



LC-MS Chemical Profiling of Dichloromethane Fraction of Methanol Extract of *Syzygium samarangense* Stem Bark

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ABSTRACT

Prior studies have demonstrated that *Syzygium samarangense* possesses antioxidant, antifungal, and anti-inflammatory properties as well as other advantages. The fruit is widely consumed, yet the medical benefits of the stem bark as an alternative therapeutic option have not been explored. This research was performed to determine the specific chemicals contained in the dichloromethane fraction of the methanol extract of this plant. The powdered plant sample was extracted with methanol by maceration and fractionated with dichloromethane. The resulting dichloromethane fraction was then concentrated, and its chemical constituents were identified using LC-MS based on the Willey and NIST libraries. The findings of this research showed the presence of 40 phytoconstituents in five main classes of secondary metabolites: phenolics, flavonoids, terpenoids, steroids, and others. This study explains the basic structures, compound derivatives, biological functions of the compounds identified in F2 with reference to relevant literature. The stem bark of *S. samarangense* predominantly contains compounds with significant bioactivity, including antioxidants, antifungal, anti-inflammatory, anticancer, and antiviral properties. This research offers valuable insight into the groundwork for the utilization of this plant and the selection of natural and synthetic substances.

Keywords: Chemical profile, Liquid Chromatography-Mass Spectrometry, methanol extract, *Syzygium samarangense*.

Introduction

Syzygium samarangense or Jambu Semarang (in Indonesian), is one of Indonesia's local plants whose medical benefits still need further exploration. However, its mineral compositions, including iron, potassium, calcium, sodium, and magnesium, are higher than that of *S. aqueum*.¹ In addition to having higher magnesium levels, it also contains tannins and mild antibiotic properties against *S. aureus*, *M. smegmatis*, and *C. albicans*.² The plant also contains desmethoxymatteucinol, 5-*O*-methyl-4'-desmethoxymatteucinol, oleanic acid, and β -sitosterol as bioactive compounds.³⁻⁵ Furthermore, the plant also has high nutritional content: protein (92.9%), carbohydrates (6%), crude fibre (0.46%), and ash content (0.21%).⁶ Several studies have reported the leaves' analgesic, anti-inflammatory, and remarkable CNS effects.^{1,6} The extract has shown potent antihelmintic properties.^{4,5} Reports also showed that the stem bark extract of this plant has antidiabetic potential with inhibitory activity against the α -glucosidase enzyme.⁷ Phytochemical analysis of the stem bark extract showed the presence of terpenoids, steroids, tannins, saponins, phenolics, and flavonoids,⁸ and possessed excellent antioxidant properties.^{9,10} The LC-MS analysis of a fraction from the VLC of the dichloromethane extract showed the presence of four flavonoid compounds, namely pinocembrin, uvangoletin, sterkurensin, and aurethiacin.¹¹

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There are no reports regarding the study of the chemical profile of the dichloromethane fraction resulting from the partitioning of the methanol extract of *S. samarangense* stem bark, making researchers interested in conducting this research.

The current aimed to investigate the chemical profile of the dichloromethane fraction of the crude methanol extract through LC-MS analysis, followed by an in-depth study of the diversity of chemical structures and bioactivity properties with a view of establishing its medicinal potential and use as a herbal agent/medicine.

Methods

Plant Collection and Identification

The plant stem bark sample of *S. samarangense* was collected from a local area in Kediri, East Java, located between Latitude 7.8480° S and longitude 112.0178° E, Indonesia, in October 2018. The plant was identified at the Herbarium-LIPI, Purwodadi, East Java, Indonesia, where voucher specimen no. 1498/IPH.06/HM/X/2018 was deposited.

Preparation, Extraction, and Fraction of Plant material

The stem bark of *S. samarangense* (c.a. 15 kg) was washed with tap water and dried under sunlight for one week. It was then dried in an oven at a reduced temperature (not more than 50 °C). The dried material was ground to a powder using an electric grinder to obtain 9.5 kg and transferred to an airtight container.¹¹ The powdered stem bark (5 kg) of *S. samarangense* was macerated using methanol (c.a. 15 L) at ambient temperature (25 °C). The container with its contents was sealed by foil and kept for 24 hours, accompanied by occasional shaking and stirring. The whole mixture was then filtered using a Buchner funnel, and the filtrate was concentrated at 50 °C with a vacuum rotary evaporator to obtain a thick, dark-brown crude extract (25 g), kept in a refrigerator at 4 °C until further use.

The crude extract was dissolved slightly in methanol and water (300 mL) and then partitioned using *n*-hexane (300 mL x 3, F1) in a separating funnel. The aqueous fraction was then separated with dichloromethane (300 mL x 3, F2) in a separating funnel. The fractions

were concentrated at 40 °C with a vacuum rotary evaporator. ~~under a vacuum using a rotary evaporator at 40°C.~~ The dried fraction (F2) obtained was stored in a refrigerator until further analysis.

Identification Using LC-MS ESI

The chemical content of the F2 fraction was analysed using the LC-MS. LC-MS instrument (Shimadzu LC-MS 8040) equipped with a Shimadzu Shim Pack FC-ODS column (2 mm x 150 mm, 3 µm) and a column temperature of 35 °C was used. The sample injection volume was 1 µL with isocratic separation (the mobile phase used was 90% methanol with water) at a flow rate of 0.5 mL/min. The LC-MS analysis was conducted in negative ion mode based on the following parameters: source temperature of 100 °C, cone sampling voltage of 23 eV, capillary voltage of 3.0 kV, solvent discharge temperature of 350 °C, and solvent gas flow of 60 mL/hour. The mass spectrum detection range was set between m/z 10-1000 using ESI negative ion mode, with a scanning duration of 0.6 seconds per scan and a total running time of 80 minutes. LC-MS has been an effective method for identifying compounds in a matrix¹³.

Results and Discussion

The chemical profile of the dichloromethane fraction (F2) of *S. samarangense* investigated using LC-MS is shown in Figure 1. There are 40 compounds peaks identified with the MS detector in Table 1. Polyphenolic compounds (flavonoids, flavanones, flavonols), phytosterols, etc. were identified in F2. The basic structure of flavonoid and its derivatives were shown in Figure 2. Those with simple structures include 1, 3, and 5. Compound 1 (methyl salicylate) is mainly found in *Gynura procumbens*, which has a spicy taste commonly called mint.¹⁴ When making deep heating liniments (like Bengay) to relieve joint and

muscular pain, methyl salicylate is utilized in high doses as an analgesic and rubefacient. On the other hand, this substance in the body metabolizes into salicylates, including salicylic acid, a well-known NSAID.^{15,16} Compound 3 (gallic acid) is widely found in many parts of plants¹⁷ and offers antioxidant activity. Compound 3 is also widely used to measure the total phenolic content of an extract¹⁸. This compound can be utilized as an antioxidant¹⁹, antineoplastic agent²⁰, astringent²¹, and cyclooxygenase-2 (COX-2) inhibitor.²² Finally, compound 5 (eugenin) is mainly produced by *Cylindropuntia bigelovi*²³ and *Daucus carota* plants.²⁴ This compound is well-known as a potential HIF-1- α inhibitor.²³

Other phenolic derivatives identified in the fraction were tannins and phenylpropanoid groups. Tannins are phenolic compounds with more than one hydroxyl group and a more complex structure, generally called polyphenols.²⁵ Compound 37 (strictinin) is a tannin belonging to the ellagitannin group, predominantly in *Alnus japonica*²⁶ and *Balanophora japonicssa*,²⁷ was identified in this study.

