Tropical Journal of Natural Product Research

Available online at <u>https://www.tjnpr.org</u> Original Research Article



Enhancing Chemotherapeutic Efficacy: Synergistic Cytotoxic Effect of *Garcinia cowa* Bark Extract and Doxorubicin in T47D Breast Cancer Cells

Ifora Ifora^{1,3}, Dachriyanus Hamidi², Meri Susanti², Dira Hefni², Fatma S. Wahyuni^{2*}

¹Doctoral Program, Faculty of Pharmacy, Andalas University, Padang, West Sumatra 25163, Indonesia

²Faculty of Pharmacy, Andalas University, Kampus Limau Manis, Padang, West Sumatra 25163, Indonesia

³Departement of Pharmacology and Clinical Pharmacy, School of Pharmaceutical Science Padang (STIFARM Padang), West Sumatera 25147, Indonesia.

ARTICLE INFO

ABSTRACT

Article history: Received 22 October 2024 Revised 23 November 2024 Accepted 10 December 2024 Published online 01 February 2025

Copyright: © 2025 Ifora *et al.* This is an open-access article distributed under the terms of the <u>Creative</u> <u>Commons</u> Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Breast cancer remains one of the leading causes of mortality worldwide, demanding more effective and less toxic treatment strategies. The investigation of natural compounds as potential adjuvants to chemotherapy has gained urgency in recent years. The aim of this research was to investigate the cytotoxic effects of Garcinia cowa bark ethanol extract (GCBEE) alone and its potential to enhance the cytotoxic activity of doxorubicin on T47D breast cancer cells, as well as to assess the selectivity of GCBEE towards the Vero cell line. The MTT assay was utilized to evaluate the cytotoxic effects of doxorubicin (dox) and GCBEE on T47D breast cancer cells. Selectivity against normal Vero cell lines was evaluated using the selectivity index (SI). The test results were examined using a microplate reader set to operate at 595 nm wavelength. Chou Talalay's method was utilized to compute and analyze the combination index (CI) in order to ascertain the influence of the combination. Based on the 3-(4,5-dimethylthiazol-2-yl)-2,5diphenyltetrazolium bromide (MTT) assay, both GCBEE and dox exhibited concentrationdependent cytotoxic effects with IC50 values of 130 µg/mL and 0.026 µg/mL on T47D cells, respectively. GCBEE was more selective for T47D cells than Vero cells (SI = 15). GCBEE synergistically enhanced the cytotoxic effect of dox, with a CI value range of 0.19-0.89. The combination of GCBEE and doxorubicin offers a promising synergistic approach in the fight against breast cancer.

Keywords: Cytotoxic activity, Garcinia cowa, Combination index, Breast cancer, Co-Chemotherapy.

Introduction

Breast cancer continues to be a major global health issue for women and a leading cause of mortality.¹ Every year, cancer-related morbidity and mortality continue to increase. This has been complicated with the cost of chemotherapy and its devastating side-effects.² The development of multidrug resistance pathways in cancer cells frequently reduces the efficacy of chemotherapeutic medicines like doxorubicin.³ Long-term use of doxorubicin has also been reported to cause cardiotoxicity.⁴ Despite advancements in treatment, current breast cancer therapies often lack selectivity, leading to significant damage to normal cells and limiting their effectiveness.⁵ As such, It is crucial to develop new cancer treatments that can specifically target and eliminate cancer cells while protecting healthy tissues, thereby improving therapeutic outcomes and reducing adverse side effects.⁶ Thus, one of the main objectives of contemporary scientific study is to find many ways to maximise the effectiveness of doxorubicin in cancer cells and reduce the related toxicity in non-cancerous tissues.⁷

*Corresponding author. E mail: <u>fatmasriwahyuni@phar.unand.ac.id</u> Tel: +62 81374024514

Citation: Ifora I, Hamidi D, Susanti M, Hefni D, Wahyuni FS. Enhancing Chemotherapeutic Efficacy: Synergistic Cytotoxic Effect of *Garcinia cowa* Bark Extract and Doxorubicin in T47D Breast Cancer Cells. Trop J Nat Prod Res. 2025; 9(1): 67 – 72 https://doi.org/10.26538/tjnpr/v9i1.10

