid	
	randomized	was induced with LA7 cells	peroxidation in the cancer-induced rats, dose-dependently. The histopathology observation and electron microscopy affirmed apoptosis: cell shrinkage, DNA (deoxyribonucleic acid) fragmentations, cell membrane blebbing, microvillus disappearance or reduction, condensation of chromosomes and apoptotic bodies with complete membrane and activation of the caspase cascade, in the rat mammary tumours.	
Lee, 2013 ⁹	<i>In vitro/</i> non randomized	HeLa (Human Cervix Adeno carcinoma)	MTT (3-(4,5-Dimethylthiazol-2,5-diphenyltetrazoliumbromide) cell viability inspection displayed that various concentrations of the crude extracts from <i>Eucheuma cottonii</i> hindered the HeLa cells for development after 24 hours and 48 hours being incubated with crude extracts from <i>Eucheuma cottonii</i> . The concentration of extracts from <i>Eucheuma cottonii</i> 0.5, 0.7, 0.9, and 1.0 mg/mL for 24 hours indicated that HeLa cells have experienced significant DNA fragmentation. The HeLa cells induced by a concentration of 20 mg/mL 24 hours did not	

Table 1: The Summary of the reviewed studies

			undergo apoptosis.
			The induction of apoptosis by a concentration of 0.1 mg/mL for 48 hours
			has degraded.
Shamsahadi	In vivo/	LA7	The oral administration 100 mg/kg body-weight of ECE (<i>Eucheuma</i>
2013 ¹⁰	randomized		cattanii athanol avtract) was correlated with tamovifan (10 mg/kg body
2015	Tanuomizeu		conomic entation extractly was contracted with tamoxinen (10 mg/kg body-
			weight). Subcutaneous injection of LA-7 cells (6 \times 10° cells/rat) was
			performed on the rat to develop mammary tumor. The ECE was proven to
			have better effectiveness than tamoxifen in tumor development suppression
			(27%), tissues improvement (plasma, liver, and kidney) malondialdehyde
			concentrations, superoxide dismutase activity and erythrocyte glutathione
			concentrations ($P < 0.05$)
			Unlike temperior the ECE showed little toricity to the liver and bidness
			United amovinen, the ECE showed fittle toxicity to the fiver and kidneys.
Tan, 2014 ¹¹	<i>In vitro</i> /non	MCF7	The cytotoxic effect of the extract was decided through the MTT test. Cell
	randomized		apoptosis was detected using GeneTex Enhanced Apoptotic DNA Ladder
			Detection Kit. The Result of the MTT test shows that MCF-7 cancer cells
			growth was suppressed by the crude extracts of Eucheuma cottonii in a
			dose-dependent manner of IC ₅₀ at 3.5 mg/mL (24h) and 0.85 mg/mL (48h).
			The presence of fragmented DNA of cells treated with the crude extract
			indicated autotoxic effect is through exertacia
z 201 1 ¹²			
Lau, 2014 ¹²	In Vitro	HeLa	The extract's cytotoxic effect was tested by the MTT test. Kappaphycus
			alvarezii was extracted with 90% methanol, 70% acetone, and aqua. The
			90% methanol extract showed an anti-proliferative effect at 200-500
			µg/mL. The aqua extract showed no anti-proliferative effect.
Lee, 2015 ¹³	In vitro/non	HeLa (Human Cervix	By using the maximum dose of the extract (20.0mg/mL) for 24 hours
	randomized	Adeno	incubation a complete cessation in cell proliferation was observed
	Tundonnized		showing a significant autotoxia affect of Euchauma actionii avtract
		carcinoma)	showing a significant cytotoxic effect of <i>Eucneuma conomi</i> extract.
		Human lung carcinoma	The extracts of <i>Eucheuma cottonii</i> showed no effect on fibroblast, a human
		cell line (SKLU-1)	normal cell line.
		Human colon carcinoma	
		cell line (HCT-116)	
		Fibroblastwere	
Suganya 2016 ¹⁴	In vitro/non	Breast cancer (MCF7)	The pharmacological properties of native carrageenan (k) that was extracted
Sugariya, 2010	rondomized	colon (UT 20) liver (Hen	from Kannanbuqua abuquazii and commercial corresponden (Sigma Aldrich)
	Tandonnized	cololi (H1-29), livel (Hep	noin <i>Kappaphycus alvarezu</i> and commercial carrageenan (Sigma-Aldrich)
		G2) and osteosarcoma	were then evaluated. Native carrageenan exhibited an excellent anticancer
		(MG63).	activity on colon carcinoma cell lines (67.66 $\pm 0.168\%$) with the IC ₅₀ value
			of 73.87 mg/ml, and commercial carrageenan possessed a potent hindrance
			on breast cancer cell lines growth (67.33 \pm 0.077%) with the IC_{50} value of
			123.8 mg/mL, liver cancer cell line (Hep G2) with the IC ₅₀ values of both
			native and commercial carrageenans determined as 56.71 and 125 mg/mL,
			respectively. Colon cancer cell line (HT-29) with ICro value was observed
			as 72.87 and 122.8mg/mL for notive and commercial commercial
			as 75.87 and 125.8 mg/mL for native and commercial carrageenans,
			respectively. Osteosarcoma cancer cell line, with IC_{50} of 4/.85 and 55.48
			mg/mL for native and commercial carrageenans, respectively.
Arsianti, 2016 ¹⁵	In vitro	MCF-7 and HCT-116	Eucheuma cottonii extract was found to hinder the proliferation of MCF-7
			and HCT-116 cells
Prasedya, 2016 ¹⁶	In vitro/non	human cervical carcinoma	Decreased the cell viability of HeLa cells exposed with k-CO and $\lambda\text{-CO}$
	randomized	cells (HeLa) cells as and	over a 72 h period. Both k-CO and λ -CO showed IC ₅₀ values of