A phenylpropanoid group is a group of compounds with an aromatic ring substituted with a propene tail, which is generally synthesized from the amino acid phenylalanine and tyrosine pathways.²⁸ Compounds belonging to the phenylpropanoid group are compounds 2 and 6, classified as phenylpropene. Compounds 2 (eugenol) and 6 (eugenol acetate) are mainly found in clove plants and are commonly called clove oil.^{29,30} Compound 6 is a derivative of compound 2 substituted by an acetyl group; both of these compounds have antioxidant, insecticidal³⁰, antimicrobial³¹, and potency as antiviral inhibitor HIV-1 protease.³²

Flavonoids are polyphenolics found in plants with a C6-C3-C6 basic structure with carbon rings.³³ Some flavonoids, namely the chalcone and flavanone groups, were identified in the dichloromethane fraction (F2) of *S. samarangense*.

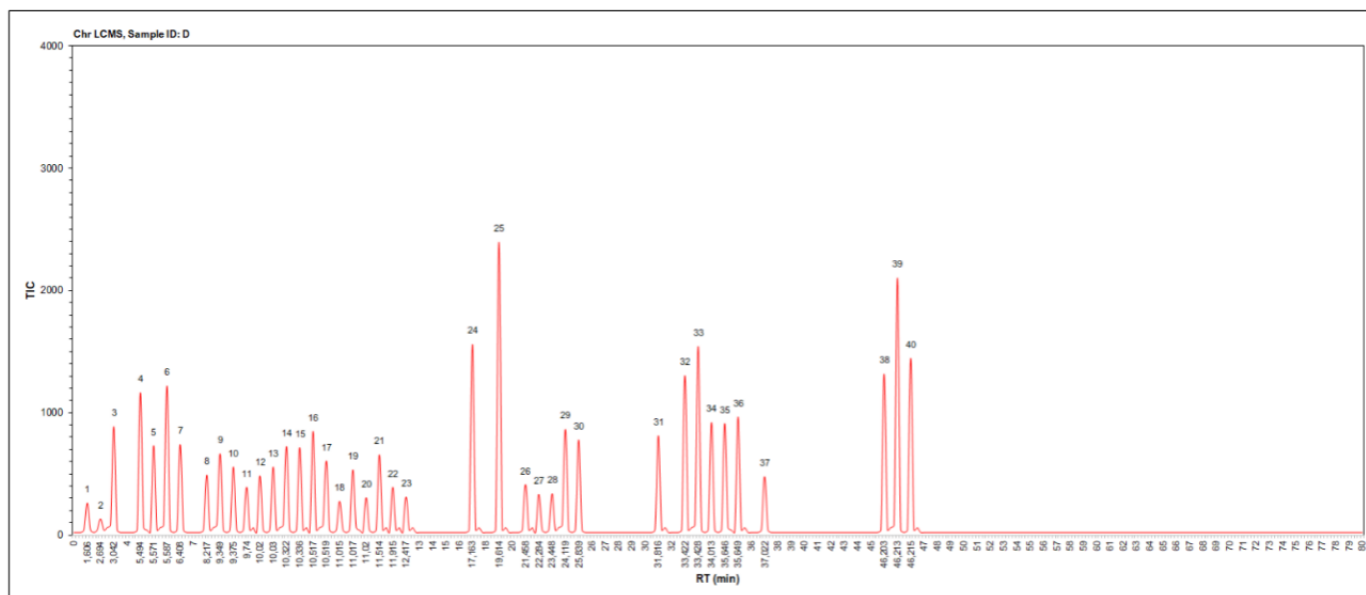


Figure 1: LC-MS Chromatogram of F2 Fraction (Dichloromethane Fraction)

The chalcone group has an open flavonoid basic structure with two rings (A and B) attached by an α,β -unsaturated carbonyl system with different substituents.³³ The chalcone group has significant bioactivity and inhibitory activity against NF- κ B receptors implicated in cancers.³⁴ The chalcone compounds identified in the LC-MS chromatogram include compounds 11, 12, 13, 15, 16, 18, 19, and 22. Compound 11 (uvangoletin) is also found in *Sarcandra glabra*³⁵ and *Monanthes trichocarpas*.³⁶ Several researchers reported that this compound has anticancer and antiviral activities. Compound 12 (stercurensin), also found in *Cleistocalyx operculatus*³⁷ and *Comptonia peregrina*³⁸, has neuraminidase antiviral activity against two influenza viruses, H1N1 and H9N2.³⁷ Similar to compound 12, is compound 13 (demethoxymatteucinol), commonly found in *Pentarrhizidium*

*orientale*³⁹ and *Cleistocalyx operculatus*³⁷, also exhibits the antiviral properties against the influenza viruses.^{37,39} Compound 15, generally called myrigalone H, can be found in *Myrica gale*⁴⁰ and is an mTOR inhibitor in cancer cells.⁴¹ Meanwhile, compound 16 (4',6'-dihydroxy-3',5'-dimethyl-2'-methoxychalcone) can be found in *S. nervosum*⁴² and has antiviral neuraminidase function against two influenza viruses, H1N1 and H9N2.^{37,39} Compound 18 (aurentiacin) found in *Myrica pensylvanica*³⁸ possesses an anti-inflammatory activity tested on mouse macrophages.^{43,44} Compound 19 (2',4'-dihydroxy-6'-methoxy-3',5'-dimethylchalcone) can be found in *S. nervosum*⁴² with similar bioactivity as compound 16.^{37,39} Compound 22 (syzygiol) can be isolated from *S. polycephaloides*, where this compound has antitumor activity against skin tumours.⁴⁵

Flavanones are the other flavonoid derivative identified in the fraction. The flavanone group generally has 15 carbon rings with two phenyl groups (A and B), a heterocyclic ring (oxygen-embedded carbon ring), and a ketone group on carbon number 4. This group has other derivatives with several substituted hydroxyl groups in its basic structure. Flavanones that are substituted with one hydroxyl group are called hydroxyflavanones. Compound 17 (7-hydroxy-5-methoxy-6,8-dimethylflavanone) is a dihydroxyflavone derivative of pinocembrin where the hydroxyl group attached at C-5 is methylated. This compound can be found in *S. samarangense*⁴⁶ and *Couroupita guianensis*.⁴⁷ The next in this group is a substituted compound with two hydroxyl groups called dihydroxyflavone³³, in which compounds belonging to the dihydroxyflavone groups are compounds 8, 9, 10, and 20 (Table 1). Pinocembrin and its derivatives can be found predominately in *Piper sarmentosum*⁴⁸ and *Cryptocarya chartacea*.⁴⁹ In general, this group exhibits antioxidant⁵⁰, anticancer^{51,52}, anti-inflammatory¹², antineoplastic⁵³ activities, and vasodilatory and neuroprotective properties.⁵⁴ The next hydroxylated flavone group with four hydroxyl groups is tetrahydroxyflavones. A compound belonging to this group found in the fraction is compound 14 (kaempferol), which is often extracted from Witch-hazel⁵⁵, grapefruit⁵⁶, and *Lotus ucrainicus*.⁵⁷ Kaempferol is a well-known antioxidant that works by reducing oxidative stress.⁵⁸ It is often used in cancer treatment⁵⁹ and has antibacterial activity.⁶⁰

The next group is that with five hydroxyl groups, commonly referred to as pentahydroxyflavone. Compound 29 (myricitrin), a pentahydroxyflavone myricetin derivative (Figure 4), belongs to this group. The hydroxyl group attached at C-3 is substituted with an α -L-rhamnopyranosyl group. This compound is found and has been isolated from *Myrica cerifera*⁶¹, *S. levinei*⁶², and *Limonium aureum*.^{63,64} This

compound has antiallergic properties⁶⁵, and its protein kinase C inhibitors have a significant role in cancer treatment.⁶⁶ Another group, characterized by six hydroxyl groups, is commonly referred to as hexahydroxyflavone. The compounds found in this group are myricetin and its derivatives, compounds 21, 31, and 36. Many myricetins and their derivatives are found in *Morella rubra*⁶⁷ and *Ficus auriculata*⁶⁸. These compounds have anti-inflammatory (COX-1 inhibitors)^{69,70}, antineoplastic^{20,71}, and antioxidant⁶⁷ activities.