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria

The combination of doxorubicin with chemopreventive agents is needed to increase doxorubicin activity, overcome drug resistance, and reduce side effects.8 Developing novel treatment strategies for breast cancer continues to be one of the most challenging fields in cancer research.9 Numerous tropical plants have biological properties that may have medical uses.¹⁰ The family Guttiferae includes Garcinia cowa Roxb, also referred to as "kandis," which is found in tropical rainforests in countries like Thailand, Indonesia, Malaysia, and the Philippines.¹¹ Traditional medicine has utilised several parts of Garcinia cowa. Bark, latex, leaves, and roots in the management of burns, wounds, pain, and inflammation.12 The chemical composition and biological activity of various parts of Garcinia cowa have been studied previously.13 Garcinia cowa contains various active compounds, including cowanin, rubraxanthone, cowaxanthone, and norcowanin.¹³⁻¹⁶ Several studies have consistently reported that a variety of plant extracts possess anticancer activity, including the ability to suppress the growth of T47D breast cancer cells.^{17–20} Xanthone isolated from methanol extract of bark can suppress the growth of MCF-7 cells.²¹ This demonstrates that the Garcinia cowa could serve as a lead in the search for anticancer compounds. This study was conducted to investigate the cytotoxic effects of an ethanolic extract of Garcinia cowa Bark in combination with doxorubicin on T47D breast cancer cells.

Materials and Methods

Plant material

The bark of *Garcinia cowa* Roxb. was collected in December 2022 at Kudu Gantiang, Pariaman City, West Sumatra. The plant material has been deposited at the Herbarium of Andalas University, West Sumatra,

Indonesia, under the voucher number 556-ID/ANDA/XII/2022. Dr. Nurainsa, a botanist from Andalas University's Herbarium, identified the *Garcinia cowa* Roxb. bark.

Preparation of plant material and extraction

Plant materials were sliced into tiny pieces (3-5 mm thick) and allowed to air-dry in a shaded area for 7 days. The dried bark of *Garcinia cowa* was ground into a powder using a traditional grinder. The materials were then soaked for 24 hours at room temperature in 70% ethanol with intermittent stirring and then filtered. This process was repeated thrice. The filtrates were combined and concentrated under a vacuum using a rotary evaporator at 45°C till a brownish semisolid extract was formed. The extract was kept in a refrigerator at 4°C for further pharmacological testing.

Cell culturing procedure

T47D cells, a kind of human breast cancer, were provided for the study by Prof. Masashi Kawaichi of the Nara Institute of Science and Technology in Japan. Vero cells were collected from the Gajah Mada University. T47D and Vero cells were grown in Dulbecco's modified Eagles medium, which contained 10% fetal bovine serum (Gibco, Grand Island, NY, USA), 1% penicillin-1% streptomycin (Gibco, Grand Island, NY, USA), and 0.5% fungizon (Gibco, Grand Island, NY, USA), in a flask in a humidified atmosphere (5% CO₂) at 37°C.²²

Single Cytotoxic assay

T47D cells were seeded in 96-well plate with 1x10⁴ cells/well and divided into control and treatment group. Then, incubated in 37°C with 5% CO2 for 24 hours. Cells were treated with final concentrations of GCBEE were 18.75, 37.5, 75, 150, and 300 $\mu g/mL$ while the concentrations of doxorubicin were 0.001, 0.01, 0.1, 1 and 10 µg/mL. The culture media was withdrawn and the cells were cleaned with PBS (Sigma) after a 48-hour incubation period. 100 µL of diluted 5 mg/mL MTT on PBS (Sigma) was applied to each well after being diluted with Dulbecco's Modified Eagle Medium (DMEM). After four hours of incubation, 10% sodium dodecyl sulfate (SDS) in HCL 0.01 N was added to stop the reaction. After that, the plate was incubated for one night at room temperature in a dark environment. The plate was agitated for ten minutes to ensure that the formazan had dissolved, and then the absorbance was measured at a wavelength of 595 nm using an ELISA reader (Bio-Rad, USA). Every treatment was administered in triplicate, and the cytotoxic activity was quantified using the IC_{50} method, which determines the quantity needed to lower the population's absorbance of cells by 50% in comparison to the untreated (control) cells.