		human umbilical vein endothelial cells (HUVEC)	550.8µg/mL and 475 ± 12 µg/mL. Both carrageenans had no significant cytotoxic effect on HUVEC. Cell cycle profiles of k-CO treated cells showed increased arrest in the G2/M phase. Cells treated with λ -CO needed a longer time to finish one cycle, around 59 ± 4.6 hours and with k-CO treatment which was 50.2 ± 2.9 hours. Most cells treated with λ -CO were unable to perform cell division. The cell cycle in cells treated with λ -CO, the progress continues as FUCCI cells change color, except the cells cannot divide and would later die. Cells treated with k-CO were able to divide at least once before cell death
Chang, 2017 ¹⁷	<i>In vitro</i> /non randomized <i>In vivo</i> / randomized	Breast cancer cell line (MCF-7) Investigated toxicity effect of high dosage <i>Kappaphicus alvarezii</i> extracts in rats and determined the effect of <i>Kappaphicus alvarezii</i> on 7, 12-dimethylbenz[a] anthracene (DMBA) mammary carcinogenesis in rats	The tumor growth rate in the untreated group of mice was found necessarily higher than the experimental group of rats. The specific tumor growth rate in the untreated group of mice was found necessarily higher than the experimental group of $\pm 0.270 \text{ mm}^3/\text{t}$ and <i>Kappaphicus alvarezii</i> extract treated group is $0.097 \pm 0.060 \text{ mm}^3/\text{t}$.
Bakar, 2017 ¹⁸	In vivo/ randomized	Mammary tumor was induced by subcutaneously injecting LA7 cells in female rat mammary pads	 After 2 weeks of cancer development, the mice were orally-administered with either SECE (Seaweed <i>Eucheuma cottonii</i> ethanol extract) 150 mg/kg body weight (BW) and 300 mg/kg (BW) or tamoxifen. The Electron microscopy-imaging results affirmed the presence of macrophage activity. Hematoxylin and eosin staining showed that the seaweed extract restored the tumor histopathological alterations to normal. The extract hindered tumor growth and regulated the immune responses. This was proven by the microscopic observations, the increased spleen weight, size, spleen CD19 B cells, and blood immunoglobulin G (IgG) levels. The extract also raised the circulating total white blood cells, lymphocytes, segmented neutrophils count, T cells (CD3), T-helper cells (CD4), cytotoxic T cell (CD8), and nuclear factor-kappa beta expressions. The extract raised cancer cell death, by upregulating the Birc5, Chk1, and p53 levels and downregulated the tumor development cellular Mdm2 (transformed mouse 3T3 cell double minute 2) messenger RNA (mRNA) expression. The extract did not show toxicity at 150 mg/kg BW in rats. The lectin-rich SECE displayed suppression of tumor by developing immune responses and upregulating the canver cell apoptosis mRNA expressions.
Arsianti, 2018 ¹⁹	In vitro	HeLa	Ethanol, n-hexane, chloroform, and ethyl acetate extracts of <i>Eucheuma cottonii</i> displayed strong cytotoxic activity against cervical HeLa cells with IC_{50} of 7.54 µg/mL, 5.73 µg/mL, 4.82 µg/mL and 4.34 µg/mL
Arsianti, 2020 ²⁰	In Vitro	A-549	The cytotoxic effect of Eucheuma cottonii was tested by MTT, controlled

by cisplatin. *Eucheuma cottonii* extract with ethanol, ethyl acetate, and nhexane solvents was shown to inhibit proliferation, with IC_{50} of 251.73 µg/mL, 261.41 µg/mL, 3508 µg/mL.