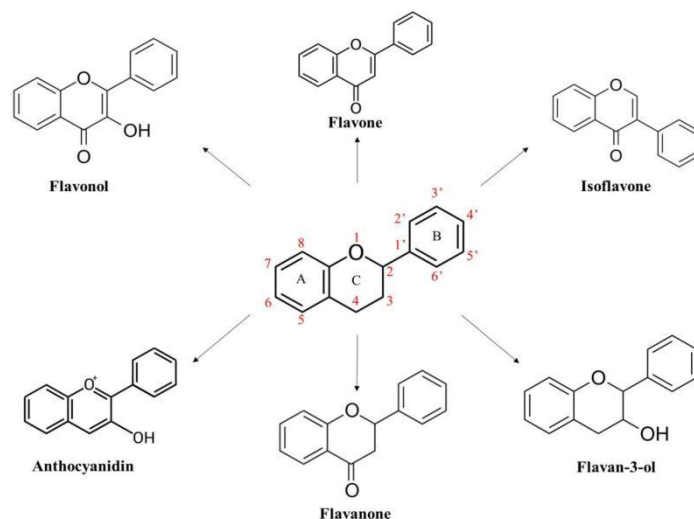
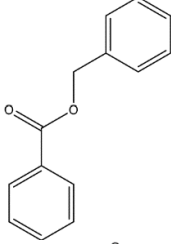
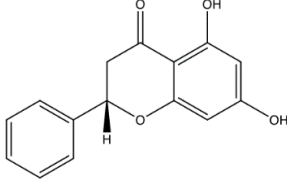
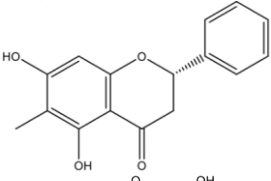
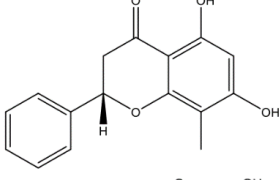
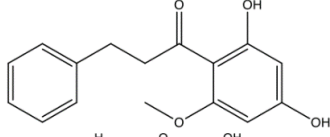
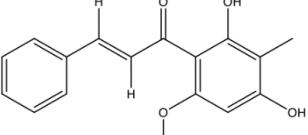
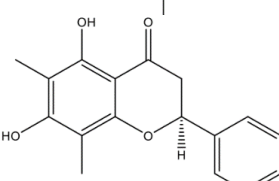
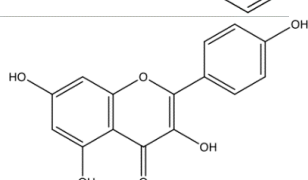
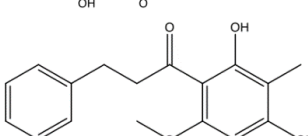
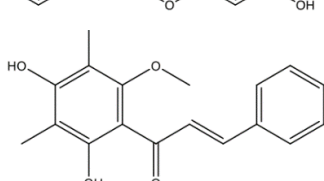
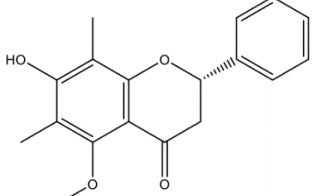
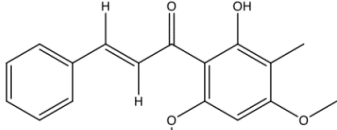
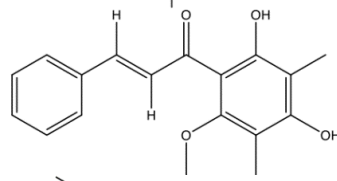
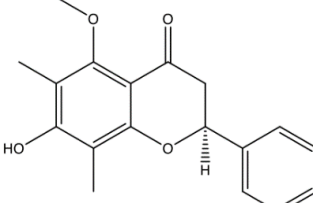
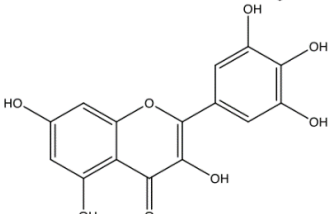
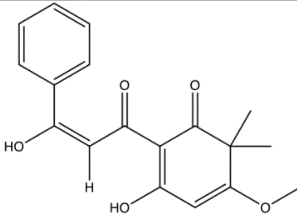
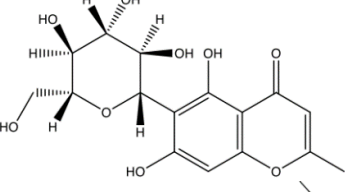
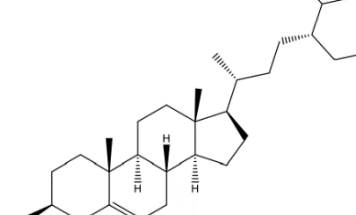


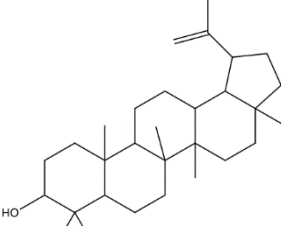
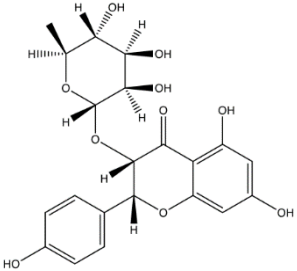
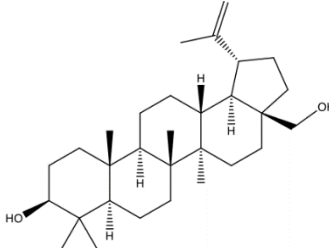
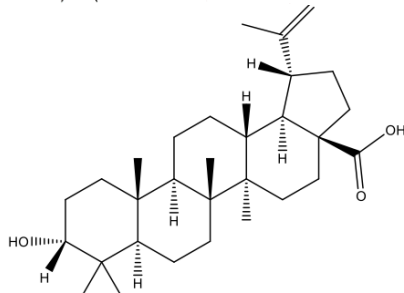
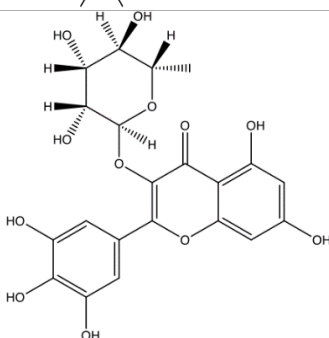
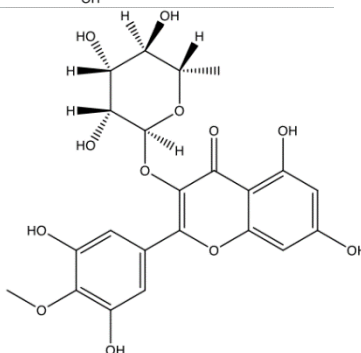
Figure 2: Basic structure of flavonoid and its derivatives