Combination Cytotoxic Assay

The combination test was carried out using the same method after determining the IC₅₀ value of a single treatment. In the combination test, the concentrations of GCBEE were 520, 260, 130, 65, and 32 µg/mL, while the concentrations of dox were 0.104, 0.052, 0.026, 0.013, and 0.007 µg/mL, corresponding to 4, 2, 1, ½, and ¼ of their respective IC₅₀ values. Next, the Combination Index (CI) was calculated. Based on the Chou-Talalay method, CompuSyn software calculates CI values that show the impact of medication combinations.²³

Selectivity index

To determine the cytotoxic selectivity of GCBEE, the Selectivity Index (SI) was calculated according to the following equation (1).²⁵

$$SI = \frac{IC50 \text{ on Normal Cells}}{IC50 \text{ on Cancer Cells}}$$
(1)

Extract is non-selective if SI is less than 3, but selective if SI is greater than $3.^{26}$

Statistical analysis

The *in vitro* experiment data were presented as mean \pm S.E.M. Using the obtained absorbance, the percentage (%) of cell viability was determined. Graphical illustration of the viability percentage and IC₅₀ value data were analyzed and presented using GraphPad Prism (GraphPad Prism, 9.0.0). The effects of medication combinations are displayed by CI values calculated using CompuSyn software (Vers.1.0).

Results and Discussion

Single Cytotoxic assay and Selectivity index Cytotoxicity is categorized by the National Cancer Institute (NCI) based on IC_{50} values. Extracts with IC_{50} values less than 20 µg/mL are

Table 1: Interpretation of CI value.²⁴

CI	Interprestation	CI	Interprestation
< 0.1	Very strongly	0.90-1.10	Nearly additive
	synergistic		
0.1-0.3	Strongly synergistic	1.10-1.20	Slight antagonist
0.3-0.7	Synerggistic	1.20-1.45	Middle antagonist
0.7-0.85	Moderate synergistic	1.45-3.3	Antagonist
0.85-	Slight synergistic	>3.3	Strongly
0.90			antagonist

considered highly active, moderately active if IC₅₀ 21-200 µg/mL, weakly active if IC₅₀ = 201–500 µg/ml and IC₅₀ values above 500 µg/mL indicate inactive or no cytotoxicity.²⁷ The inhibitory concentration (IC₅₀) is the amount needed to prevent 50% of cell growth. Lower IC₅₀ values signify larger efficacy in inhibiting cell division.^{28,29}

The GCBEE exhibited an ICso of 130 µg/mL on T47D cells, indicating moderate cytotoxic activity. In contrast, dox showed a significantly lower IC50 of 0.026 µg/mL, highlighting its potent cytotoxicity as a chemotherapeutic agent. GCBEE's IC50 on Vero cells was considerably higher, at 1.955 µg/mL, suggesting a lower toxic effect on noncancerous cells. The selectivity index (SI) is a crucial metric to evaluate the specificity of GCBEE towards cancer cells relative to non-cancerous cells. A higher SI indicates better selectivity and potential as an anticancer agent.³⁰ For GCBEE, the SI was 15 for T47D cells versus Vero cells (Table 2). An SI > 3 is considered to exhibit high selectivity, underscoring GCBEE's promising anti-cancer potential with relatively low toxicity towards normal cells. Dox did not have an SI calculation in this study as the IC50 on Vero cells was not determined, but it is known to have significant toxicity in both cancer and normal cells.³¹ The results of this study demonstrate that Garcinia cowa bark ethanol extract (GCBEE) possesses selective cytotoxic activity against T47D breast cancer cells, as evidenced by an IC50 value of 130 µg/mL and a high selectivity index (SI = 15) when compared to Vero cells. The SI value exceeding 3 suggests that GCBEE has a strong potential as an anti-cancer agent with reduced toxicity towards normal, non-cancerous cells.²⁶ This high selectivity makes GCBEE a promising candidate for further investigation in cancer therapy, especially as a natural compound with possibly fewer side effects than conventional chemotherapeutic agents like doxorubicin.

Table 2: IC_{50} of GCBEE and dox on T47D cells, Vero cells and SI^a values of GCBEE at 48 h after administration.

Cell Lines				
IC ₅₀ (µg/mL)	T47D	Vero		
GCBEE	130	1.955		
Dox	0.026	-		
^a SI				
GCBEE	15	1		

^aSI : refers to Selectivity Index.SI values > 3 was considered as high selectivity.

- : IC₅₀ was not calculated.

