

GC-MS Analysis of Phytoconstituents in Fractions of *Corydalis adiantifolia*Iftikhar Ali^{1*}, Saleha S. Khan², Ajmal Khan³, Zulfiqar Ali¹, Huma A. Bhatti⁴, Jean J.K. Bankeu⁵, Viqar U. Ahmad⁴¹Department of Chemistry, Karakoram International University, 15100-Gilgit, Gilgit-Baltistan, Pakistan.²Department of Chemistry, Sardar Bahadur Khan Women University, Quetta, Balochistan, Pakistan.³UoN Chair of Oman's Medicinal Plants and Marine Natural Products, University of Nizwa, Nizwa, Sultanate of Oman.⁴HEJ Research Institute of Chemistry, University of Karachi, Karachi-75270, Pakistan.⁵Department of Chemistry, Faculty of Science, The University of Bamenda, P.O. Box 39, Bambili, Cameroon.

ARTICLE INFO

Article history:

Received 16 May 2018

Revised 03 June 2018

Accepted 05 June 2018

Published online 07 June 2018

Copyright: © 2018 Ali *et al.* This is an open-access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Corydalis adiantifolia Hook.f. & Thomson (Fumariaceae) is a herb known as shampoo (*Balti*) in Shigar valley, Baltistan. In addition, other *Corydalis* species have been reported as antidiabetic, anticancer, anti-inflammatory, and analgesic. Phytochemical study has exhibited the presence of mostly alkaloids in various *Corydalis* species. *C. adiantifolia*, to the best of our knowledge, has not been investigated so far for any kind of phytochemical analysis.

In our present study, the *n*-hexane, dichloromethane and ethyl acetate fractions of the methanol extract of *Corydalis adiantifolia* Hook.f. & Thomson (Fumariaceae) were studied through gas chromatography-mass spectrometry (GC-MS) analysis. A number of phytoconstituents are found in these fractions. The phytoconstituents reported in our present study include different esters, long chain alcohols, ketones, aldehydes and carboxylic acids, phenols, etc. More interestingly, different alkaloids are being reported in this study. Protopine, hydrastine, hydrastinine, oxyhydrostinine, (RS)-stylophine, oxoberberine, D-bicuculline and norsanguinarine are the main alkaloids reported through GC-MS in different fractions of *C. adiantifolia*.

Keywords: *Corydalis adiantifolia*, GC-MS Analysis, Fractions, Baltistan.

Introduction

Corydalis adiantifolia Hook.f. & Thomson (Fumariaceae) is a wild herb and its roots are reported for topical use as hair tonic.¹ In literature, *Corydalis* species and/or its active principles have been reported for a number of activities such as antithrombotic, anticoagulant,² hepatoprotective,³⁻⁴ neuroprotective,⁵ antitumor,⁶ anti-inflammatory,⁷ antinociceptive,⁸ analgesic,⁹ and antimalarial¹⁰ etc. Various compounds like phenylpropanoid amides,¹¹ lignanamides,⁶ flavonoid glycosides,¹² triterpenoids,¹³ have been isolated from different *Corydalis* species. However, alkaloids¹⁴⁻¹⁵ are the mostly reported compounds from a number of *Corydalis* species. Keeping in view the medicinal use of *C. adiantifolia* by the local community, it is our aim to investigate the plant species of *C. adiantifolia* for phytoconstituents. For this purpose, the fractions of the plant sample were studied through gas chromatography-mass spectrometry (GC-MS) method. Gas chromatography-mass spectrometry (GC-MS) technique has been utilized for the identification of phytoconstituents in certain plant species,¹⁶⁻¹⁸ and it is an important technique for this purpose. In our present study, the phytoconstituents found in *n*-hexane, dichloromethane and ethyl acetate fractions of *C. adiantifolia* have been sorted out.

*Corresponding author. E mail: iftikhar.ali@kiu.edu.pk
Tel: +92-3339665739

Citation: Ali I, Khan SS, Khan A, Ali Z, Bhatti HA, Bankeu JJK, Ahmad VU. GC-MS Analysis of Phytoconstituents in Fractions of *Corydalis adiantifolia*. Trop J Nat Prod Res. 2018; 2(6):282-289. doi.org/10.26538/tjnpr/v2i6.5

© 2018 Natural Product Research Group, Faculty of Pharmacy, University of Benin. All rights reserved.

Materials and Methods

Plant material

The plant material of *C. adiantifolia* (Fumariaceae) was collected from Shigar valley in August 2015 and was identified by Dr. Sher Wali Khan, Department of Biological Sciences, Karakoram International University (KIU), Gilgit. The voucher specimen (SKN-01) was deposited in the Department of Biological Sciences, KIU. The plant material was washed with tap water to remove the dust and other pollutants. Whole plant material was collected, dried in shade and crushed to powder using a grinder.

Extraction and fractionation

The crushed form of *C. adiantifolia* (1 kg) was soaked and extracted with 100% MeOH (3 L) for a week at room temperature. The methanol extract was evaporated to dryness using rotary evaporator at 40°C under reduced pressure.

The methanol extract (25 g) was further fractionated with *n*-hexane (CAH), dichloromethane (CAD), and ethyl acetate (CAE) and water residue was left behind. CAH, CAD and CAE were further analyzed through GC-MS for active principles.