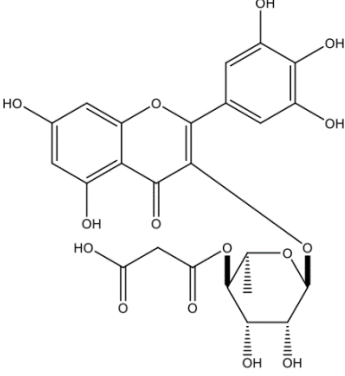
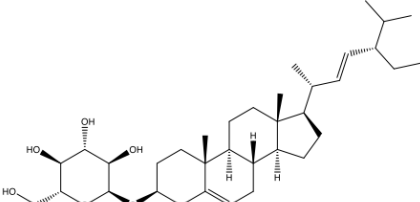
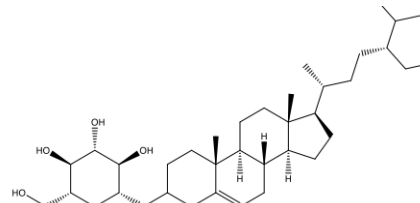
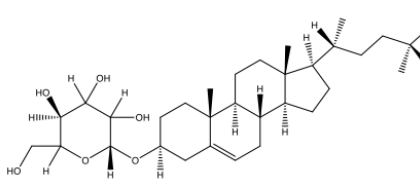
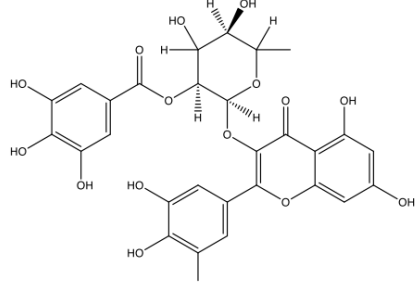
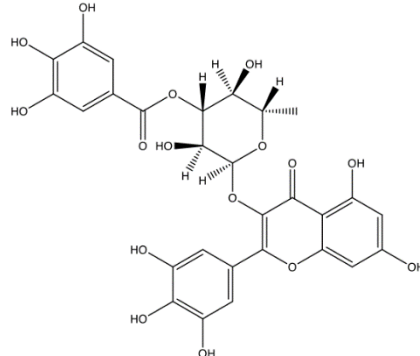
Table 1: Identified Compounds based on LC-MS Chromatogram of F2 Fraction (Dichloromethane Fraction)

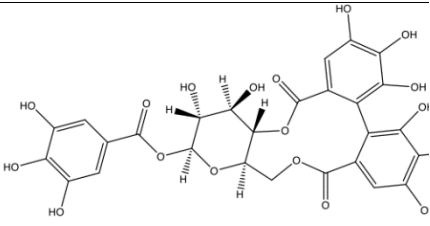
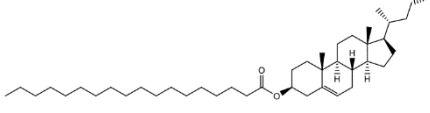
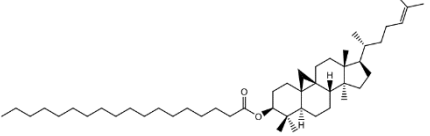
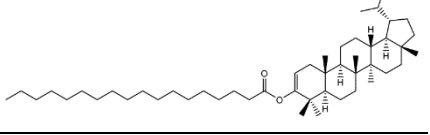
Comp. No.	RT (Min)	Composition (%)	Compound Results Analysis	Identified Compound
1	1.606	0.81312	Methyl salicylate CF: C ₈ H ₈ O ₃ ; EM: 152.0473; MW: 152.490; m/z: 152.0473 (100%), and 153.0507 (8.7%)	
2	2.694	0.40981	Eugenol CF: C ₁₀ H ₁₂ O ₂ ; EM: 164.0837; MW: 164.2040; m/z: 164.0837 (100%), and 165.0871 (10.8%)	
3	3.042	2.75891	Gallic acid CF: C ₇ H ₆ O ₅ ; EM: 170.0215; MW: 170.1200; m/z: 170.0215 (100%), 171.0249 (7.6%), and 172.0258 (1.0%)	
4	5.494	3.62398	β -Caryophyllene CF: C ₁₅ H ₂₄ ; EM: 204.1878 ; MW: 204.3570 ; m/z: 204.1878 (100%), and 205.1912 (1.2%)	
5	5.571	2.27671	Eugenin CF: C ₁₁ H ₁₀ O ₄ ; EM: 206.0579; MW: 206.1970; m/z: 206.0579 (100%), and 207.0613 (11.9%)	
6	5.587	3.79729	Eugenol acetate CF: C ₁₂ H ₁₄ O ₃ ; EM: 206.0943; MW: 206.2410; m/z: 206.0943 (100%), and 207.0976 (13.0%)	

7	6.408	2.30441	Benzyl benzoate CF: C ₁₄ H ₁₂ O ₂ ; EM: 212.0837; MW: 212.2480; m/z: 212.0837 (100%), 213.0871 (15.1%), and 213.0871 (1.1%)	
8	8.217	1.52868	Pinocebrin CF: C ₁₅ H ₁₂ O ₄ ; EM: 256.0736; MW: 256.2570; m/z: 256.0736 (100.0%), 257.0769 (16.2%), and 258.0803 (1.2%)	
9	9.349	2.06804	(-)-Strobopinin CF: C ₁₄ H ₁₄ O ₄ ; EM: 270.0892; MW: 270.2840; m/z: 270.0892 (100.0%), 271.0926 (17.3%), and 272.0959 (1.4%)	
10	9.375	1.73286	8-Methylpinocebrin CF: C ₁₆ H ₁₄ O ₄ ; EM: 270.0892; MW: 270.2840; m/z: 270.0892 (100.0%), 271.0926 (17.3%), and 272.0959 (1.4%)	
11	9.74	1.21678	Uvangoletin CF: C ₁₆ H ₁₆ O ₄ ; EM: 272.1049; MW: 272.3000; m/z: 272.1049 (100.0%), 273.1082 (17.3%), and 274.1116 (1.4%)	
12	10.02	1.50924	Stercurensin CF: C ₁₇ H ₁₆ O ₄ ; EM: 284.1049; MW: 284.3110; m/z: 284.1049 (100.0%), 285.1082 (18.4%), and 286.1116 (1.6%)	
13	10.03	1.73178	Demethoxymatteucinol CF: C ₁₇ H ₁₆ O ₄ ; EM: 284.1049; MW: 284.3110; m/z: 284.1049 (100.0%), 285.1082 (18.4%), and 286.1116 (1.6%)	
14	10.322	2.25465	Kaempferol CF: C ₁₅ H ₁₀ O ₆ ; EM: 286.0477; MW: 286.2390; m/z: 286.0477 (100.0%), 287.0511 (16.2%), 288.0520 (1.2%), and 288.0544 (1.2%)	
15	10.336	2.22455	2',4'-Dihydroxy-6'-methoxy-3'-methyl-dihydrochalcone CF: C ₁₇ H ₁₈ O ₄ ; EM: 286.1205; MW: 286.3270; m/z: 286.1205 (100.0%), 287.1239 (18.4%), and 288.1272 (1.6%)	
16	10.517	2.64109	4',6'-Dihydroxy-3',5'-dimethyl-2'-methoxychalcone CF: C ₁₈ H ₁₈ O ₄ ; EM: 298.1205; MW: 298.3380; m/z: 298.1205 (100.0%), 299.1239 (19.5%), and 300.1272 (1.8%)	

17	10.519	1.88630	7-Hydroxy-5-methoxy-6,8-dimethylflavanone CF: C ₁₈ H ₁₈ O ₄ ; EM: 298.1205 ; MW: 298.3380 m/z: 298.1205 (100.0%), 299.1239 (19.5%), and 300.1272 (1.8%)	
18	11.015	0.85809	Aurentiacin CF: C ₁₈ H ₁₈ O ₄ ; EM: 298.1205; MW: 298.3380; m/z: 298.1205 (100.0%), 299.1239 (19.5%), and 300.1272 (1.8%)	
19	11.017	1.66310	2',4'-Dihydroxy-6'-methoxy3',5'-dimethylchalcone CF: C ₁₈ H ₁₈ O ₄ ; EM: 298.1205; MW: 298.3380; m/z: 298.1205 (100.0%), 299.1239 (19.5%), and 300.1272 (1.8%)	
20	11.02	0.94841	(+)-6,8-Di-C-methylpinocembrin-5-methylether CF: C ₁₈ H ₁₈ O ₄ ; EM: 298.1205; MW: 298.3380; m/z: 298.1205 (100.0%), 299.1239 (19.5%), and 300.1272 (1.8%)	
21	11.514	2.04365	Myricetin CF: C ₁₅ H ₁₀ O ₈ ; EM: 318.0376; MW: 318.2370; m/z: 318.0376 (100.0%), 319.0409 (16.2%), 320.0418 (1.6%), and 320.0443 (1.20%)	
22	11.915	1.21666	Syzygiol CF: C ₁₈ H ₁₈ O ₅ ; EM: 314.1154 ; MW: 314.3370 m/z: 314.1154 (100.0%), 315.1188 (19.5%), 316.1221 (1.8%), and 316.1197 (1.0%)	
23	12.417	0.96577	Biflorin CF: C ₁₆ H ₁₈ O ₉ ; EM: 354.0951 ; MW: 354.3110; m/z: 354.0951 (100.0%), 355.0984 (17.3%), 356.0993 (1.8%), and 356.1018 (1.4%)	
24	17.163	4.85456	β -Sitosterol CF: C ₂₉ H ₅₀ O ; EM: 414.3862 ; MW: 414.7180 m/z: 414.3862 (100.0%), 415.3895 (31.4%), 416.3929 (2.7%), and 416.3929 (2.0%)	