ISSN 2616-0684 (Print) ISSN 2616-0692 (Electronic)

Although GCBEE's IC₅₀ value is higher than that of doxorubicin (0.026 μ g/mL) (Figure 1), its significantly lower cytotoxicity towards Vero cells offers a considerable advantage in reducing side effects. Doxorubicin, while effective at killing cancer cells, is known for its severe side effects due to its non-selective nature, affecting both cancerous and normal cells. The moderate cytotoxicity of GCBEE towards cancer cells combined with its minimal impact on normal cells supports its potential as a safer alternative or complementary therapy in breast cancer treatment. Future research could further explore the bioactive compounds within GCBEE responsible for the cytotoxic

activity. Additionally, *in vivo* studies are needed to confirm the therapeutic potential and safety of GCBEE in a biological system. The development of targeted delivery systems could further enhance its efficacy, reduce required dosages, and mitigate any residual toxicity towards normal cells. This study provides a solid foundation for the continued exploration of GCBEE as a natural anti-cancer agent, offering a balance between efficacy and safety that is critical for developing new cancer therapies.

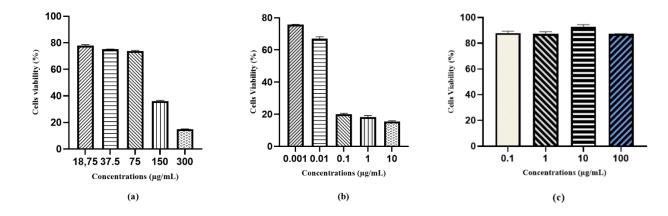


Figure 1: Cell viability (%) profile of T47D cells following treatment with GCBEE (IC₅₀= 130 μ g/mL) (a), doxorubicin (IC₅₀= 0.026 μ g/mL) (b), and Cell survivability (%) of Vero cells following treatment with GCBEE (IC₅₀= 1.955 μ g/mL) (c) for 48 h single test application. Viability values are presented as mean (n = 3) ± SEM.

Combination Cytotoxic Assay

The cytotoxic effects of GCBEE on T47D cells are concentrationdependent (Figure 2a). As the concentration increases from $32.5 \,\mu$ g/mL to $520 \,\mu$ g/mL, cell viability decreases significantly. The viability of the cells decreases significantly at the maximum dose of $520 \,\mu$ g/mL, suggesting that GCBEE is an effective means of lowering cell viability on its own. Figure 2b shows the cytotoxic effects of dox on T47D cells. Similar to GCBEE, dox exhibits a concentration-dependent reduction in cell viability. At lower concentrations (0.007 μ g/mL), the impact on cell viability is minimal. However, as the concentration rises to 0.104 μ g/mL, there is a substantial reduction in cell viability, indicating dox's well-known efficacy as a chemotherapeutic agent. These findings are consistent with dox's mechanism of action, which involves DNA intercalation and inhibition of topoisomerase II, resulting in the death of rapidly proliferating cells, like cancer cells.³² This validates dox's potency against breast cancer cells in a controlled in vitro environment.

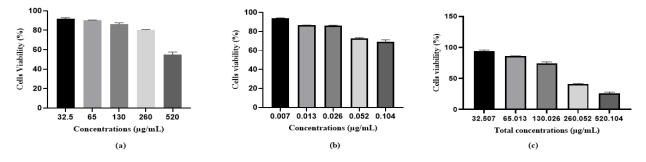


Figure 2: Cell viability (%) profile of GCBEE (a), dox (b), and combination GCBEE + dox (c) on T47D cells in the combination test application. All values are expressed as means $(n = 3) \pm SEM$.

The combined treatment of GCBEE and dox demonstrates an even greater reduction in cell viability compared to each compound alone, particularly at higher concentrations. At 520.104μ g/mL (Figure 2c), the combination achieves a near-complete reduction in cell viability. The combination effect is likely synergistic, as indicated by a greater-than-expected reduction in cell viability, which suggests that GCBEE may enhance the cytotoxic effects of dox on T47D cells. This could be due to complementary mechanisms of action between GCBEE and dox. These findings support the potential of GCBEE as an adjuvant to traditional chemotherapy, especially to increase dox's effectiveness in the treatment of breast cancer. Further studies are necessary to explore

the exact molecular interactions and to determine if similar effects can be achieved in vivo.