Gas chromatography-mass spectrometry (GC-MS) analysis

The *n*-hexane (CAH), dichloromethane (CAD), and ethyl acetate (CAE) fractions were subjected to GC analysis and the GC-MS was done by means of 'Agilent GC-MS triple quad 7000 GC 7890A'. Helium was utilized as a carrier gas with a flow rate of 1.2 mL/min for all samples. The initial pressure was kept 9.1473 psi and run time 65 min for CAH and CAD and ZebronZB-5MS column was used for both samples at 40°C. For CAE the initial pressure was kept 9.7852 psi, the run time was kept 78 min at 40 °C and Agilent 19091J-433: 1825.64217 column was used. Analysis was performed in split mode in front SS inlet He. A split flow of 12 mL/min was kept for CAH and CAD while 36 mL/min split flow was maintained for CAE. The injection volume for CAH and CAD was kept 2 µL each while it was 1.5 µL for CAE. The oven program for CAH and CAD was kept on at 40°C for 10 min, then

10°C/min to 190°C for 15 min and then 10°C/min to 290°C for 20 min. While the oven program for CAE was kept on at 50°C for 10 min, then 10°C/min to 180°C for 20 min and then 10°C/min to 280°C for 25 min. Electron impact (EI) was the ion source and triple quadrupole detector was used.

The identification of compounds of each fraction was based on the computer evaluation of mass spectra through NIST based AMDIS V 2.69 (Automated mass spectral deconvolution and identification software), direct comparison of peaks and retention time with those for standard compounds, with eight peak index.¹⁹ A mass spectral survey was performed using NIST library for spectral comparison and identification.

Results and Discussion

The results of GC-MS analysis of n-hexane (CAH), dichloromethane (CAD) and ethyl acetate (CAE) fractions of *C. adiantifolia* are presented in Table 1, Table 2 and Table 3, respectively. Each table contains the data on retention time (min), name of compound, molecular formula, molecular weight and concentration i.e. peak area (%) for every individual compound identified through the technique. The GC-MS analysis of CAH, CAD and CAE revealed the presence of 24, 10 and 31 compounds, respectively.

The major compounds found in CAH include phthalic acid, mono(2-ethylhexyl) ester; 10-nonadecanol; hexadecanoic acid, methyl ester; β -Sitosterol and 9,12-Octadecadienoic acid (Z,Z)-methyl ester, etc.

The mentionable phytoconstituents found in CAD were protopine; benzo[f]quinolin-3(2H)-one, 1,4-dihydro-1-(2,5-dimethoxyphenyl)-; isobenzofuran-1,3-dione, 4,5-dimethoxy-; hydrastine; oxyhydrastinine and (RS)-stylophine, etc. However, the ethyl acetate fraction (CAE) was found to contain more of the compounds. Among those compounds benzo[f]quinolin-3(2H)-one, 1,4-dihydro-1-(2,5-dimethoxyphenyl)-; protopine; n-hexadecanoic acid; 3,4-methylenedioxypropiofenone; 1(3H)-isobenzofuranone, 6,7-dimethoxy; norsanguinarine; γ -sitosterol; D-bicuculline; hydrastine; hydrastinine and oxoberberine.

The literature survey of some of the chemical principles found in the fractions of *C. adiantifolia* mentioned above reveals the medicinal value of the plant species. Protopine; an alkaloid, has been reported for its anticonvulsant activity,²⁰ microtubule-stabilizing effects,²¹ anti-inflammatory activity reducing effects,²² neuroprotective effects,²³⁻²⁴ anticholinesterase effects,²⁵ etc. Sanguinarine compounds have been reported for anticancer,²⁶⁻²⁷ anti-osteoporosis,²⁸⁻²⁹ and norsanguinarine has been reported for antifungal activity.³⁰ Hydrastine has been reported for the effects on dopamine biosynthesis,³¹ PAK4 kinase inhibition³² etc. Oxoberberine has exhibited effects on sodium current in human atrial myocytes.³³

The GC-MS chromatograms of CAH, CAD and CAE are given in Figure 1, Figure 2 and Figure 3, respectively.

The mass spectra of the identified compounds reported in CAH, CAD and CAE fractions of *C. adiantifolia* through GC-MS analysis along with their structures are provided as Figure 4, Figure 5 and Figure 6, respectively.

