

**Comparative Antibacterial Activity of Biosynthesized Silver Nanoparticles and Aqueous Extract of Unripe Pawpaw Peels Against *E.coli***Temitayo I. Adesipe^{1,2*} and Abiodun H. Adebayo^{1,3}¹Dept. of Biochemistry, College of Science and Technology Covenant University, Ota, Ogun State, Nigeria²Dept. of Science Laboratory Technology, Federal Polytechnic Ilaro, Ogun state, Nigeria³Vice Chancellor, Covenant University, Ota Ogun State, Nigeria

ARTICLE INFO

Article history:

Received 12 November 2020

Revised 21 January 2021

Accepted 17 February 2021

Published online 01 March 2021

ABSTRACT

One of the most important issues of current medicine is bacterial drug resistance as well as uropathogenic tract infections and the primary factor of these issues are extraintestinal pathogenic *E. coli* strains. This study examined and compared the antibacterial activity of unripe pawpaw peel aqueous extract (UPPA), biosynthesized silver nanoparticles (b-AgNPs) and Ampicillin (Positive control) against *E.coli*. AgNPs was prepared by reducing 1mM aqueous AgNO₃ solution with UPPA. Primary characterization of synthesized AgNPs was carried out using UV-visible spectroscopy. The total phenol content (TPC) and antibacterial activities of the samples (UPPA and AgNPs) were carried out following standard analytical methods. Synthesis of AgNPs was confirmed by a color change of the reaction mixture to brown after 24hrs of incubation at room temperature, UV-visible spectroscopy also revealed an absorption maxima at 450 nm which is characteristic of AgNPs. The result of this study shows that the biosynthesized AgNPs contains a higher amount of total phenolics and consequently showed a greater antibacterial activity than the unripe pawpaw peel extract alone based on the zone of inhibition exhibited by both samples on the test organism used. However, no zone of inhibition was exhibited by the standard drug (Ampicillin) against the test organism. This study therefore indicates that the antibacterial performance of aqueous extract of unripe pawpaw peels in combating drug resistant pathogenic organisms could be improved to a great extent if incorporated for the synthesis of AgNPs.

Keywords: Silver nanoparticles, Unripe pawpaw peels, Total Phenol content, Uropathogenic *E. coli*.

Copyright: © 2021 Adesipe and Adebayo. This is an open-access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Introduction

Uropathogenic *E.coli* (UPEC) is a common pathotype of extraintestinal pathogenic *E.coli* (ExPEC) which greatly emanates from intestinal microbiota.¹ ExPEC has been implicated to cause bacteremia in older adults, meningitis in neonates and the majority of urinary tract infections (UTIs) in young healthy women.² Estimation shows that 40-50% of women and 5% of men will develop a UTI in their life time.³ However, an increase in morbidity and mortality rates caused by UTI are due to untreated or mistreated UTI which results in kidney and systemic infections.¹ Also the extended usage of last-line antibiotics as a result of recurrence of infection ultimately causes resistance to antibiotics.⁴ This rise in antibiotics resistance of ExPEC has hindered the effective management of UTI.⁵ The tactics used by this pathogenic bacteria to evolve antibiotics resistance includes active efflux of drugs, variation of target sites and degradation of enzymes.⁶ Since medicinal plants contain a wide variety of secondary metabolites, they are being promoted as alternate source of antimicrobials and resistance modifying agents.⁷⁻⁹ Recently, phenolic compounds found in plant has been implied to show significant antimicrobial activity,¹⁰⁻¹² and also have the ability to attack multidrug