The most common usage of drug combinations is to treat the chronic diseases, like AIDS and cancer. The major goals are to reduce dosage and toxicity, produce a synergistic therapeutic effect, and prevent the onset of drug resistance.³³ The quantitative definitions of synergism (CI < 1), additive effect (CI = 1), and antagonism (CI > 1) as shown in Table 1, as well as the techniques for their computer simulation, are obtained by combining the median-effect equation with the CI equation.^{24,34} Based on the Chou and Talalay approach, the CI was computed to

ascertain if the combination of GCBEE and dox has an antagonistic, additive, or synergistic effect. 35

The Combination Index (CI) values for T47D breast cancer cells when *Garcinia cowa* bark ethanol extract (GCBEE) and doxorubicin were combined at different doses are shown in Table 3, as determined by the Chou-Talalay method using Compusyn software. At a total dose of 520.104 µg/mL, the combination affects 74.2% of the T47D cells (Fa = 0.742), with a CI value of 0.19, indicating a strong synergistic effect. A dose of 260.052 µg/mL affects 58.9% of the cells (Fa = 0.589) with a CI value of 0.25, suggesting a synergistic interaction. The 130.026 µg/mL dose affects 26.0% of the cells (Fa = 0.260) with a CI value of 0.89, still indicating synergism, albeit weaker compared to the higher doses. At a lower dose of 65.013 µg/mL, the fraction affected drops to 14.2% (Fa = 0.142), and the CI value increases to 1.27, suggesting antagonism. The smallest dose of 32.507 µg/mL affecteds only 6.3% of the cells (Fa = 0.063) with a CI value of 2.23, indicating strong antagonism.

 Table 3: Combination Index values of combination GCBEE

 and doxorubicin on T47D cells

Total Dose	Fraction	Combination Index	
(µg/mL)	Affected (Fa)	(CI) Value	
520.104	0.742	0.19	
260.052	0.589	0.25	
130.026	0.260	0.89	
65.013	0.142	1.27	
32.07	0.063	2.23	

The cytotoxicity analysis of the combination of *Garcinia cowa* bark ethanol extract (GCBEE) and doxorubicin on T47D cells reveals a significant synergistic effect at higher doses, as demonstrated by CI values well below 1 at doses of 520.104 μ g/mL and 260.052 μ g/mL. The highest degree of synergy is observed at the dose of 520.104

 μ g/mL, where 74.2% of the cells were killed, and the CI value reaches an impressive 0.19, indicating potent synergy. This suggests that at higher dose combinations, the two agents work in concert to exert enhanced cytotoxic effects, likely reducing the required concentration of doxorubicin to achieve similar levels of cytotoxicity, thereby potentially minimizing its toxic side effects.

The dose-effect curve Figure 3a clearly demonstrates that doxorubicin (dox) alone (red curve) has a potent cytotoxic effect at very low doses, while GCBEE (green curve) requires higher doses to reach similar levels of efficacy. The combination of the two (GCBDox, blue curve) shows an improved response compared to GCBEE alone, suggesting that combining these agents enhanceds their cytotoxic effect, particularly at higher doses.

The combination index (CI) plot (Figure 3b) further supports this finding. CI values below 1 across a range of affected fractions (Fa) indicate a synergistic effect between GCBEE and dox, especially as the fraction affected increases. This synergy suggests that the combination is more effective in killing cancer cells compared to each agent used individually, particularly at doses higher than 0.2 Fa. However, as the dose decreases, the CI values progressively increase, with the combination becoming antagonistic at the lower doses of 65.013 µg/mL and 32.507 µg/mL. This shift from synergy to antagonism at lower doses may indicate that below a certain concentration, the interaction between GCBEE and dox becomes antagonistic, possibly due to inadequate cellular exposure to either agent. This finding is also in line with the isobologram curve (Figure 3c), The combination of Garcinia cowa extract and doxorubicin showed a synergistic effect at Fa 0.5, 0.75, and 0.9, with the isobologram curve below the straight line, indicating that the combination dose is more effective than the single agent. Similar findings related to the synergistic properties of the combination of natural extracts and conventional cytotoxic drugs have also been reported by several researchers, such as those conducted by previous researchers who reported that synergistic effects are demonstrated when an anti-cancer medication and a natural chemical are used together, improving total therapeutic activity against cancer cells.36

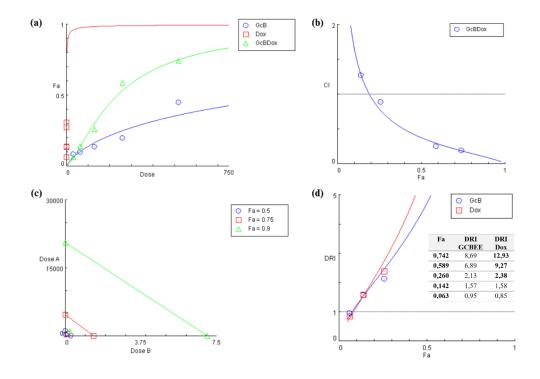


Figure 3: Concentration-effect curve (a), combination index plot (b), and isobologram (c) drug reduction index plot (d).