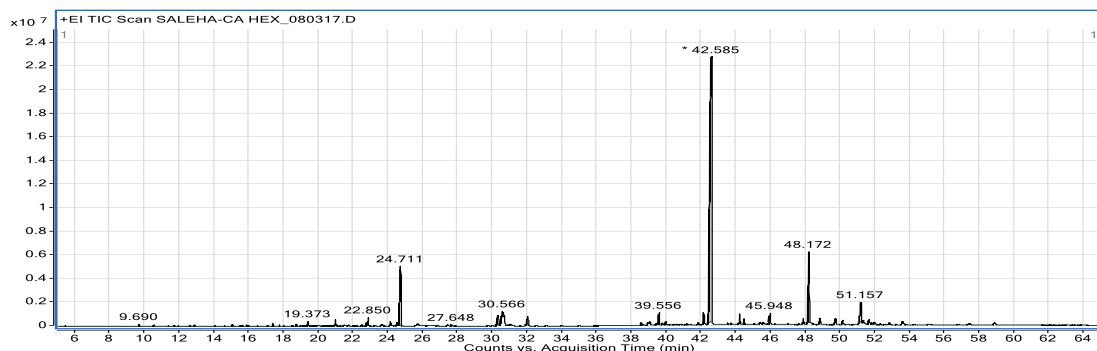


Figure 1: GC-MS chromatogram for n-hexane fraction of *C. adiantifolia*

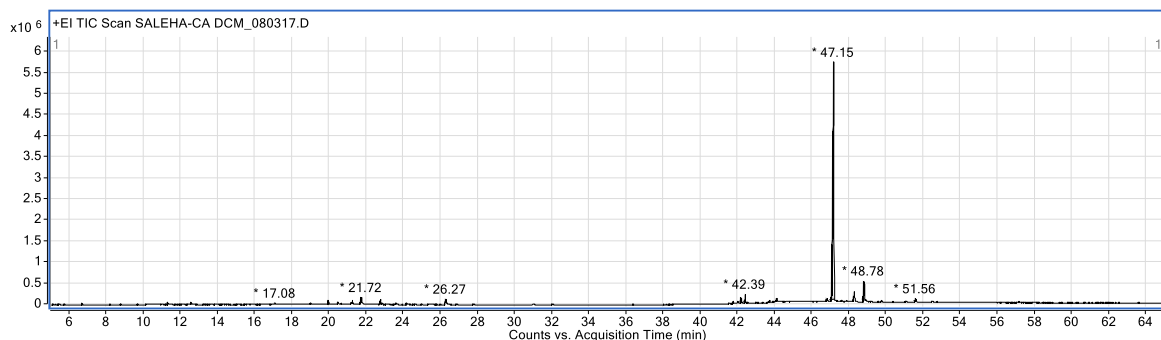


Figure 2: GC-MS chromatogram for dichloromethane fraction of *C. adiantifolia*

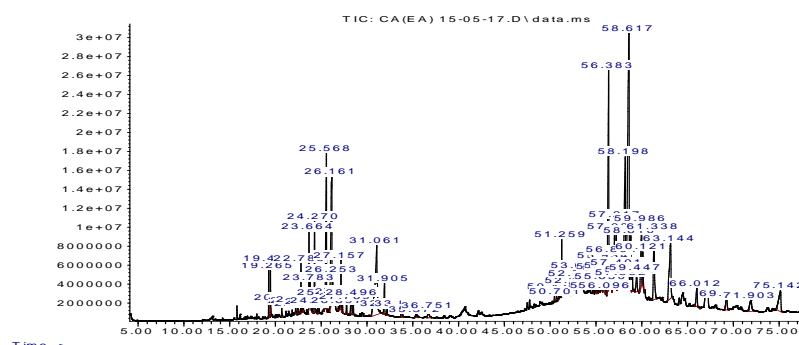


Figure 3: GC-MS chromatogram for ethyl acetate fraction of *C. adiantifolia*

Table 1: GC-MS spectral analysis of n-hexane fraction of *C. adiantifolia*.

Peak No.	RT (min)	Name of compound	Molecular formula	Molecular weight	Peak area (%)
1	9.70	2-Heptenal, (Z)-	C ₇ H ₁₂ O	112	0.10
2	17.38	Nonanoic acid, 9-oxo-, methyl ester	C ₁₀ H ₁₈ O ₃	186	0.12
3	19.38	Methyl 10-oxo-8-decenoate	C ₁₁ H ₁₈ O ₃	198	0.19
4	20.96	Methyl tetradecanoate	C ₁₅ H ₃₀ O ₂	242	0.30
5	22.85	2-Pentadecanone, 6,10,14-trimethyl-	C ₁₈ H ₃₆ O	268	0.17
6	23.64	1-Hexadecanol	C ₁₆ H ₃₄ O	242	0.19
7	24.13	7-Hexadecenoic acid, methyl ester, (Z)-	C ₁₇ H ₃₂ O ₂	268	0.36
8	24.53	7-Hexadecenoic acid, methyl ester, (Z)-	C ₁₇ H ₃₂ O ₂	268	0.35
9	24.68	Hexadecanoic acid, methyl ester	C ₁₇ H ₃₄ O ₂	270	6.60
10	27.65	Heptadecanoic acid, methyl ester	C ₁₈ H ₃₆ O ₂	284	0.12
11	30.31	9,12-Octadecadienoic acid (Z,Z)-, methyl ester	C ₁₉ H ₃₄ O ₂	294	1.84
12	30.55	9,12,15-Octadecatrienoic acid, methyl ester, (Z,Z,Z)-	C ₁₉ H ₃₂ O ₂	292	1.23
13	31.99	Octadecanoic acid, methyl ester	C ₁₉ H ₃₈ O ₂	298	1.64
14	38.55	1,2-15,16-Diepoxylhexadecane	C ₁₆ H ₃₀ O ₂	254	0.23
15	39.56	Eicosanoic acid, methyl ester	C ₂₁ H ₄₂ O ₂	326	0.78
16	39.96	Z-(13,14-Epoxy)tetradec-11-en-1-ol acetate	C ₁₆ H ₂₈ O ₃	268	0.23
17	42.13	Heneicosane	C ₂₁ H ₄₄	296	0.64
18	42.58	Phthalic acid, mono(2-ethylhexyl) ester	C ₁₆ H ₂₂ O ₄	278	72.68
19	44.19	Heptacosane	C ₂₇ H ₅₆	380	0.55
20	44.44	Tetracosanoic acid, methyl ester	C ₂₅ H ₅₀ O ₂	382	0.24
21	47.85	10-Nonadecanone	C ₁₉ H ₃₈ O	282	0.16
22	48.16	10-Nonadecanol	C ₁₉ H ₄₀ O	284	8.12
23	48.81	Benzo[f]quinolin-3(2H)-one, 1,4-dihydro-1-(2,5-dimethoxyphenyl)-	C ₂₁ H ₁₉ NO ₃	333	0.34
24	51.14	β-Sitosterol	C ₂₉ H ₅₀ O	414	2.82

Table 2: GC-MS spectral analysis of dichloromethane fraction of *C. adiantifolia*