25	19.614	7.45610	<p>Lupeol CF: C₃₀H₅₀O ; EM: 426.3862 ; MW: 426.7290 m/z: 426.3862 (100.0%), 427.3895 (32.4%), 428.3929 (2.7%), and 428.3929 (2.4%)</p>	
26	21.458	1.28346	<p>Isoengeletin CF: C₂₁H₂₂O₁₀ ; EM: 434.1213 ; MW: 434.3970 m/z: 434.1213 (100.0%), 435.1247 (22.7%), 436.1280 (2.5%), and 436.1255 (2.1%)</p>	
27	22.284	1.03783	<p>Betulin CF: C₃₀H₅₀O₂ ; EM: 442.3811 ; MW: 442.7280 m/z: 442.3811 (100.0%), 443.3844 (32.4%), 444.3878 (2.7%), and 444.3878 (2.4%)</p>	
28	23.448	1.05291	<p>Epibetulinic acid CF: C₃₀H₄₈O₃ ; EM: 456.3603 ; MW: 456.7110 m/z: 456.3603 (100.0%), 457.3637 (32.4%), 458.3671 (2.7%), and 458.3671 (2.4%)</p>	
29	24.119	2.69024	<p>Myricitrin CF: C₂₁H₂₀O₁₂; EM: 464.0955; MW: 464.3790; m/z: 464.0955 (100.0%), 465.0988 (22.7%), 466.0997 (2.5%), and 466.1022 (2.5%)</p>	
30	25.839	2.42486	<p>Mearnsitrin CF: C₂₂H₂₂O₁₂; EM: 478.1111; MW: 478.4060; m/z: 478.1111 (100.0%), 479.1145 (23.8%), 480.1178 (2.7%), and 480.1154 (2.5%)</p>	

31	31.816	2.52852	<p>Myricetin-3-<i>O</i>-(4''-<i>O</i>-malonyl)-α-L-rhamnopyranoside CF: C₂₄H₂₂O₁₅ ; EM: 550.0959 ; MW: 550.4250 m/z: 550.0959 (100.0%), 551.0992 (26.0%), 552.1026 (3.2%), and 552.1001 (3.1%)</p>	
32	33.422	4.05880	<p>Stigmasterol-3-<i>O</i>-β-D-glucoside CF: C₃₅H₅₈O₆ ; EM: 574.4233 ; MW: 574.8430 m/z: 574.4233 (100.0%), 575.4267 (37.9%), 576.4300 (4.3%), 576.4300 (2.7%), and 576.4276 (1.2%)</p>	
33	33.428	4.80307	<p>β-Sitosterol-D-glucoside CF: C₃₅H₆₀O₆ ; EM: 576.4390 ; MW: 576.8590 m/z: 576.4390 (100.0%), 577.4423 (37.9%), 578.4457 (4.3%), 578.4457 (2.7%), and 578.4432 (1.2%)</p>	
34	33.013	2.86462	<p>Campesterol glucoside CF: C₃₄H₅₈O₆; EM: 562.4233 ; MW: 562.8320 m/z: 562.4233 (100.0%), 563.4267 (36.8%), 564.4300 (6.6%), and 564.4276 (1.2%)</p>	
35	35.646	2.83992	<p>Desmanthin 1 CF: C₂₈H₂₄O₁₆; EM: 616.1064; MW: 616.4840; m/z: 616.1064 (100.0%), 617.1098 (30.3%), 618.1131 (4.4%), and 618.1107 (3.3%)</p>	
36	35.649	3.00663	<p>Myricetin-3-(3''-galloylrhamnoside) CF: C₂₈H₂₄O₁₆ ; EM: 616.1064 ; MW: 616.4840 m/z: 616.1064 (100.0%), 617.1098 (30.3%), 618.1131 (4.4%), and 618.1107 (3.3%)</p>	

37	37.022	1.48540	Strictinin CF: C ₂₇ H ₂₂ O ₁₈ ; EM: 634.0806 ; MW: 634.4550 m/z: 634.0806 (100.0%), 635.0840 (29.2%), 636.0873 (4.1%), 636.0849 (3.7%), and 637.0882 (1.1%)	
38	46.203	4.09850	Sitosteryl stearate CF: C ₄₇ H ₈₄ O ₂ ; EM: 680.6471 ; MW: 681.1870 m/z: 680.6471 (100.0%), 681.6505 (50.8%), 682.6538 (12.6%), and 683.6572 (1.2%)	
39	46.213	6.54282	Cycloartenyl stearate CF: C ₄₈ H ₈₄ O ₂ ; EM: 692.6471 ; MW: 693.1980 m/z: 692.6471 (100.0%), 693.6505 (51.9%), 694.6538 (13.2%), and 695.6572 (1.4%)	
40	46.215	4.49788	Lupenyl stearate CF: C ₄₈ H ₈₄ O ₂ ; EM: 692.6471 ; MW: 693.1980 m/z: 692.6471 (100.0%), 693.6505 (51.9%), 694.6538 (13.2%), and 695.6572 (1.4%)	

Note = CF: Chemical Formula; EM: Exact Mass; MW: Molecular Weight; m/z: mass per charge

Beyond the chalcone and flavanone groups, other groups of polyphenols identified in F2 include the flavonol group, notably compound 30 (mearncitrin) and its derivatives featuring two hydroxyl groups, commonly referred to as dihydroflavonols, such as compound 26 (isoengeletin). Compound 30 (mearncitrin) is a glycoside derived from mearncitrin substituted by the α -L-rhamnopyranosyl. This compound has been isolated from *Byrsonima coccolobifolia*⁷² and *Myrsine africana*⁷³, known as natural antioxidants from plants⁷⁴. Meanwhile, compound 26 (isoengeletin) is a dihydroflavonol derivative substituted by the α -L-rhamnopyranosyl at the third position. This compound can be found in *Smilax glabrae*⁷⁵ and *Iryanthera lancifolia*⁷⁶ and is being researched for psoriasis, hyperuricemia, and gout.⁷⁷

Terpenoids are a group of compounds commonly identified as oils derived from 5-carbon isoprene, namely monoterpenes, diterpenes, triterpenes, etc. The isoprene structure as the basic structure for the terpenoid group is presented in Figure 6.⁷⁸ In general, terpenoids are compounds with different bioactivity in the pharmaceutical field, especially in medicinal chemistry.⁷⁹ In this fraction, compounds belonging to the terpenoids are compounds 25, 27, 28, and 40. These compounds can be grouped into triterpene-type terpenoids because of the presence of 3 basic structures of isoprenes. Compound 25 (lupeol) is a pentacyclic triterpenoid with anticancer^{80,81} and anti-inflammatory activities.⁸⁰ This compound can be found in mango⁸², *Camellia japonica*⁸³, *Acacia visco*⁸⁴, and *Abronia villosa*.⁸⁵ Compound 27 (betulin) is a triterpene commonly isolated from birch trees.⁸⁶ This compound is a lupeol derivative with hydroxyl groups attached at C-3 and C-28 known to have anticancer activity.⁸⁷ Compound 28 (epibetulinic acid) is a derivative of betulinic acid (also a lupeol derivative) found in *Microtropis fokienensis*⁸⁸, *Hypericum lancifolium*⁸⁹, and birch trees.⁹⁰ Epibetulinic acid has anti-inflammatory, antineoplastic⁹¹, and anti-HIV⁹² activities. Compound 40 (lupenyl stearate) is a lupeol derivative substituted with a stearyl group at the β -position, commonly found in *S. Samarangense*.⁹³