Additionally, other studies also reports that substances derived from plants have a significant influence as therapeutic agents, particularly cytotoxic agents, whether used alone or combined with other medications.³⁷

The DRI plot and the accompanying table of DRI (Figure 3d) values provide essential insights into how much the doses of GCBEE and doxorubicin can be reduced when used in combination, without compromising efficacy.At an Fa of 0.742, the DRI for GCBEE is 8.69106 and for dox is 12.8295, indicating that the simultaneous use of both drugs significantly reduced the dosage while still achieving high levels of cell death. This reflects a considerable improvement in therapeutic potential, especially for doxorubicin, whose dose reduction could help alleviate its known toxic side effects, such as cardiotoxicity. As the Fa decreases, the DRI values decrease as well, suggesting that dose reduction is less prominent at lower fractions affected. However, even at lower Fa (e.g., 0.142), the DRI values remain above 1 for both agents, reinforcing the idea that dose reduction is still possible across a broad range of therapeutic effects. GCBEE shows consistent dose reduction across various Fa levels, implying that this natural extract, when combined with doxorubicin, can enhance the overall therapeutic response and mitigate the need for higher doses of the chemotherapeutic agent. The DRI data suggest that combining GCBEE with doxorubicin can lead to synergistic cytotoxicity against T47D breast cancer cells. This synergism allows for lower doses of both agents to achieve similar therapeutic effects, which is particularly significant for doxorubicin due to its well-documented toxicity profile. Reducing the dose of doxorubicin while maintaining its anticancer efficacy offers a significant clinical advantage, as it could minimize adverse effects, improve patient tolerance, and enhance the quality of life for those undergoing chemotherapy.

Conclusion

These results strongly indicate that *Garcinia cowa* Bark ethanolic extract displayed higher selectivity for T47D cells over Vero cells and synergistically enhanced cytotoxic activity of doxorubicin. These findings suggest the potential of GCBEE as a selective and synergistic adjunct to dox in cancer therapy. The results of this study open avenues for further preclinical and clinical investigations to explore the full therapeutic potential of this combination. Future studies should aim to confirm these synergistic effects *in vivo* and elucidate the underlying molecular mechanisms behind the observed synergy, thus paving the way for developing more effective and safer combination therapies for breast cancer treatment.

Conflict of Interest

The authors declare no conflicts of interest.

Author's Declaration

The authors hereby declare that the work presented in this article are original and that any liability for claims relating to the content of this article will be borne by them.

Acknowledgments

The research was funded by the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia through the Basic Research scheme of the Doctoral Dissertation Research cluster with contract No. 041/E5/PG.02.00.PL/2024 subcontract No. 14/UN16.19/PT.01.03/PL/2024, for which the authors are grateful. We want to express our sincere gratitude to Prof. Masashi Kawaichi of the Nara Institute of Science and Technology, Japan, for providing the T47D breast cancer cells for this study, with special assistance from Gajah Mada University.

References

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, bray F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide 2221.

for 36 cancers in 185 Countries. CA Cancer J Clin. 2021;71(3):209–249.