Peak No.	RT (min)	Name of compound	Molecular formula	Molecular weight	Peak area (%)
1	19.95	1,3-Dioxolo[4,5-g]isoquinolin-5-ol, 5,6,7,8-tetrahydro-6-methyl-	C ₁₁ H ₁₃ NO ₃	207	0.42
2	21.73	Isobenzofuran-1,3-dione, 4,5-dimethoxy-	C ₁₀ H ₈ O ₅	208	2.15
3	26.27	Oxyhydrostinine	C ₁₁ H ₁₁ NO ₃	205	1.30
4	42.17	Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester	C ₁₉ H ₃₈ O ₄	330	0.80
5	42.38	Phthalic acid, diisooctyl ester	C ₂₄ H ₃₈ O ₄	390	1.28
6	46.79	(RS)-Stylopine	C ₁₉ H ₁₇ NO ₄	323	0.48
7	47.15	Protopine	C ₂₀ H ₁₉ NO ₅	353	85.63
8	48.25	Hydrastine	C ₂₁ H ₂₁ NO ₆	383	2.14
9	48.77	Benzo[f]quinolin-3(2H)-one, 1,4-dihydro-1-(2,5-dimethoxyphenyl)-	C ₂₁ H ₁₉ NO ₃	333	4.69
10	51.57	Spiro[androstande-3,2'-thiazolidine], (5α)-	C ₂₁ H ₃₅ NS	333	0.93

Table 3: GC-MS spectral analysis of ethyl acetate fraction of *C. adiantifolia*.

Peak No.	RT (min)	Name of compound	Molecular formula	Molecular weight	Peak area (%)
1	19.26	4-Methoxymethylphenol	C ₈ H ₁₀ O ₂	138	0.73
2	19.45	2-Methoxy-4-vinylphenol	C ₉ H ₁₀ O ₂	150	0.74
3	20.69	Vanillin lactoside	C ₂₀ H ₂₈ O ₁₃	476	0.29
	22.22	Cinnamic acid, 4-hydroxy-3-methoxy-, [5-hydroxy-2-hydroxymethyl-6-[2-(4-hydroxy-3-methoxyphenyl)ethoxy]-4-(6-methyl-3,4,5-trihydroxytetrahydropyran-2-yl)oxy]tetrahydropyran-3-yl ester	C ₃₁ H ₄₀ O ₁₅	652	
4					0.18
5	22.45	Octahydrobenzo[b]pyran, 4a-acetoxy-5,5,8a-trimethyl-	C ₁₄ H ₂₄ O ₃	240	0.11
6	22.79	3',5'-Dimethoxyacetophenone	C ₁₀ H ₁₂ O ₃	180	0.79
7	23.06	9-Hexadecenoic acid	C ₁₆ H ₃₀ O ₂	254	0.09
8	23.66	Hydrastinine	C ₁₁ H ₁₃ NO ₃	207	1.59
	24.04	1b,4a-Epoxy-2H-cyclopenta[3,4]cyclopropa[8,9]cycloundec[1,2-b]oxiren-5(6H)-one, 7-(acetyloxy)decahydro-2,9,10-trihydroxy-3,6,8,8,10a-pentamethyl-	C ₂₂ H ₃₂ O ₈	424	
9					0.16
10	24.26	Ethenylcyanide, 3-[3,4-methylenedioxyphenyl]-	C ₁₀ H ₇ NO ₂	173	2.92
11	25.52	1(3H)-Isobenzofuranone, 6,7-dimethoxy-	C ₁₀ H ₁₀ O ₄	194	7.66
12	26.06	3,4-Methylenedioxypropiofenone	C ₁₀ H ₁₀ O ₃	178	9.33
	26.89	3,3a-Epoxydicyclopenta[a,d]cyclooctan-4β-ol, 9,10a-dimethyl-6-methylene-3β-isopropyl-	C ₂₀ H ₃₂ O ₂	304	
13					0.17
14	27.16	3,7,11,15-Tetramethyl-2-hexadecen-1-ol	C ₂₀ H ₄₀ O	296	1.33
15	27.31	2-Propenoic acid, 3-(4-hydroxy-3-methoxyphenyl)-, methyl ester	C ₁₁ H ₁₂ O ₄	208	0.42
16	27.76	Ethanol, 2-(9-octadecenyl)-, (Z)-	C ₂₀ H ₄₀ O ₂	312	0.38
17	28.49	2-Isobenzazol, 1,3-dioxo-2-methyl-4,5-methylenedioxy-	C ₁₀ H ₇ NO ₄	205	0.54
18	31.04	n-Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	256	10.27
	51.25	3',8,8'-Trimethoxy-3-piperidyl-2,2'-binaphthalene-1,1',4,4'-tetrone	C ₂₈ H ₂₅ NO ₇	487	
19					1.02
20	52.39	Ethyl iso-allocholate	C ₂₆ H ₄₄ O ₅	436	0.16
21	56.38	Protopine	C ₂₀ H ₁₉ NO ₅	353	12.15
	56.89	2-[4-methyl-6-(2,6,6-trimethylcyclohex-1-enyl)hexa-1,3,5-trienyl]cyclohex-1-en-1-carboxaldehyde	C ₂₃ H ₃₂ O	324	
22					0.59
23	57.03	Hydrastine	C ₂₁ H ₂₁ NO ₆	383	1.66
24	57.22	Stigmastan-3,5-diene	C ₂₉ H ₄₈	396	2.21
25	57.39	9,12-Octadecadienoic acid, 2-phenyl-1,3-dioxan-5-yl ester, cis-	C ₂₈ H ₄₂ O ₄	442	0.67
	58.61	Benzo[f]quinolin-3(2H)-one, 1,4-dihydro-1-(2,5-dimethoxyphenyl)-	C ₂₁ H ₁₉ NO ₃	333	
26					23.69
27	59.92	D-Bicuculline	C ₂₀ H ₁₇ NO ₆	367	4.12
28	61.36	γ-Sitosterol	C ₂₉ H ₅₀ O	414	4.99
29	63.16	Norsanguinarine	C ₁₉ H ₁₁ NO ₄	317	6.39
30	66.02	Oxoberberine	C ₂₁ H ₂₁ NO ₄	351	1.47
	75.05	7-Azadibenz[a,e]azulen-12-one, 5,6,7,7a,12,12a-hexahydro-7-methyl-8,9-bis(methoxy)-2,3-methylenedioxy-12a-acetyloxy-	C ₂₃ H ₂₃ NO ₇	425	
31					3.16