resistant bacteria.^{13,14} For example, an aqueous extract of unripe pawpaw peel has been reported to contain phenolics¹⁵ and consequently exhibit a broad range of antibacterial properties.¹⁶ However because of poor solubility, low bioavailability and poor permeability, the delivery of herbal therapeutic molecules as drugs is challenging. These limitations can therefore be overcome by attaching or encapsulating the herbal drug with suitable nanomaterials; nutrient delivery systems through bioactive nanoencapsulation, which will further enhance the pharmacokinetics of the medicinal plant and improve their performance to a great extent.¹⁷⁻¹⁹ A nanoparticle (or nanopowder or nanocluster or nanocrystal) is a microscopic particle with at least one dimension less than 100nm.²⁰⁻²² The recent researches on the synthesis, characterization, and applications of nanoparticles are provided by metal nanoparticles,^{23,24} especially silver nanoparticles because they have unique features and have been shown to be non- or less toxic towards human when compared to other types of metal nanoparticles.^{25,26} Several methods of nanoparticle synthesis including physical, chemical and biological methods are either being developed or improved in order to enhance their properties and reduce the production costs.^{27,28} However, green synthesis; biological method of synthesizing nanoparticles which uses plants extract as reducing and capping agents are more advantageous than other methods of synthesis because it is a single-step method and it is human friendly.²⁹⁻³¹ Recently, as a further step towards the development of greener and more sustainable processes, attempts have been made to replace plant parts with agro industrial wastes.^{32,33} Biogenic silver NPs has been reported to have a wide spectrum of applications as an antimicrobial agent and for drug delivery.³⁴ In the present study, aqueous extracts of unripe pawpaw peels were used for a simple, low-cost and green method of AgNPs synthesis. The prepared nanoparticles were characterized primarily by UV-Visible spectroscopy and the

*Corresponding author. E mail: iyanu.adesipe@federalpolyilaro.edu.ng
Tel: +2347066025250

Citation: Adesipe TI and Adebayo AH. Comparative Antibacterial Activity of Biosynthesized Silver Nanoparticles and Aqueous Extract of Unripe Pawpaw Peels Against *E.coli*. Trop J Nat Prod Res. 2021; 5(2):359-363. doi.org/10.26538/tjnpr/v5i2.25

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

antibacterial activity against *E. coli* was compared to the aqueous extract of unripe pawpaw peels.

Materials and Methods

Unripe pawpaw was first obtained from local market in April 2018. All chemicals and reagent used were of analytical grade

Sample preparation

The peels were removed using table knife, washed thoroughly with distilled water and diced into tiny pieces.

Extract preparation

The aqueous extract of the peel was prepared according to the method of Abhay and Rupa³⁵ and was denoted as UPPA-unripe pawpaw peel aqueous extract. About 25 g of the unripe pawpaw peel was weighed into a beaker containing 100 mL distilled water and heated in a water bath at 60°C for 30min. The aqueous extracts were separated by filtration with Whatmann No. 1 filter paper (pore size 0.45 µm) and then centrifuged at 1000 rpm for 10 min.

Synthesis of silver nanoparticles

The modified method of Nooshinet *al.*,³⁶ was adopted for the green synthesis of silver nanoparticles from UPPA. In brief 10 ml of aqueous unripe peel extract was added into a flask containing 90 ml of 1 mM silver nitrate (AgNO₃) solution and was allowed to incubate for 24hrs at room temperature.

Characterization of silver nanoparticles

After overnight incubation at room temperature, the reduction of pure Ag⁺ ions was monitored by measuring the UV spectrum of the reaction medium within the range of 300 – 600 nm range.

Determination of total phenol content

Total Phenol Content was determined quantitatively using the Folin Ciocalteu reagent with Gallic acid as the standard.^{37,38} The phenolic content was calculated as Gallic acid equivalents GAE/g of samples on the basis of a standard curve of Gallic acid. All determinations were carried out in triplicate.

Comparative antibacterial activity of extract and biosynthesized AgNPs

Preparation of Samples for Antimicrobial Assay

The test organism employed in this study was obtained from the Molecular Biology Laboratory of Covenant University. UPPA (unripe pawpaw peel aqueous extract) and biosynthesized Silver nanoparticles (b-AgNPs) were oven-dried at 60 °C.¹⁸ The resultant dried extracts were dissolved separately in Phosphate Buffer Saline (PBS) to make a concentration of 4.7 g/mL solution each.