- 2. Globocan. Cancer Incident in Indonesia. Int agency Res cancer. 2020;858:1–2.
- 3. Sun YS, Zhao Z, Yang ZN, Xu F, Lu HJ, Zhu ZY, Shi W, Jiang J, Yau P, Zhu H. Risk factors and preventions of breast cancer. Int J Biol Sci. 2017;13(11):1387–1397.
- Rawat PS, Jaiswal A, Khurana A, Bhatti JS, Navik U. Doxorubicin-induced cardiotoxicity: An update on the molecular mechanism and novel therapeutic strategies for effective management. Biomed Pharmacother [Internet]. 2021;139:111708. Available from: https://doi.org/10.1016/j.biopha.2021.111708
- Perkins DW, Steiner I, Haider S, Robertson D, Buus R, O'Leary L, Clare M, Isacke. Therapy-induced normal tissue damage promotes breast cancer metastasis. iScience [Internet]. 2024;27(1):108503. Available from: https://doi.org/10.1016/j.isci.2023.108503
- Navya PN, Kaphle A, Srinivas SP, Bhargava SK, Rotello VM, Daima HK. Current trends and challenges in cancer management and therapy using designer nanomaterials. Nano Converg [Internet]. 2019;6(1):1–30. Available from: https://doi.org/10.1186/s40580-019-0193-2
- Hanušová V, Boušová I, Skálová L. Possibilities to increase the effectiveness of doxorubicin in cancer cells killing. Drug Metab Rev. 2011;43(4):540–557.
- Talib WH, Awajan D, Hamed RA, Azzam AO, Mahmod AI, Al-yasari IH. Combination Anticancer Therapies Using Selected Phytochemicals. Molecules. 2022;27(5452):1–50.
- Witzel I, Müller V. Targeted Therapies in Breast Cancer: New Approaches and Old Challenges. Breast Care. 2015;10(3):157–158.
- Shaikh AM, Shrivastava B, Apte KG, Navale SD. Medicinal Plants as Potential Source of Anticancer Agents: A Review. J Pharmacogn Phytochem JPPB) APT Res Found. 2016;5(2):291–295.
- Lim T. Cajanus cajan. In: Edible Medicinal And Non-Medicinal Plants. Springer, Dordrecht. Springer, Dordrecht; 2012. 549–568 p.
- Lin F, Luo B, Cheng Z, Li P, Long C. Ethnobotanical Study on *Garcinia (Clusiaceae)* in China. Acta Soc Bot Pol. 2021;90.
- Ritthiwigrom T, Laphookhieo S, Pyne SG. Chemical constituents and biological activities of *Garcinia cowa* Roxb. Maejo Int J Sci Technol. 2013;7(2):212–231.
- Hefni D, Dachriyanus, Wahyuni FS, Yerizel E, Arisanty D, Yusra LN. *Cowanin*, a Cytotoxic Xanthone from Asam Kandis (*Garcinia cowa*, Roxb.) Reduced Cell Migration and Induced Cell Cycle Arrest on T47D Human Cancer Cell. Int J Adv Sci Eng Inf Technol. 2020;10(5):2164–2169.
- Siridechakorn I, Maneerat W, Sripisut T, Ritthiwigrom T, Cheenpracha S, Laphookhieo S. Biphenyl and xanthone derivatives from the twigs of a *Garcinia* sp. (*Clusiaceae*). Phytochem Lett [Internet]. 2014;8(1):77–80. Available from: http://dx.doi.org/10.1016/j.phytol.2014.02.004
- Wahyuni FS. Anticancer Compound from *Garcinia cowa* Roxb Induce Cell Cycle Arrest. Int J Pharm Sci Rev Res Relat. 2015;166–168.
- Husni E, Nahari F, Wirasti Y, Wahyuni FS, Dachriyanus. Cytotoxicity study of ethanol extract of the stem bark of asam kandis (*Garcinia cowa* Roxb.) on T47D breast cancer cell line. Asian Pac J Trop Biomed. 2015;5(3):249–252.
- Edityaningrum CA, Khairurrizki A, Nurani LH, Bachri MS, Yuliani S, Utami D, Kintako, Nurkhasanah, Irham LM, Zakaria ZA. Co-Chemotherapy Effect of The Extract of *Hibiscus Sabdariffa* and Cisplatin Against. Trop J Nat Prod Res. 2024;8:7509–7513.
- Rahmawati N, Ismail NH, Hamidi D, Wahyuni FS. Cytotoxic Activity Screening of Various Uncaria Spp Plants on T47d Breast Cancer. Trop J Nat Prod Res. 2023;7 :2218–
- 20. Engel N, Oppermann C, Falodun A, Kragl U. Proliferative

effects of five traditional Nigerian medicinal plant extracts on human breast and bone cancer cell lines. J Ethnopharmacol. 2011;137(2):1003–1010.