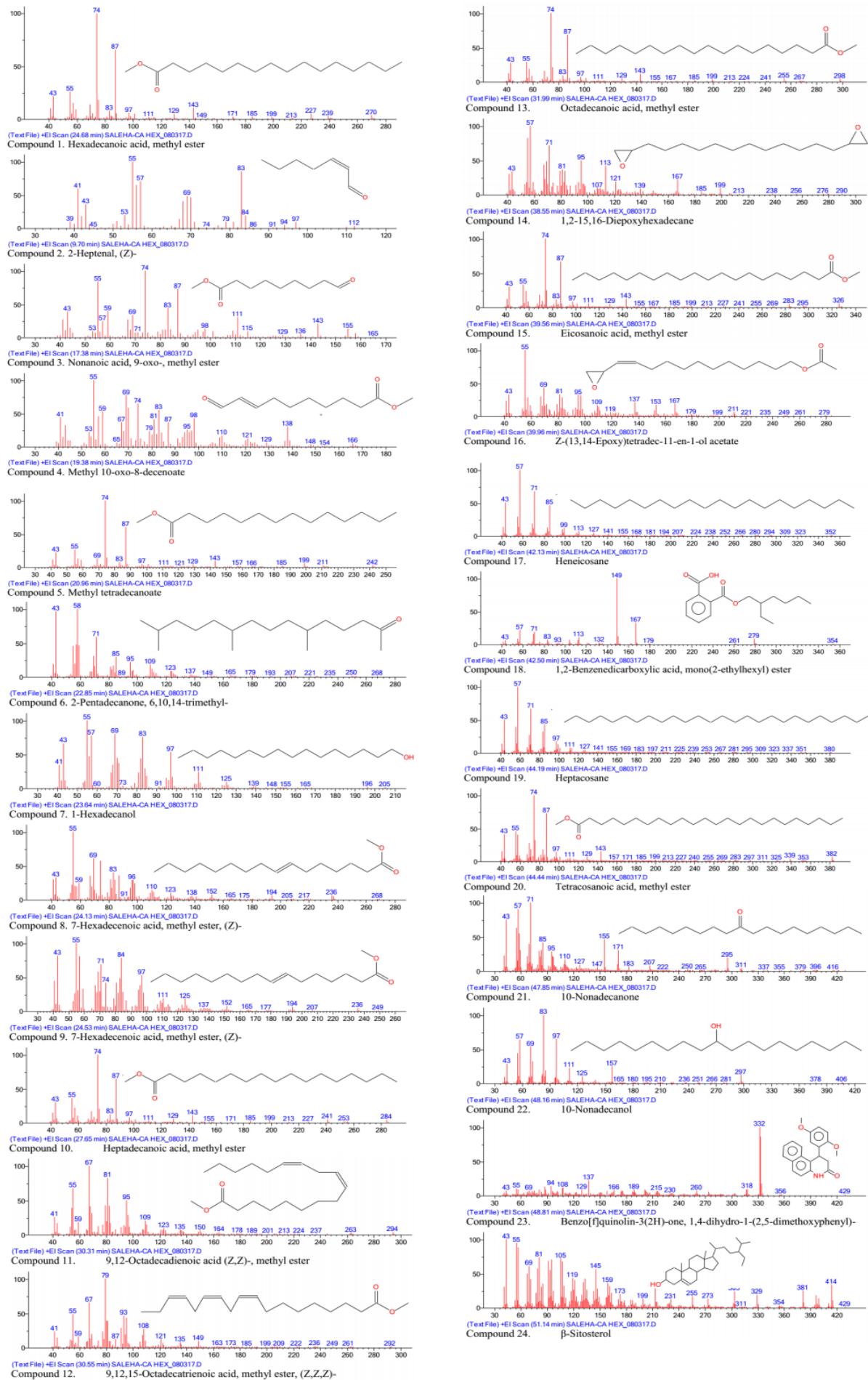


Figure 4: Mass spectra of identified compounds from n-hexane fraction of *C. adiantifolia*.