Antibacterial assay

The antibacterial potency of the samples were accessed using Agar well diffusion method as described by Abhay and Rupa,³⁵ with appropriate modifications. In brief, pre-sterilized Mueller Hinton agar was dispensed into petri dishes and allowed to solidify. After solidification of the agar plates, 20 µl of standardized inoculum of the test organism (*Escherichia coli*) was seeded on agar plate and wells of 5mm diameter were bored using a cork borer. About 50 µl of UPPA and b-AgNPs were loaded into the wells.

Plates were incubated at 37 °C for 24 hrs, and was inspected for Zone of inhibition. The zone of inhibition was compared with that formed by Ampicillin.

Statistical analysis

All experiments were done in triplicate and expressed as the mean± S.D. Graphical representation was done using Microsoft Excel 2007 package.

Results and Discussion

Addition of the UPPA to aqueous AgNO₃ solution resulted in a color change of the reaction mixture to colloidal brown indicating AgNP formation (b-AgNPs) as shown in Figure 1.

Previous studies have reported that AgNPs can be biosynthesized via peels such as Onion peels,^{39,40} Orange peels,³² Black pomengranate peels,⁴¹ Canvedish banana peel,⁴² unripe plantain peel,²⁹ and ripe pawpaw peels.⁴³ In this study, AgNPs was synthesized by reducing aqueous silver ions using UPPA after incubation at room temperature for 24 h in the dark. The colour of the reaction mixture turned brown and this color change has been previously reported by several researchers to indicate the formation of AgNPs.^{18,29,44,45,46} These authors mentioned that the color change is as a result of the surface plasmon resonance of precipitated AgNPs.

Characterization of Silver Nanoparticles

The ultraviolet-visible (UV-Vis) spectrum of AgNPs (Figure 2) was recorded from the reaction medium at 24 h interval. The AgNPs from the unripe pawpaw peel extracts gave absorbance peak of 450 nm. The peak due to silver ion at 300 nm was found missing.

The most useful characterization pertinent to the synthesis of silver nanoparticles has been established to be UV-visible spectroscopy.^{47,48} The biosynthesized AgNPs in this study was primarily characterized using UV-Vis spectrophotometer and an absorption peak of 450 nm was observed, which is typical of AgNPs as they usually display a surface Plasmon resonance band ranging from 400-550 nm.^{18,49,50,51} An absorption spectra of 450 nm has been reported for several biosynthesized AgNPs.^{44,52,53}

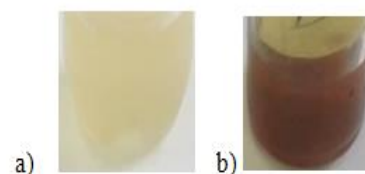


Figure 1: Unripe pawpaw peel aqueous extract before (a) and after (b) reaction with silver nitrate.

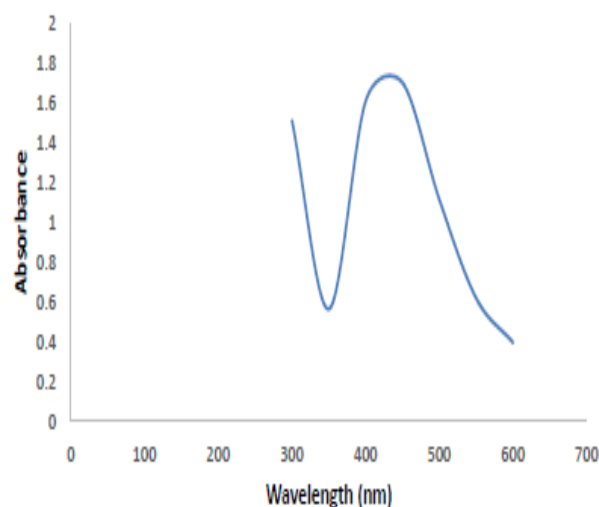


Figure 2: UV-Vis spectrum of AgNPs synthesized from aqueous extract of unripe pawpaw peels