- Wahyuni FS, Shaari K, Stanslas J, Lajis NH, Dachriyanus. Cytotoxic xanthones from the stem bark of *Garcinia cowa* Roxb. J Chem Pharm Res. 2015; 7(1): 227-236.
- Nugroho AE, Hermawan A, Putri DDP, Novika A, Meiyanto E. Combinational effects of hexane insoluble fraction of *Ficus septica* Burm. F. and doxorubicin chemotherapy on T47D breast cancer cells. Asian Pac J Trop Biomed. 2013;3(4):297–302.
- Martin N & Chou T. for Drug Combinations and for General Dose-Effect Analysis. Vol. 2005. 2010. 3–4 p.
- Artanti AN, Pujiastuti UH, Prihapsara F, Rakhmawati R. Synergistic Cytotoxicity Effect by Combination of Methanol Extract of Parijoto Fruit (*Medinilla speciosa* Reinw. ex. Bl) and Cisplatin Against Hela Cell Line. Indones J Cancer Chemoprevention. 2020;11(1):16.
- Rivanti E, Shabrina BA, Nurzijah I, Ayu C, Hermawan A. Heartwood of Secang (*Caesalpinia sappan* L.) Ethanolic Extract Show Selective Cytotoxic Activities on T47D and Widr Cells But Not on Hela Cells. Indones J Cancer Chemoprevention. 2017;7(2):60.
- Da'i M, Meilinasary KA, Suhendi A, Haryanti S. Selectivity Index of *Alpinia galanga* Extract and 1' -Acetoxychavicol Acetate on Cancer Cell Lines. Indones J Cancer Chemoprevention. 2019:95–100.
- Anywar GU, Kakudidi E, Oryem-Origa H, Schubert A, Jassoy C. Cytotoxicity of Medicinal Plant Species Used by Traditional Healers in Treating People Suffering From HIV/AIDS in Uganda. Front Toxicol. 2022;4; 1–8.
- He Y, Zhu Q, Chen M, Huang Q, Wang W, Li Q, Huang Y, Di W. The changing 50% inhibitory concentration (IC₅₀) of cisplatin: A pilot study on the artifacts of the MTT assay and the precise measurement of density-dependent chemoresistance in ovarian cancer. Oncotarget. 2016;7(43):70803–70821.
- Okafor CE, Ijoma IK, Igboamalu CA, Ezebalu CE, Eze CF, Osita-Chikeze JC, Uzo CE, Ekwuekwe AL. Secondary

metabolites, spectra characterization, and antioxidant correlation analysis of the polar and nonpolar extracts of *Bryophyllum pinnatum* (Lam) Oken. Biotechnologia. 2024;105(2):121–136.

- Lica JJ, Wiecz M, Grabe GJ, Heldt M, Jancz M, Misiak M, Gucwa K, Brankiewicz W, Maciejewska N, Stupak A, Baginski M, Rolka K, Hellmann A, Składanowski A . Effective Drug Concentration and Selectivity Depends on Fraction of Primitive Cells. Int J Mol Sci. 2021;22(4931):1– 24.
- Fraczkowska K, Bacia M, Przybyło M, Drabik D, Kaczorowska A, Rybka J, Stefanko E, Drobczynskic S, Masajada J, Podbielska H, Wrobelb T, Kopaczynskaa M. Alterations of biomechanics in cancer and normal cells induced by doxorubicin. Biomed Pharmacother. 2018;97:1195–1203.
- Mujwar S, Kołat D, Kciuk M, Gieleci A, Kałuzi Z. Doxorubicin — An Agent with Multiple Mechanisms of Anticancer Activity. Cells. 2023;12(659):26–32.
- Ashton JC. Drug combination studies and their synergy quantification using the chou-talalay method-letter. Cancer Res. 2015;75(11):2400.
- Huang L, Jiang Y, Chen Y. Predicting Drug Combination Index and Simulating the Network-Regulation Dynamics by Mathematical Modeling of Drug-Targeted EGFR-ERK Signaling Pathway. Sci Rep [Internet]. 2017;7:1–9. Available from: http://dx.doi.org/10.1038/srep40752
- 35. Chou TC. Drug combination studies and their synergy quantification using the chou-talalay method. Cancer Res. 2010;70(2):440–446.
- 36. Boţa M, Vlaia L, Jîjie AR, Marcovici I, Crişan F, Oancea C, Dehelean CA, Dehelean T, Moacă E. Exploring Synergistic Interactions between Natural Compounds and Conventional Chemotherapeutic Drugs in Preclinical Models of Lung Cancer. Pharmaceuticals. 2024;17(5):1–36.
- Pezzani R, Salehi B, Vitalini S, Iriti M, Zuñiga FA, Sharifi-Rad J, Martorell M, Martins N. Synergistic effects of plant derivatives and conventional chemotherapeutic agents: An update on the cancer perspective. Med. 2019;55(4):1–16.