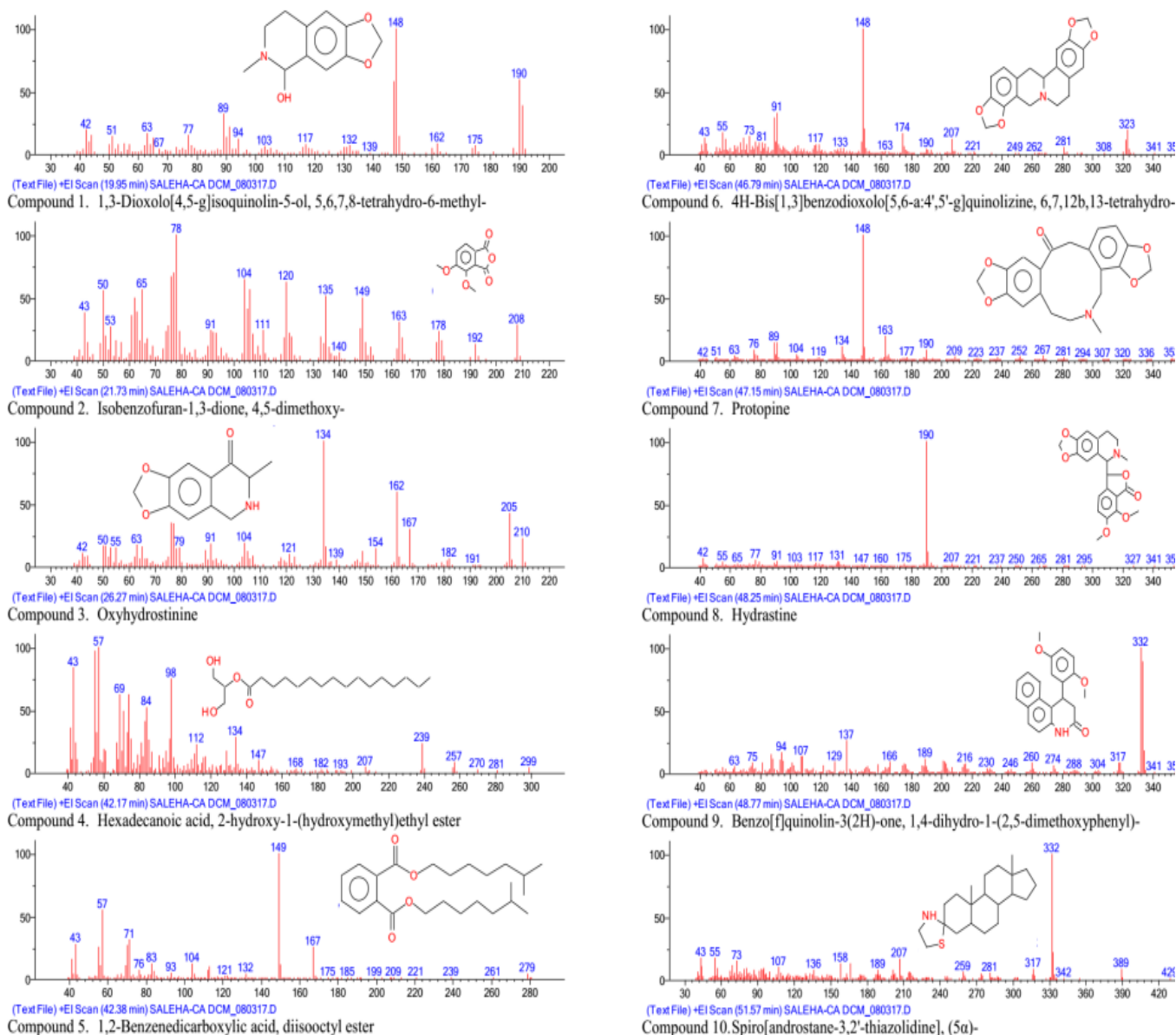


Figure 5: Mass spectra of identified compounds from DCM fraction of *C. adiantifolia*

Conclusion

The n-hexane (CAH), dichloromethane (CAD) and ethyl acetate (CAE) fractions of *Corydalis adiantifolia* extracted in methanol were separately analyzed using gas chromatography-mass spectrometry (GC-MS) technique. Total number of compounds found in CAH, CAD and CAE were 24, 10 and 31, respectively. Mostly esters were reported in CAH fraction of the sample and certain important alkaloids such as protopine, hydrastine, oxyhydrostinine, (RS)-stylopinine were found in CAD fraction of *C. adiantifolia*. While, the protopine, hydrastine, hydrastinine, oxoberberine, D-bicuculline and norsanguinarine were reported in ethyl acetate fraction of the plant.

Conflict of interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

Acknowledgments

The authors thank Dr. Sher Wali Khan, Associate Professor, Department of Biological Sciences, Karakoram International University Gilgit for the identification of the plant sample. The authors wish to acknowledge Higher Education Commission of Pakistan for the financial support through National Support Program for Universities (Project No. NRP-3466).