The antioxidant effect was tested by comparing it with ascorbic acid. The ethanol extract reduced DDDH free radicals with IC₅₀ of 559.76 μ g/mL.

Studies	Cell line	Solvent	Time Incubated (hours)	IC ₅₀
			24	$25\pm0.1\mu\text{g/ml}$
	MCF7		48	$22\pm0.3\mu\text{g/ml}$
Normal 2012^7		Mathemal	72	$20\pm0.2\mu\text{g/ml}$
Namvar, 2012		Methanol	24	$50\pm0.4\mu g/ml$
	MB-MDA231		48	$50\pm0.6\mu g/ml$
			72	$42\pm0.3\mu\text{g/ml}$
1 0010 ⁹			24	N 7.4
Lee, 2013	HeLa	Methanol	48	NA
T 2014			24	3.5mg/ml
Tan,2014	MCF/	Methanol	48	0.85mg/ml
		Methanol	24	
Lau, 2014 ¹²	HeLa	Aceton	24	NA
		Aqua	24	
	HeLa			$\pm 0.5 \text{mg/ml}$
Lee,2015 ¹³	SK-Lu1	Methanol	24	$\pm 0.5 \text{mg/ml}$
	HCT116			$\pm 0.5 \text{mg/ml}$
	MCF7			103.2 µg/ml
$201c^{14}$	HT29	NT A	NA	73.87µg/ml
Suganya, 2016	HepG2	NA		56.71µg/ml
	MG63			47.85µg/ml
Prasedya, 2016 ¹⁶	II.I .	NA	72	550.8 ± 7.6
	HeLa			µg/ml
Arsianti, 2016 ¹⁵	DT-ethanol			$149,5 \pm 2.8$
		D1-ethanol		µg/mL
		DU-Ethanol		$75.7\pm1.3~\mu\text{g/mL}$
			oform 4 exane acetate	189.0 ± 2.2
	MCF7 Chloroform DT Hexane Ethyl acetate	Chloroform		µg/mL
		DTH		111.5 ± 1.9
		DI Hexane		µg/mL
				259.0 ± 2.6
		Etnyl acetate		µg/mL
		DT-ethanol		$65.3\pm1.8~\mu\text{g/mL}$
		DU Ethanol		419.1 ± 2.7
	ИСТ 116	DU-Ethanol	4	µg/mL
	1101 110	Chloroform		$99.3 \pm 1.8 \ \mu\text{g/mL}$
		DT Hexane		$43.0\pm1.3~\mu\text{g/mL}$
		Ethyl acetate		$21.4 \pm 1.4 \; \mu g/mL$
Chang, 2017 ¹⁷	MCF7	Methanol	24	4.1 ± 0.69

Table 2: The IC₅₀ value of *Eucheuma cottonii's* extract in the included studies

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				mg/mL
		Etanol	48	7.54 µg/ml
Assignt: 2018^{19}	HeLa	n-Hexana		5.73 µg/ml
Arsianti, 2018		Chloroform		4.82 µg/ml
		Etil asetat		4.34 µg/ml
	A-549	Etanol		251.73µg /mL
Arsianti, 2020 ²⁰		Ethil asetat	4	261.41µg / mL
		n-Hexana		$3508 \mu g \ / \ mL$



Figure 3: Radiosensitizer mechanism of *Eucheuma cottonii* (EC = *Eucheuma cottonii*, black line shows radiation effect, red line shows *Eucheuma cottonii* effect, arrow shows activating effect, T model shows inhibiting effect)^{18,21-24}



Figure 4: Immunomodulator effect of *Eucheuma cottonii* by increasing monocytes and regulating IL4 and IFN_γ.^{18,24,25}

Conclusion

Our systematic review and further analysis of several selected pathway found that *Eucheuma cottonii* is able to suppress the proliferation of various cancer cells and cause cancer cell death in the form of apoptosis through regulation of proapoptotic proteins (Chk1 and p53) and anti-apoptosis (Birc5, Bag 1 and MDM 2) and also interfere with the cell cycle control mechanisms by a cessation of the cell cycle at the G2/M phase. We discovered the future potential of *Eucheuma cottonii* as a promising radiosensitizer. All journals in this systematic review are still preclinical trials of the effects of *Eucheuma cottonii* extract on cancer cells. Further studies are needed to determine whether the extract can synergize with radiation as a radiosensitizer.