Steroids are compounds with a basic structure of seventeen C atoms bonded in four rings (with three cyclohexane rings and one cyclopentane ring).^{94,95} The compounds within this group are steroid derivatives with a hydroxyl group, namely sterols⁹⁶. Sterols found in plants are called phytosterols, while the ones found in animals are called zoosterols. These derivative compounds have many uses, such as inhibiting human cholesterol absorption. Compounds classified as phytosterols include compounds 24, 32, 33, 34, 38, and 39. Compound 24 (β -sitosterol) is a sterol derivative substituted with a β -hydroxyl

group at the third position. This compound can be found in *Typhonium trilobatum*⁹⁷ and *Elodea canadensis*⁹⁸, used as an antioxidant⁹⁹, anticholesteremic drug¹⁰⁰, and reduces the toxic effects of radiation.⁹⁷

Compound 32 (stigmasterol-3-O- β -D-glucoside) is a glycoside compound derived from stigmasterol, which is substituted with β -D-glucopyranosyl at position 3 with glycosidic bonds, which are also classified as steroid saponins. This compound is often found in *Symplocos lancifolia*, which is used as an antioxidant and antibacterial.¹⁰¹ Compound 33 (β -sitosterol-D-glucoside) is a glycoside compound derived from sitosterol, which is substituted with D-glucopyranosyl found in *Dimocarpus longan*¹⁰² and *Erigeron canadensis*¹⁰³ having antioxidant¹⁰⁴ and anti-inflammatory properties.⁴⁸ Compound 38 (sitosteril stearate) is a sterol compound derived from sitosterol, substituted by a stearyl group at the β -position. This compound is commonly found in *S. samarangense*, which has antioxidant and anti-inflammatory properties.⁹³ Compound 39 (cycloartenyl stearate) is a cycloartenol derivative compound substituted with a stearyl group found in *S. samarangense*, with analgesic and anti-inflammatory activities.⁹³

In addition to the main compound groups described above, the ester and quinone groups were detected in this fraction. The ester group found in this fraction is compound 7 (benzyl benzoate), a benzoic acid derivative. This compound is easily found and isolated from the genus *Polyalthia*¹⁰⁵ plants and *Populus tremula*¹⁰⁶ as a scabicide and acaricide.¹⁰⁷ Meanwhile, the quinone group found in this fraction is compound 23 (biflorin), belonging to the naphthoquinone group. This compound is found in *S. aromaticum*¹⁰⁸ and *Capraria biflora*¹⁰⁹ and has antimicrobial, antitumor, and antimutagenic activities.¹⁰⁹

Conclusion

The findings of this study indicated that the dichloromethane fraction of methanol extract of the stem bark of *S. samarangense* contains 40 phytoconstituents categorised into phenolics, flavonoids, terpenoids, and steroids, each with distinct bioactivities. This information is insightful for further research and understanding of the properties of these compounds. Additional research on isolating compounds from F2 is necessary to identify lead compounds for the development of pharmaceutical products.

Conflict of Interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

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References

- Abdulrahman MD, Hama HA. Anticancer of genus *Syzygium*: a systematic review. *Explor Target Antitumor Ther.* 2023; 4(2): 273–293.
- Idris NS, Khandaker MM, Rashid ZM, Majrashi A, Alenazi MM, Nor ZM, Mohd Adnan AF, Mat N. Polyphenolic compounds and biological activities of leaves and fruits of *Syzygium samarangense* cv. "Giant Green" at three different maturities. *Horticulturae.* 2023; 9(3): 326.
- Srivastava R, Shaw AK, Kulshreshtha DK. Triterpenoids and chalcone from *Syzygium samarangense*. *Phytochem.* 1995; 38(3): 687–689.
- Aung EE, Kristanti AN, Aminah NS, Takaya Y, Ramadhan R. Plant description, phytochemical constituents and bioactivities of *Syzygium* genus: A review. *Open Chem.* 2020; 18(1): 1256–1281.
- Tarigan C, Pramastya H, Insanu M, Fidrianny I. *Syzygium samarangense*: review of phytochemical compounds and pharmacological activities. *Biointerface Res Appl Chem.* 2022; 12(2): 2084–2107.
- Mollika S, Islam N, Parvin N, Kabir A, Sayem MW, Luthfunnesa SR. Evaluation of analgesic, anti-inflammatory and CNS activities of the methanolic extract of *Syzygium samarangense* leave. *Glob J Pharmacol.* 2014; 8(1): 39-46.
- Tukiran T, Setiawan AR, Constaty IC, Safitri FN. The potency of Java Apple (*Syzygium samarangense*) as α -glucosidase and α -amylase inhibitor: an in-silico approach. *Trop J Nat Prod Res.* 2023; 7(8): 3741–3755.
- Madhavi M, Ram MR. Phytochemical screening and evaluation of biological activity of root extracts of *Syzygium samarangense*. *Int J Res Pharm Chem.* 2015; 5(4): 753–763.
- Metasari S, Elfita E, Muharni M, Yohandini H. Antioxidant compounds from the stem bark of *Syzygium samarangense* L. *Molekul.* 2020; 15(3): 175–183.
- Constanty IC, Tukiran T. Aktivitas antioksidan dari fraksi n-heksana kulit batang tumbuhan jambu semarang (*Syzygium samarangense*). *J Kim Ris.* 2021; 6(1): 1–7.
- Tukiran T, Suyatno S, Safitri FN. Identification of the chemical constituents of the selected fraction of the dichloromethane extract of *Syzygium samarangense* stem bark using LC-ESI-MS and evaluation its potential as antifungal agent. *Indones J Chem.* 2021; 21(2): 340–349.
- Sururi AM, Raihan M, Aisa ER, Safitri FN, Constaty IC. Anti-inflammatory activity of stem bark dichloromethane fraction of *Syzygium samarangense* extract as COX-2 inhibitor: a bioinformatics approach. *J Kim Ris.* 2022; 7(2): 94–100.
- Marzouk MM, Elkhateeb A, El-Shabrawy M, Farid MM, Kawashty SA, Abdel Hameed ESS, Hussein SR. Chemical profiling of *Farsetia aegyptia* Turra and *Farsetia longisiliqua* Decne and their chemosystematic significance. *Trop J Nat Prod Res.* 2020; 4(11): 953–960.
- Elias TS, Dykeman PA. Edible wild plants: a North American field guide to over 200 natural foods. Sterling Publishing Company, Inc.; 2009.
- Tramèr MR. It's not just about rubbing—topical capsaicin and topical salicylates may be useful as adjuvants to conventional pain treatment. *BMJ.* 2004; 328(7446): 998.
- Mason L, Moore RA, Edwards JE, McQuay HJ, Derry S, Wiffen PJ. Systematic review of efficacy of topical rubefaciants containing salicylates for the treatment of acute and chronic pain. *BMJ.* 2004; 328(7446): 995.
- Haslam E, Cai Y. Plant polyphenols (vegetable tannins): gallic acid metabolism. *Nat Prod Rep.* 1994; 11: 41–66.
- Tukiran, Putri DO. Chemical profile by LC-MS analysis from the selected fraction of methanol extract of *Syzygium malaccense*. *Rasayan J Chem.* 2021; 14(1): 295–305.
- Limanto A, Simamora A, Santoso AW, Timotius KH. Antioxidant, α -glucosidase inhibitory activity, and molecular docking study of gallic acid, quercetin and rutin: a comparative study. *Mol Cell Biomed Sci.* 2019; 3(2): 67–74.
- Varela-Rodríguez L, Sánchez-Ramírez B, Hernández-Ramírez VI, Varela-Rodríguez H, Castellanos-Mijangos RD, González-Horta C, Chávez-Munguía B, Talamás-Rohana P. Effect of gallic acid and myricetin on ovarian cancer models: a possible alternative antitumoral treatment. *BMC Complement Med Ther.* 2020; 20(1): 110.
- Budavari S, O'Neil MJ, Smith A, Heckelman PE, Kinneary JF. The Merck index, an encyclopedia of chemicals, drugs, and biologicals, Whitehouse Station, NY: Merck Research Laboratories Division of Merck & Co. Inc Monogr. 1996; 2006: 289.
- Anish RJ, Mohanan B, Aswathy TR, Nair A, Radhakrishnan KV, Rauf AA. An integrated approach to the structural characterization, long-term toxicological and anti-inflammatory evaluation of *Pterospermum rubiginosum* bark extract. *J Ethnopharmacol.* 2023; 308: 116262.
- Klausmeyer P, Zhou Q, Scudiero DA, Uranchimeg B, Melillo G, Cardellina JH, Shoemaker RH, Chang CJ, McCloud TG. Cytotoxic and HIF-1 α inhibitory compounds from *Crossosoma bigelovii*. *J Nat Prod.* 2009; 72(5): 805–812.
- Czepa A, Hofmann T. Structural and sensory characterization of compounds contributing to the bitter off-taste of carrots (*Daucus carota* L.) and carrot puree. *J Agric Food Chem.* 2003; 51(13): 3865–3873.
- Arapitsas P. Hydrolyzable tannin analysis in food. *Food Chem.* 2012; 135(3): 1708–1717.
- Shimozu Y, Kuroda T, Tsuchiya T, Hatano T. Structures and antibacterial properties of isorugosins H–J, oligomeric ellagitannins from *Liquidambar formosana* with characteristic bridging groups between sugar moieties. *J Nat Prod.* 2017; 80(10): 2723–2733.
- Jiang ZH, Tanaka T, Iwata H, Sakamoto S, Hirose Y, Kouno I. Ellagitannins and lignan glycosides from *Balanophora japonica* (Balanophoraceae). *Chem Pharm Bull.* 2005; 53(3): 339–341.
- Barros J, Serrani-Yarce JC, Chen F, Baxter D, Venables BJ, Dixon RA. Role of bifunctional ammonia-lyase in grass cell wall biosynthesis. *Nat Plants.* 2016; 2(6): 1–9.
- Pop DM, Timar MC, Varodi AM, Beldean EC. An evaluation of clove (*Eugenia caryophyllata*) essential oil as a potential alternative antifungal wood protection system for cultural heritage conservation. *Maderas, Cienc. Tecnol.* 2022; 24.
- Haro-González JN, Castillo-Herrera GA, Martínez-Velázquez M, Espinosa-Andrews H. Clove essential oil (*Syzygium aromaticum* L. Myrtaceae): extraction, chemical composition, food applications, and essential bioactivity for human health. *Molecules.* 2021; 26(6387): 1-25.
- Marchese A, Barbieri R, Coppo E, Orhan IE, Daglia M, Nabavi SF, Izadi M, Abdollahi M, Nabavi SM, Ajami M. Antimicrobial activity of eugenol and essential oils containing eugenol: a mechanistic viewpoint. *Crit Rev Microbiol.* 2017; 43(6): 668–689.