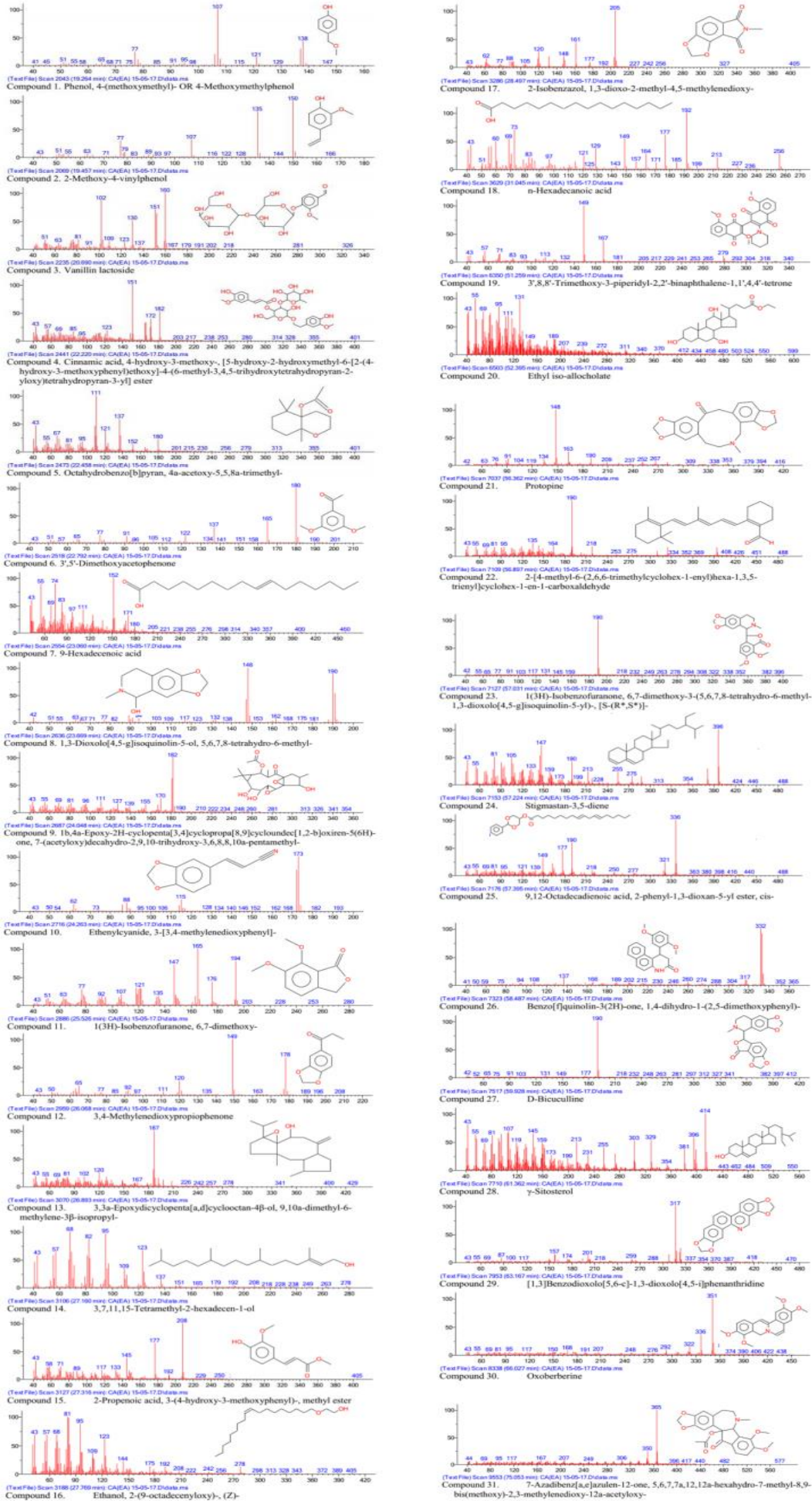


Figure 6: Mass spectra of identified compounds from Ethyl acetate fraction of *C. adiantifolia*.