Total phenolic content

The total phenolic contents of the unripe pawpaw peel extract (UPPA) and biosynthesized AgNPs (b-AgNPs) were expressed as mg of Gallic acid (GAE)/ g of extract as shown in table 1 below;

The result of total phenolic content (TPC) of peel extract alone or AgNPs showed that TPC was higher in the biosynthesized AgNPs (85.30 ± 0.57 mgGAE/g) compared to the aqueous peel extract alone (23.95 ± 0.57 mgGAE/g). Similar to this result, Sepidehet *et al.*,⁵⁴ Abdel-Aziz *et al.*,⁵⁵ and Sultana *et al.*,⁵⁶ reported a higher total phenol content in synthesized AgNPs compared to the *Prosopis farcta* fruit extract, *Chenopodium murale* and *Houttuyniacordata* leaf extracts respectively.

Antimicrobial Activities of Samples against *E.coli*

The antimicrobial activity of UPPA, b-AgNPs and standard antimicrobial drug, Ampicillin against pathogenic *E.coli* strain is shown in Figure 3 below;

The antimicrobial results revealed that AgNPs synthesized using aqueous extract of unripe pawpaw peel had a greater antibacterial activity with zone of inhibition of 36 mm against *E.coli* when compared with the peel extract alone with zone of inhibition of 27 mm. However, the standard antibacterial drug (Ampicillin) showed no zone of inhibition against the test pathogen. The result presented in this study validates that AgNPs have a broad antibacterial effect on antibiotic-resistant bacteria strains.⁵⁷ Contrary to this result, Muhammad *et al.*,⁵⁸ in his study reported that Callus extract of *F. indica* showed no antibacterial activity against *E.coli*, even though the Callus extract mediated AgNPs showed significant antibacterial activity against *E.coli* when compared to Ciprofloxacin. However, in this present work, the greater antibacterial activity exhibited by AgNPs synthesized using unripe pawpaw peel extract against *E.coli* could be as a result of capped phenolic compounds. Phenolic group has been implicated to enhance the conversion of silver nitrate to AgNPs due to its electron donating ability.^{54,59}

Table 1: Total phenolic content of extracts of unripe pawpaw peel and biosynthesized AgNPs

| Samples | Phenolic contents (mgGAE/g) |
|---------|-----------------------------|
| b-AgNPs | 85.30 ± 0.57 |
| UPPA | 23.95 ± 0.57 |

Values are expressed as mean \pm SD, n = 3

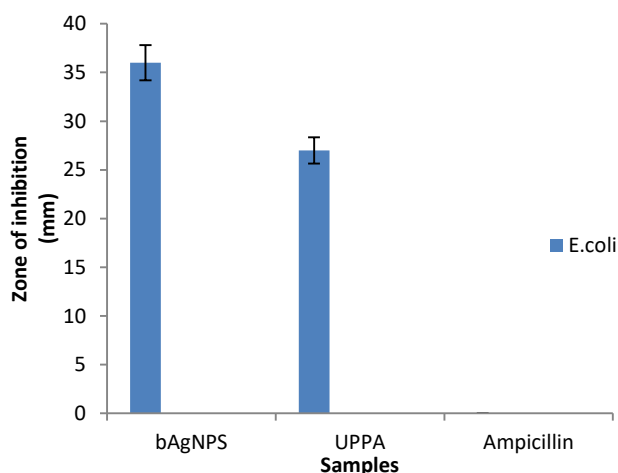


Figure 3:Antibacterial activity of b-AgNPs, UPPA and Ampicillin against *E.coli*

Conclusion

This study has revealed that silver nanoparticles can be synthesized in a simple and cost effective method using aqueous extract of unripe pawpaw peel. The total phenolic compound was higher in b-AgNPs compared to the pawpaw peel extract alone. Ampicillin showed no antibacterial activity against *E. coli* whereas, b-AgNPs showed a higher antibacterial activity against *E. coli* compared to the peel extract alone, indicating that the antimicrobial performance of aqueous extract of unripe pawpaw peels can be improved to a great extent if incorporated for the synthesis of AgNPs.