32. Sururi AM, Maharani DK, Wati FA. Potensi senyawa eugenol dari cengkeh (*Syzygium aromaticum*) sebagai inhibitor protease HIV-1 (PR). *Unesa J Chem*. 2023; 12(1): 26–30.
33. De Souza Farias SA, Da Costa KS, Martins JBL. Analysis of conformational, structural, magnetic, and electronic properties related to antioxidant activity: revisiting flavan, anthocyanidin, flavanone, flavonol, isoflavone, flavone, and flavan-3-ol. *ACS Omega*. 2021; 6(13):8908–8918.
34. Piccolella S, Pacifico S. Plant-derived polyphenols: a chemopreventive and chemoprotectant worth-exploring resource in toxicology. *Adv Mol Toxicol*. 2015; 161–214.
35. Shen J, Zhu X, Wu Z, Shi Y, Wen T. Uvangoletin, extracted from *Sarcandra glabra*, exerts anticancer activity by inducing autophagy and apoptosis and inhibiting invasion and migration on hepatocellular carcinoma cells. *Phytomedicine*. 2022; 94: 153793.
36. Maeda G, Van Der Wal J, Gupta AK, Munissi JJE, Orthaber A, Sunnerhagen P, Nyandoro SS, Erdélyi M. Oxygenated cyclohexene derivatives and other constituents from the roots of *Monanthes trichocarpa*. *J Nat Prod*. 2020; 83(2): 210–215.
37. Dao TT, Tung BT, Nguyen PH, Thuong PT, Yoo SS, Kim EH, Kim SK, Oh WK. C-methylated flavonoids from *Cleistocalyx operculatus* and their inhibitory effects on novel influenza (H1N1) neuraminidase. *J Nat Prod*. 2010; 73(10): 1636–1642.
38. Wollenweber E, Kohorst G, Mann K, Bell JM. Leaf gland flavonoids in *Comptonia peregrina* and *Myrica pensylvanica* (Myricaceae). *J Plant Physiol*. 1985; 117(5): 423–430.
39. Huh J, Ha TKQ, Kang KB, Kim KH, Oh WK, Kim J, Sung SH. C-methylated flavonoid glycosides from *Pentarrhizidium orientale* rhizomes and their inhibitory effects on the H1N1 influenza virus. *J Nat Prod*. 2017; 80(10): 2818–2824.
40. Mathiesen L, Malterud KE, Sund RB. Antioxidant activity of fruit exudate and C-methylated dihydrochalcones from *Myrica gale*. *Planta Med*. 1995; 61(06): 515–518.
41. Perera ND, Tomas D, Wanniarachchillage N, Cuic B, Luikinga SJ, Rytova V, Turner BJ. Stimulation of mTOR-independent autophagy and mitophagy by rilmenidine exacerbates the phenotype of transgenic TDP-43 mice. *Neurobiol Dis*. 2021; 154: 105359.
42. Utama K, Khamto N, Meepowpan P, Aobchey P, Kantapan J, Sringarm K, Roytrakul S, Sangthong P. Effects of 2',4'-dihydroxy-6'-methoxy-3',5'-dimethylchalcone from *Syzygium nervosum* seeds on antiproliferative, DNA damage, cell cycle arrest, and apoptosis in human cervical cancer cell lines. *Molecules*. 2022; 27(4): 1154.
43. Kim YJ, Kim HC, Ko H, Amor EC, Lee JW, Yang HO. Inhibitory effects of aurentiacin from *Syzygium samarangense* on lipopolysaccharide-induced inflammatory response in mouse macrophages. *Food Chem Toxicol*. 2012; 50(3–4): 1027–1035.
44. Janpajit S, Sillapachaiyaporn C, Theerasri A, Charoenkiatkul S, Sukprasansap M, Tencommao T. *Cleistocalyx nervosum* var. Paniala Berry seed protects against TNF- α -stimulated neuroinflammation by inducing HO-1 and suppressing NF- κ B mechanism in BV-2 microglial cells. *Molecules*. 2023; 28(7): 3057.
45. Nishizawa M, Yamada H, Sano J, Itô S, Hayashi Y, Ikeda H, Chairul, Shiro M, Tokuda H. Structure of syzygiol: a skin-tumor promotion inhibitor. *Tetrahedron Lett*. 1991; 32(2): 211–212.
46. Lim TK. *Syzygium samarangense* BT - edible medicinal and non medicinal plants. Volume 3, Fruits. In: Lim TK, editor. Dordrecht: Springer Netherlands. 2012; 778–786.
47. Martínez A, Conde E, Moure A, Domínguez H, Estévez RJ. Protective effect against oxygen reactive species and skin fibroblast stimulation of *Couroupita gutanensis* leaf extracts. *Nat Prod Res*. 2012; 26(4): 314–322.
48. Pan L, Matthew S, Lantvit DD, Zhang X, Ninh TN, Chai H, Carcache De Blanco EJ, Soejarto DD, Swanson SM, Kinghorn AD. Bioassay-guided isolation of constituents of *Piper sarmentosum* using a mitochondrial transmembrane potential assay. *J Nat Prod*. 2011; 74(10): 2193–2199.
49. Allard PM, Dau ETH, Eydoux C, Guillemot JC, Dumontet V, Poullain C, Canard B, Guéritte F, Litaudon M. Alkylated flavanones from the bark of *Cryptocarya chartacea* as dengue virus NS5 polymerase inhibitors. *J Nat Prod*. 2011; 74(11): 2446–2453.
50. Li X, Zhai Y, Xi B, Ma W, Zhang J, Ma X, Miao Y, Zhao Y, Ning W, Zhou H, Yang C. Pinocebrin ameliorates skin fibrosis via inhibiting tgf- β 1 signaling pathway. *Biomolecules*. 2021; 11(8):1240.
51. Sharma N, Sharma M, Shakeel E, Jamal QMS, Kamal MA, Sayeed U, Khan MKA, Siddiqui MH, Arif JM, Akhtar S. Molecular interaction and computational analytical studies of pinocebrin for its antiangiogenic potential targeting VEGFR-2: a persuader of metastasis. *Med Chem (Los Angeles)*. 2018; 14(6): 626–640.
52. Kumar N, Shrungeswara AH, Mallik SB, Biswas S, Mathew J, Nandakumar K, Mathew J, Lobo R. Pinocebrin-enriched fractions of *Elytranthe parasitica* (L.) Danser modulates apoptotic and MAPK cellular signaling in HepG2 cells. *Anticancer Agents Med Chem*. 2018; 18(11): 1563–1572.
53. Jiang L, Yang Y, Feng H, Zhou Q, Liu Y. Pinocebrin inhibits the proliferation, migration, invasiveness, and epithelial-mesenchymal transition of colorectal cancer cells by regulating LACTB. *Cancer Biother Radiopharm*. 2022; 37(7): 527–536.
54. Yang R, Yang J, Li Z, Su R, Zou L, Li L, Xu X, Li G, Liu S, Liang S, Xu C. Pinocebrin inhibits P2X4 receptor-mediated pyroptosis in hippocampus to alleviate the behaviours of chronic pain and depression comorbidity in rats. *Mol Neurobiol*. 2022; 59(12): 7119–7133.
55. Thring TS, Hili P, Naughton DP. Antioxidant and potential anti-inflammatory activity of extracts and formulations of white tea, rose, and witch hazel on primary human dermal fibroblast cells. *J Inflamm*. 2011; 8(1): 1–7.
56. Lucarini M, Durazzo A, Di Stefano V, Di Lena G, Lombardi-Boccia G, Santini A. Bioactive phytochemicals from grape seed oil processing by-products BT - bioactive phytochemicals from vegetable oil and oilseed processing by-products. In: Ramadan Hassanien MF, editor. Cham: Springer International Publishing; 2023; 289–308.
57. Kovalev S V. Flavonoids from *Lotus ucrainicus* and *L. arvensis*. *Chem Nat Compd*. 2009; 45: 550–551.
58. Sharma V, Joseph C, Ghosh S, Agarwal A, Mishra MK, Sen E. Kaempferol induces apoptosis in glioblastoma cells through oxidative stress. *Mol Cancer Ther*. 2007; 6(9): 2544–2553.
59. Zhang Y, Chen AY, Li M, Chen C, Yao Q. *Ginkgo biloba* extract kaempferol inhibits cell proliferation and induces apoptosis in pancreatic cancer cells. *J Surg Res*. 2008; 148(1): 17–23.
60. Sadeghi-Kiakhani M, Hashemi E, Miri FS, Tehrani-Bagha AR, Etezad SM. Dyeing of nylon fabric with two natural dyes, saffron (*Crocus sativus* L.) and weld (*Reseda luteola* L.), and study their dyeing, antioxidant, and antibacterial properties. *Fibers Polym*. 2023; 24(3): 1083–1092.
61. Huang Q, Gao B, Wang L, Hu YQ, Lu WG, Yang L, Luo ZJ, Liu J. Protective effects of myricitrin against osteoporosis via reducing reactive oxygen species and bone-resorbing cytokines. *Toxicol Appl Pharmacol*. 2014; 280(3): 550–560.
62. Zou J, Mi Y, Chen W, Liu Q, Wang J, Lou L, Zhao W. Alkyl phloroglucinol derivatives from *Syzygium levinei* and their differentiation-inducing activity. *Planta Med*. 2006; 72(06): 533–538.
63. Ye G, Huang C. Flavonoids of *Limonium aureum*. *Chem Nat Compd*. 2006; 42: 232–234.