References

- Abbas Z, Khan SM, Alam J, Khan SW, Abbasi AM. Medicinal plants used by inhabitants of the Shigar Valley, Baltistan region of Karakorum range-Pakistan. *J Ethnobiol Ethnomed.* 2017; 13(1):53.
- Chang S, Yang Z, Han N, Liu Z, Yin J. The antithrombotic, anticoagulant activity and toxicity research of ambinine, an alkaloid from the tuber of *Corydalis ambigua* var. *amurensis*. *Regul Toxicol Pharmacol.* 2018; 95:175-181.
- Wu F, Zheng H, Yang ZT, Cheng B, Wu JX, Liu XW, Tang CL, Lu SY, Chen ZN, Song FM, Ruan JX, Zhang HY, Liang YH, Song H, Su ZH. Urinary metabonomics study of the hepatoprotective effects of total alkaloids from *Corydalis saxicola* Bunting on carbon tetrachloride-induced chronic hepatotoxicity in rats using (1)H NMR analysis. *J Pharm Biomed Anal.* 2017; 140:199-209.
- Liang YH, Tang CL, Lu SY, Cheng B, Wu F, Chen ZN, Song F, Ruan JX, Zhang HY, Song H, Zheng H, Su ZH. Serum metabonomics study of the hepatoprotective effect of *Corydalis saxicola* Bunting on carbon tetrachloride-induced acute hepatotoxicity in rats by (1)H NMR analysis. *J Pharm Biomed Anal.* 2016; 129:70-79.
- Yang JL, Ha TKQ, Lee BW, Kim J, Oh WK. PTP1B inhibitors from the seeds of *Iris sanguinea* and their insulin mimetic activities via AMPK and ACC phosphorylation. *Bioorg Med Chem Lett.* 2017; 27(22):5076-5081.
- Zhang B, Huang R, Hua J, Liang H, Pan Y, Dai L, Liang D, Wang H. Antitumor lignanamides from the aerial parts of *Corydalis saxicola*. *Phytomed.* 2016; 23(13):1599-1609.
- Yang C, Zhang C, Wang Z, Tang Z, Kuang H, Kong AN. Corynoline Isolated from *Corydalis bungeana* Turcz. Exhibits Anti-Inflammatory Effects via Modulation of Nfr2 and MAPKs. *Molecules.* 2016; 21(8):E975.
- Wang L, Zhang Y, Wang Z, Gong N, Kweon TD, Vo B, Wang C, Zhang X, Chung JY, Alachkar A, Liang X, Luo DZ, Civelli O. The Antinociceptive Properties of the *Corydalis yanhusuo* Extract. *PLoS One.* 2016; 11(9):e0162875.
- Muhammad N, Shrestha RL, Adhikari A, Wadood A, Khan H, Khan AZ, Maione F, Mascolo N, De Feo V. First evidence of the analgesic activity of govaniadine, an alkaloid isolated from *Corydalis govaniiana* Wall. *Nat Prod Res.* 2015; 29(5):430-437.
- Wangchuk P, Keller PA, Pyne SG, Willis AC, Kamchonwongpaisan S. Antimalarial alkaloids from a Bhutanese traditional medicinal plant *Corydalis dubia*. *J Ethnopharmacol.* 2012; 143(1):310-313.
- Peng ZT, Chao LH, Huo HX, Chen XN, Yao HN, Zhang Y, Zhao YF, Tu PF, Zheng J, Li J. [Phenylpropanoid amides from whole plants of *Corydalis edulis*]. *Zhongguo Zhong Yao Za Zhi.* 2018; 43(1):109-113.
- Xie C, Veitch NC, Houghton PJ, Simmonds MS. Flavonoid glycosides and isoquinolinone alkaloids from *Corydalis bungeana*. *Phytochemistry.* 2004; 65(22):3041-3047.
- Kim KH, Lee IK, Choi SU, Lee JH, Moon E, Kim SY, Lee KR. New triterpenoids from the tubers of *Corydalis ternata*: structural elucidation and bioactivity evaluation. *Planta Med.* 2011; 77(13):1555-1558.
- Zhang J, Zhang QY, Tu PF, Xu FC, Liang H. Mucroniferanines A-G, Isoquinoline Alkaloids from *Corydalis mucronifera*. *J Nat Prod.* 2018; 81(2):364-370.
- Naseri M, Emami SA, Asili J, Tayarani-Najaran Z, Dehghan G, Schneider B, Iranshahi M. Rupestrines A-D, alkaloids from the aerial parts of *Corydalis rupestris*. *Bioorg Chem.* 2018; 77:651-659.
- Zeb A, Ullah F, Ayaz M, Ahmad S, Sadiq A. Demonstration of biological activities of extracts from *Isodon rugosus* Wall. Ex Benth: Separation and identification of bioactive phytoconstituents by GC-MS analysis in the ethyl acetate extract. *BMC Complement Altern Med.* 2017; 17(1):284.
- Sardar AA, Perveen A, Khan ZU, Farid S and Khan I. Phytochemical screening, GC-MS analysis and in vitro antioxidant activity of pollen of *Centella asiatica* (Linn) urban a traditional medicinal plant. *Pak J Pharm Sci.* 2017; 30(6):2239-2245.
- Chander MP, Vinod Kumar K, Lall C, Vimal Raj R, Vijayachari P. GC/MS profiling, in vitro anti-leptospiral and haemolytic activities of *Boesenbergia rotunda* (L.) Mansf. used as a medicinal plant by Nicobarese of Andaman and Nicobar Islands. *Nat Prod Res.* 2016; 30(10):1190-1192.
- Center MSD. Eight peak index of mass Spectra. Mass Spectrometry Data Centre, AWRE, Aldermaston, Reading. 1974.
- Prokopenko Y, Tsyvunin V, Shtrygol S, Georgiyants V. *In Vivo* Anticonvulsant Activity of Extracts and Protopine from the *Fumaria schleicheri* Herb. *Sci Pharm.* 2015; 84(3):547-554.
- Chen CH, Liao CH, Chang YL, Guh JH, Pan SL, Teng CM. Protopine, a novel microtubule-stabilizing agent, causes mitotic arrest and apoptotic cell death in human hormone-refractory prostate cancer cell lines. *Cancer Lett.* 2012; 315(1):1-11.
- Bae DS, Kim YH, Pan CH, Nho CW, Samdan J, Yansan J, Lee JK. Protopine reduces the inflammatory activity of lipopolysaccharide-stimulated murine macrophages. *BMB Rep.* 2012; 45(2):108-113.
- Xiao X, Liu J, Hu J, Zhu X, Yang H, Wang C, Zhang Y. Protective effects of protopine on hydrogen peroxide-induced oxidative injury of PC12 cells via Ca(2+) antagonism and antioxidant mechanisms. *Eur J Pharmacol.* 2008; 591(1-3):21-27.
- Xiao X, Liu J, Hu J, Li T, Zhang Y. Protective effect of protopine on the focal cerebral ischaemic injury in rats. *Basic Clin Pharmacol Toxicol.* 2007; 101(2):85-89.
- Kim SR, Hwang SY, Jang YP, Park MJ, Markelonis GJ, Oh TH, Kim YC. Protopine from *Corydalis ternata* has anticholinesterase and anti-amnesic activities. *Planta Med.* 1999; 65(3):218-221.
- Zhang S, Leng T, Zhang Q, Zhao Q, Nie X, Yang L. Sanguinarine inhibits epithelial ovarian cancer development via regulating long non-coding RNA CASC2-EIF4A3 axis and/or inhibiting NF-kappaB signaling or PI3K/AKT/mTOR pathway. *Biomed Pharmacother.* 2018; 102:302-308.
- Ma Y, Yu W, Shrivastava A, Alemi F, Lankachandra K, Srivastava RK, Shankar S. Sanguinarine inhibits pancreatic cancer stem cell characteristics by inducing oxidative stress and suppressing sonic hedgehog-Gli-Nanog pathway. *Carcinogenesis.* 2017; 38(10):1047-1056.
- Zhang F, Xie J, Wang G, Zhang G, Yang H. Anti-osteoporosis activity of Sanguinarine in preosteoblast MC3T3-E1 cells and an ovariectomized rat model. *J Cell Physiol.* 2018; 233(6):4626-4633.
- Ma Y, Sun X, Huang K, Shen S, Lin X, Xie Z, Wang J, Fan S, Ma J, Zhao X. Sanguinarine protects against osteoarthritis by suppressing the expression of catabolic proteases. *Oncotarget.* 2017; 8(38):62900-62913.
- Singh S, Jain L, Pandey MB, Singh UP, Pandey VB. Antifungal activity of the alkaloids from *Eschscholzia californica*. *Folia Microbiol (Praha).* 2009; 54(3):204-206.
- Kim SH, Shin JS, Lee JJ, Yin SY, Kai M, Lee MK. Effects of hydrastine derivatives on dopamine biosynthesis in PC12 cells. *Planta Med.* 2001; 67(7):609-613.
- Guo B, Li X, Song S, Chen M, Cheng M, Zhao D, Li F. (-)-beta-hydrastine suppresses the proliferation and invasion of human lung adenocarcinoma cells by inhibiting PAK4 kinase activity. *Oncol Rep.* 2016; 35(4):2246-2256.
- Chi JF, Chu SH, Lee CS, Su MJ. Effects of 8-oxoberberine on sodium current in rat ventricular and human atrial myocytes. *Can J Cardiol.* 1997; 13(11):1103-1110.