The results outlined in this study are novel, thus unripe pawpaw peel mediated AgNPs could emerge as an alternative to combat extraintestinal pathogenic *E. coli* resistant strains after proper pharmacologic evaluations.

Conflict of interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

References

- Ruta P, Frederic, S, Rita P, Vidmantas P, Thomas JW, Ruta P, Rasa S, Rasa B, Liliija K, Povilas K. Molecular characterization of uropathogenic *Escherichia coli* Reveals emergence of drug resistant O15, O22 and O25 serogroups. *Med.* 2019; 55:733.
- Jan TP and Michael W. Extraintestinal pathogenic *Escherichia coli*, a common human pathogen: challenges for vaccine development and progress in the field. *J Infect Dis.*2016; 213(1):6-13.
- Makrina TD, Gomes M, Adi I, Benjamin AR, Daniel JW, Minh-Duy P, David LP, Mark A S.Uropathogenic *Escherichia coli* mediated urinary tract infection. *Curr Drug Targets.* 2012;13(11):1386-1399.
- Mediavilla JR, Patrawalla A, Chen L. Colistin- and Carbapenem-Resistant *Escherichia coli* Harboring Mcr-1 and Blandm-5, causing a complicated urinary tract infection in a patient from the United States. *Mbio.* 2016; 7(4):01191-01196
- Kudinha T. The Pathogenesis of *Escherichia Coli* Urinary Tract Infection. In: Samie Amidou, editors. *Escherichia coli—Recent Advances on Physiology, Pathogenesis and Biotechnological Applications.* InTech. 2017. 45-61 p.
- Khameneh B, Iranshahy M, Soheili V, Sedigheh B, Bazzaz F. Review on plant antimicrobials: a mechanistic viewpoint. *Antimicrob. Resist Infect Control.*2019; 8:118.
- Rossiter SE, Fletcher MH, Wuest WM. Natural products as platforms to overcome antibiotic resistance. *Chem Rev.* 2017; 117(19):12415-12474.
- Ana CA, Andrew M, Manuel S. Plants as sources of new antimicrobials and resistance-modifying agents. *Nat Prod Rep.* 2012; 29(9):1007-1021.
- Takó M, Kerekes EB, Zambrano C, Kotogán A, Papp T, Krisch J, Vágvölgyi C. Plant Phenolics and Phenolic-Enriched Extracts as Antimicrobial Agents against Food-Contaminating Microorganisms. *Antioxidants (Basel, Switzerland).* 2020; 9(2):165.
- Lynda B, Valérian F, Pierre L, Yohann C, Lucie L, Nadia O, Pascal D, Claire B. Antibacterial properties of polyphenols: characterization and QSAR