Conflict of Interest Disclosure

Authors declare no conflicts of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

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References

- 1. New Global Cancer Data: GLOBOCAN 2018 | UICC [Internet]. [cited 2020 Nov 3]. Available from: https://www.uicc.org/news/new-global-cancer-dataglobocan-2018
- Miller KD, Nogueira L, Mariotto AB, Rowland JH, Yabroff KR, Alfano CM, Jemal A, Kramer JL, Siegel RL. Cancer treatment and survivorship statistics, 2019. CA Cancer J Clin. 2019; 69(5):363-385.
- Kuruba V and Gollapalli P. Natural radioprotectors and their impact on cancer drug discovery. Rad Oncol J. 2018; 36(4):265-275.
- Van Weelden,W.J., Sekarutami,S.M., Bekkers,R.L.M, Kaanders,J.H., Bussink J, Gondhowiardjo SA, Leer,J.W. The Effect of Carbogen Breathing and Nicotinamide Added to Standard (Chemo)Radiation Treatment of Advanced Cervical Cancer in Indonesia. Int J Gynecol Cancer. 2014;24:1628-1635.
- Malik A, Sultana M, Qazi A, Qazi MH, Parveen G, Waquar S, Ashraf AB, Rasool M. Role of Natural Radiosensitizers and Cancer Cell Radioresistance: An Update. Anal Cell Pathol. 2016; 2016:1-8.
- Liu Z, Gao T, Yang Y, Meng F, Zhan F, Jiang Q, Sun X. Anti-Cancer Activity of Porphyran and Carrageenan from Red Seaweeds. Molecules 2019; 24(23):1-14.
- Namvar F, Mohamed S, Ghasemi S, Behravan J. Polyphenolrich seaweed (*Eucheuma cottonii*) extract suppresses breast tumour via hormone modulation and apoptosis induction. Food Chem. 2012; 130(2):376-82.
- Noor A, Gunasekaran S, Vijayalakshmi MA. Article in Pharmacognosy Research · October 2017. Pharmacog Res. 2018; 10:24-30.
- Lee JW and Teo SS. *In-vitro* Cytotoxicity and Anticancer Activity of *Eucheuma Cottonii* Extracts Against Hela Cell Line. Malay J Sci. 2013; 32:189-196.