- (Quantitative Structure–Activity Relationship). Models Front. Microbiol. 2019; 10(2019):829.
11. Ouerghemmi I, Iness BR, Fatma ZR, Soumaya B, Luisa P, Riadh K, Brahim M, Moufida ST. Antioxidant and antimicrobial phenolic compounds from extracts of cultivated and wild-grown Tunisian *Rutachalepensis*. J Food Drug Anal. 2017; 25(2):350-359.
 12. Rempe CS, Burris KP, Lenaghan SC, Stewart CN Jr. The Potential of Systems Biology to Discover Antibacterial Mechanisms of Plant Phenolics. FrontMicrobiol.2017; 8:422.
 13. Abreu AC, McBain AJ, Simões M. Plants as sources of new antimicrobials and resistance-modifying agents. Nat Prod Rep.2012; 29:1007-1021.
 14. Dada FA, Nzewuji FO, Esan AM, Oyeleye SI, Adegbola, V.B.Phytochemical and antioxidant analysis of aqueous extracts of unripe pawpaw (*Carica papaya* Linn.) fruit's peel and seed. IJRRAS. 2016; 27(3):68-71.
 15. Fasoyinu BT, Oyetayo OV, Ajayi AT. Antibacterial Activity of Fermenting Unripe Pawpaw Parts (*Carica papaya*) against Some Enteric Bacteria. Int J Pathogen Res. 2019; 3(2):1-10.
 16. Martínez-Ballesta M, Gil-Izquierdo Á, García-Viguera C, Domínguez-Perles R. Nanoparticles and Controlled Delivery for Bioactive Compounds: Outlining Challenges for New "Smart-Foods" for Health. Foods (Basel, Switzerland). 2018; 7(5):72.
 17. Gloria AO, Anthony JA, Emmanuel OA, Samuel WO. Characterization, antibacterial and antioxidant properties of silver nanoparticles synthesized from aqueous extracts of *Allium sativum*, *Zingiberofficinale*, and *Capsicum frutescens*. Pharmacogn. Mag 2017; 13(50):201-208.
 18. Okafor F, Janen A, Kukhtareva T, Edwards V, Curley M. Green synthesis of silver nanoparticles, their characterization, application and antibacterial activity. Int J Environ Res Public Health. 2013; 10:5221-5225.
 19. Muhammad N, Bilal H, AbbasiM, Younas WA, Taimoor K.A review of the green syntheses and antimicrobial applications of gold nanoparticles. Green Chem Lett Rev. 2017;10(4):216-227.
 20. Jaison J, Ahmed B, Yen, SC, Alain D, Michael KD. Review on nanoparticles and nanostructured materials: history, sources, toxicity and regulations. Beilstein J Nanotechnol. 2018; 9:1050-1074.
 21. Khan I, Saeed K, Khan I. Nanoparticles: Properties, applications and toxicities. Arab J Chem. 2019; 12(7):908-931
 22. Chen J, Qin G, Wang J, Yu J, Shen B, Li S, Ren Y, Zuo L, Shen W, Das B. One-step fabrication of sub-10-nm plasmonic nanogaps for reliable SERS sensing of microorganisms. Biosens Bioelectron.2013;44:191-197.
 23. Beyene HD, Werkneh AA, Bezabh HK, Ambaye TG. Synthesis paradigm and applications of silver nanoparticles, a review. Sustain Mater Technol.2017;13:18-23.
 24. Ratan ZA, Haidere MF, Nurunnabi M, Shahriar SM, Ahammad AS, Shim YY, Reaney MJ, Cho JY. Green Chemistry Synthesis of Silver Nanoparticles and Their Potential Anticancer Effects. Cancers. 2020; 12:855.
 25. Rai M, Yadav A, Gade A. Silver nanoparticles as a new generation of antimicrobials. Biotechnol Adv. 2009; 27(1):76-83.
 26. Ealia SM and Saravanakumar MP. A review on the classification, characterisation, synthesis of nanoparticles and their application. IOP Conf. Ser.: Mater Sci Eng.2017; 263:032019.
 27. Sivakumar SR, Sridhar V, Abdul K. antioxidant, antimicrobial and sewage treatment of synthesised silver nanoparticles from leaf extract of *Hygrophila auriculata (schumusch) heine*. IAJPS. 2017; 4(08):2350-2361.
 28. Olugbemi TI. Biosynthesis of Silver Nanoparticles from aqueous extract of unripe plantain peel and its antibacterial assay: A novel biological approach.IJPSH.2019; 9(5):9-16.
 29. Ana-Alexandra S, Alexandrina N, Rodica-mariana I, Ioana-Raluca S. Green synthesis of silver nanoparticles using plant extracts. Chem Sci. 2016; 1:188-193.
 30. Valli JS and Vaseeharan B. Biosynthesis of silver nanoparticles by *Cissusquadrangularis* extracts. Mater Lett.2012; 82:171-173.
 31. Skiba MI and Vorobyova VI. Synthesis of silver nanoparticles using orange peel extract prepared by plasmochemical extraction method and degradation of methylene blue under solar irradiation. Ann Mater Sci Eng. 2019; 115(2019):1-8.
 32. Borase HP, Salunke BK, Salunkhe RB. "Plant extract: a promising biomatrix for ecofriendly, controlled synthesis of Silver Nanoparticles. Appl BiochemBiotech.2014; 173(1):1-29.
 33. Janakiraman V, Govindarajan K, Cr M. Biosynthesis of silver nanoparticles from endophytic fungi, and its cytotoxic activity. BioNanoSci.2019; 9(3):573-579.
 34. Abhay T and Rupa S. Antimicrobial activities of silver nanoparticles synthesized from peel of fruits and vegetables. J Bio Innova Res and Develop Society.2016; 1:29-34.
 35. Nooshin A, Gholamreza A, Zahra JA. Green synthesis of silver nanoparticles using *Avena sativa* L. extract. Nanomed Res J. 2017; 2(1):57-63.
 36. Singleton VL and Rossi JA. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid Reagents. Am J EnolViticult. 1965;16:144-158.
 37. Chandra S, Khan S, Avula B, Lata H, Yang MH, ElSohly MA, Khan IA. Assessment of total phenolic and flavonoid content, antioxidant properties, and yield of aeroponically and conventionally grown leafy vegetables and fruit crops: a comparative study.Evid-Based ComplAltern Med. 2014; 2014:253875.
 38. Yu HY, Alyza AA, Nor KM, Fu Siong JY, Su-Yin K, Mohd ZA, Poh WC. Green Synthesis of Silver Nanoparticle Using Water Extract of Onion Peel and Application in the Acetylation Reaction. Arab J Sci Eng.2020;45:4797-4807.
 39. Olugbemi-Adesipe TI. Cost-effective green synthesis of silver nanoparticles from red onion peels aqueous and ethanol extracts and their antimicrobial activity, Proceedings of Nat Develop Conf of The School of Pure and ApplSci, The Federal Polytechnic Ilaro, Ogun State2019; 4:445-450.
 40. Sadeh K, Atefeh Z, Ali Z. Green synthesis of silver nanoparticles at low temperature in a fast pace with unique DPPH radical scavenging and selective cytotoxicity against MCF-7 and BT-20 tumor cell lines. Biotechnol Rep. 2019; 24:e00393.
 41. Kokila T, Ramesh PS, Geetha D. Biosynthesis of silver nanoparticles from Cavendish banana peel extract and its antibacterial and free radical scavenging assay: a novel biological approach. Appl Nanosci. 2015;5:911-920.
 42. Balavijayalakshmi J and Ramalakshmi V. *Carica papaya* peel mediated synthesis of silver nanoparticles and its antibacterial activity against human pathogens. J Appl Res Technol. 2017; 15(5):413-422.