- Shamsabadi FT, Khoddami A, Shamsabadi FT. Comparison of Tamoxifen with Edible Seaweed (*Eucheuma cottonii* L) Extract in Suppressing Breast Tumor. Nutrition and Cancer.2013;65(2):255-262.
- Chern HT and Vi-Sion Chang SST. Cytotoxic Activity of Eucheuma Cottonii on MCF-7 Human Breast Cancer. Malay J Sci. 2014; 33(2):155-162.
- Ying LT, Fenny VD, Sze YCC, Thau LYW. Antiproliferative Potential of Extracts from Kappaphycus Seaweeds on HeLa Cancer Cell Lines. Sains Malaysiana. 2014; 43(12):1895-1900.
- Lee JW, Wang JH, Ng KM, Tan-Rabina P, Teo S. *In-vitro* Anticancer Activity of *Eucheuma Cottonii* Extracts Againts Hela Cell Line, HUMN Lung Carcinoma Cell Line (SK-LU-1), Human Colon Carcinoma Cell Line (HCT-116), and Fibroblast. IJCMS. 2015; 1(2):69-73.
- Suganya AM, Sanjivkumar M, Chandran MN, Palavesam A, Immanuel G. Pharmacological importance of sulphated polysaccharide carrageenan from red seaweed Kappaphycus alvarezii in comparison with commercial carrageenan. Biomed Pharmacother. 2016; 84:1300-1312.
- 15. Arsianti AA, Fadilah F, Fatmawaty Y, Wibisono LK, Kusmardi S, Azizah NN, Putrianingsih R, Murniasih T, Rasyid A, Pangestuti R. Phytochemical composition and anticancer activity of seaweeds *Ulva lactuca* and *Eucheuma cottonii* against breast MCF-7 and colon HCT-116 cells. Asian J Pharm Clin Res. 2016; 9(6):115-9.
- Prasedya ES, Miyake M, Kobayashi D, Hazama A. Carrageenan delays cell cycle progression in human cancer cells *in vitro* demonstrated by FUCCI imaging. BMC Compl Altern Med. 2016; 16(1):1-9.
- Chang VS, Okechukwu PN, Teo SS. The properties of red seaweed (*Kappaphycus alvarezii*) and its effect on mammary carcinogenesis. Biomed Pharmacother. 2017; 87:296-301.
- Bakar N, Tengku IT, Mohamad SN, Mohamed S. Changes in rats' breast tumor ultrastructure and immune and messenger RNA responses caused by dietary Seaweed (*Kappaphycus alvarezii*) extract. J Microsc Ultrastruct. 2017; 5(2):70.
- Arsianti A, Aziza YAN, Kurniasari KD, Mandasari BKD, Masita R, Zulfa FR, Dewi MK, Zagloel CRZ, Azizah NN, Putrianingsih R. Phytochemical test and cytotoxic activity of macroalgae *Eucheuma cottonii* against cervical HeLa cells. Pharmacog J. 2018; 10(5):1012-1017.
- Arsianti A, Kurniawan G, Tejaputri NA, Qorina F, Fithrotunnisa Q, Azizah NN, Fajrin AM. Phytochemical Profile, Antioxidant Activity and Cell Line Study of Marine Red Macroalgae *Eucheuma cottonii* on Lung A-549 Cancer Cells. Pharmacogn J. 2020; 12(2):276-281.
- 21. Maier P, Hartmann L, Wenz F, Herskind C. Cellular pathways in response to ionizing radiation and their targetability for tumor radiosensitization. Int J Mol Sci. 2016; 17(1):102.
- Lu M, Breyssens H, Salter V, Zhong S, Hu Y, Baer C, Ratnayaka I, Sullivan A, Brown NR, Endicott J, Knapp S, Kessler BM, Middleton MR, Siebold C, Jones EY, Sviderskaya EV, Cebon J, John T, Caballero OL, Goding CR, Lu X. Restoring p53 Function in Human Melanoma Cells by Inhibiting MDM2 and Cyclin B1/CDK1-Phosphorylated Nuclear iASPP. Cancer Cell. 2013; 23(5):618-633.
- 23. Mittal R, Jaiswal P, Goel A. Survivin: A molecular biomarker in cancer. Indian J Med Res. 2015; 141:389.
- 24. Aveic S, Pigazzi M, Basso G. BAG1: The guardian of antiapoptotic proteins in acute myeloid leukemia. PLoS One. 2011; 6(10):e26907.
- 25. Shiao SL, Preethi GA, Rugo HS, Coussens LM. Immune microenvironments in solid tumors: New targets for therapy. Genes Dev. 2011; 25(24):2559-2572.
- 26. Gondhowiardjo SA, Handoko, Adham M, Rachmadi L, Kodrat H, Tobing DL, Haryoga IM, Dwiyono AG, Kristian YA, Permata TBM. Tumor Microenvironment Predicts

Local Tumor Extensivemess in PD-L1 Positive Nasopharingeal Cancer. PLoS ONE. 2020;15(3):1-10.

- Gondhowiardjo SA, Handoko, Jayalie VF, Apriantoni R, Barata AR, Senoaji F, Utami IGAAJW, Maubere F, Nuryadi E, Giselvania A. Tackling Resistence to Cancer Immunotherapy: What Do We Know?. Molecules. 2020;25(18):4096
- Smith TD, Tse MJ, Read EL, Liu WF. Regulation of macrophage polarization and plasticity by complex activation signals. Integr Biol (United Kingdom). 2016; 8(9):946-955.
- 29. Plewka D, Plewka A, Miskiewicz A, Morek M, Bogunia E.

Nuclear factor-kappa B as potential therapeutic target in human colon cancer. J Cancer Res Ther. 2018; 14(3):516-520.

- Sato H, Niimi A, Yasuhara T, Permata TBM, Hagiwara Y, Isono M, Nuryadi E, Sekine R, Oike T, Kakoti S, Yoshimoto Y, Held KD, Suzuki Y, Kono K, Miyagawa K, Nakano T, Shibata A. DNA double-strand break repair pathway regulates PD-L1 expression in cancer cells. Nat Commun. 2017; 8(1):1-11.
- Buchbinder EI and Desai A. CTLA-4 and PD-1 pathways similarities, differences, and implications of their inhibition. Am J Clin Oncol Cancer Clin Trials. 2016; 39(1):98-106.