43. Dada AO, Adekola FA, Dada FE, Adelani-Akande AT, Bello MO, Okonkwo CR, Inyinbor AA, Oluyori AP, Olayanju A, Ajanaku KO, Adetunji CO. Silver nanoparticle synthesis by *Acalypha wilkesiana* extract: phytochemical screening, characterization, influence of operational parameters, and preliminary antibacterial testing. *Heliyon*. 2019; 5:e02517.
44. Krithiga N, Rajalakshmi A, Jayachitra. Agreen synthesis of silver nanoparticles using leaf extracts of *Clitoria ternatea* and *Solanum nigrum* and study of its antibacterial effect against common nosocomial pathogens. *J Nanosci*.2015; 2015:8.
45. Banerjee P, Mantosh S, Aniruddha M, Papita D. Leaf extract mediated green synthesis of Silver Nanoparticles from widely available Indian Plants: synthesis, characterization, antimicrobial property and toxicity analysis. *Bioresour Bioprocess*.2014;1(3):1-10.
46. Dada AO, Adekola FA, Adeyemi OS, Bello MO, Adetunji CO, Awakan OJ, Grace FAA. Exploring the effect of operational factors and characterization imperative to the synthesis of silver nanoparticles. In: *Silver Nanoparticles Fabrication, Characterization and Applications*. London: IntechOpen; 2018. 165-184 p.
47. Xi-Feng Z, Zhi-Guo L, Wei S, Sangiliyandi G. Silver Nanoparticles: Synthesis, Characterization, Properties, Applications, and Therapeutic Approaches. *Int J Mol Sci*.2016; 17(9):1534.
48. Das G, Patra JK, Debnath T, Ansari A, Shin HS. Investigation of antioxidant, antibacterial, antidiabetic, and cytotoxicity potential of silver nanoparticles synthesized using the outer peel extract of *Ananas comosus* (L.). *PLoS ONE*. 2019;14(8):e0220950.
49. Mousavi BF and Zaker BS. Green synthesis of silver nanoparticles using *Artemisia turcomanica* leaf extract and the study of anti-cancer effect and apoptosis induction on gastric cancer cell line (AGS). *Artif Cells Nanomed Biotechnol*. 2018; 46(1):499-510.
50. Balashanmugam P and Kalaichelvan PT. Biosynthesis characterization of Silver Nanoparticles using *Cassia Roxburghii* DC. Aqueous Extract, and coated on Cotton Cloth for effective antibacterial activity. *Int J Nanomed*. 2015; 10:87-97.
51. Hina S, Juan D, Priyanka S, Tae HY. Ecofriendly synthesis of silver and gold nanoparticles by *Euphrasia officinalis* leaf extract and its biomedical applications. *Artif Cells Nanomed Biotechnol*. 2018; 46(6):1163-1170.
52. Ghodsieh B, Maryam MT, Mohmmad HN. Green synthesis of silver nanoparticles using aqueous extract of saffron (*Crocus sativus* L.) wastages and its antibacterial activity against six bacteria. *Asian Pac J Trop Biomed*. 2017;7(3):227-233.
53. Sepideh S, Sedigheh EB, Alireza S, Forough Y. *In-vitro* evaluation of antioxidant and antibacterial potential of green synthesized silver nanoparticles using *Prosopis farcta* fruit extract. *Iran J Pharm Res*.2019; 18(1):430-455.
54. Abdel-Aziz MS, Shaheen MS, El-Nekeety AA, Abdel-Wahhab MA. Antioxidant and antibacterial activity of silver nanoparticles biosynthesized using *chenopodium murale* leaf extract. *J Saudi ChemSoc*.2014;18(4):356-363.
55. Sultana F, Barman J, Mousmi SB. Biological approach to synthesis of silver nanoparticles using aqueous leaf extract of *Houttuynia cordata* Thunb and comparative antioxidant study of plant extract and synthesized nanoparticles. *J Mater Bio Appl*.2015;5:10-16.
56. Kim JS, Kuk E, Yu KN, Kim JH, Park SJ, Lee HJ, Kim SH, Park YK, Park YH, Hwang CY, Kim YK, Lee YS, Jeong DH, Cho MH. Antimicrobial effects of silver nanoparticles. *Nanomed*. 2007; 3(1):95-101.
57. Muhammad A, Tariq K, Muhammad A, Ayaz A.K, Muhammad A. Evaluation of the antibacterial potential of silver nanoparticles synthesized through the interaction of antibiotic and aqueous callus extract of *Fagonia indica*. *AMB Express*.2019; (9):75.
58. Philip D. *Mangifera indica* leaf-assisted biosynthesis of well-dispersed silver nanoparticles. *SpectrochimActa A Mol Biomol Spectrosc*. 2011; 78(1):327-331.