64. Liu Y, Shang RF, Cheng FS, Wang XH, Hao BC, Liang JP. Flavonoids and phenolics from the flowers of *Limonium aureum*. Chem Nat Compd. 2016; 52: 130–131.
65. Acharyya RN, Mithila S, Rani S, Islam MA, Golder M, Ahmed KS, Hossain H, Dev S, Das AK. Antiallergic and anti-hyperglycemic potentials of *Lumnitzera racemosa* leaves: in vivo and in silico studies. Proc Natl Acad Sci India Sect B - Biol Sci. 2023; 93(1): 147–158.
66. Li YR, Li GH, Zhou MX, Xiang L, Ren DM, Lou HX, Wang XN, Shen T. Discovery of natural flavonoids as activators of Nrf2-mediated defense system: structure-activity relationship and inhibition of intracellular oxidative insults. Bioorganic Med Chem. 2018; 26(18): 5140–5150.
67. Liu Y, Wang R, Ren C, Pan Y, Li J, Zhao X, Xu C, Chen K, Li X, Gao Z. Two myricetin-derived flavonols from *Morella rubra* leaves as potent α -glucosidase inhibitors and structure-activity relationship study by computational chemistry. Oxid Med Cell Longev. 2022; 2022: 9012943.
68. Mohamed YSN, Abas F, Jaafar AH, Azizan A, Zolkeflee NKZ, Abd Ghafar SZ. Antioxidant and α -glucosidase inhibitory activities of eight neglected fruit extracts and UHPLC-MS/MS profile of the active extracts. Food Sci Biotechnol. 2021; 30(2): 195–208.
69. Song X, Tan L, Wang M, Ren C, Guo C, Yang B, Ren Y, Cao Z, Li Y, Pei J. Myricetin: a review of the most recent research. Biomed Pharmacother. 2021; 134: 111017.
70. Rahmani AH, Almatroudi A, Allemailem KS, Alwanian WM, Alharbi BF, Alrumaihi F, Khan AA, Almatroodi SA. Myricetin: a significant emphasis on its anticancer potential via the modulation of inflammation and signal transduction pathways. Int J Mol Sci. 2023; 24(11): 9665.
71. Akash S, Kumer A, Rahman MM, Emran T Bin, Sharma R, Singla RK, Alhumaydhi FA, Khandaker MU, Park MN, Idris AM. Development of new bioactive molecules to treat breast and lung cancer with natural myricetin and its derivatives: a computational and SAR approach. Front Cell Infect Microbiol. 2022; 12: 952297.
72. De Sousa LRF, Ramalho SD, Burger MCDM, Nebo L, Fernandes JB, Da Silva MFDGF, Iemma MRDC, Correia CJ, Souza DHF De, Lima MIS, Vieira PC. Isolation of arginase inhibitors from the bioactivity-guided fractionation of *Byrsonima coccolobifolia* leaves and stems. J Nat Prod. 2014; 77(2): 392–396.
73. Kishore N, Twilley D, Blom Van Staden A, Verma P, Singh B, Cardinali G, Kovacs D, Picardo M, Kumar V, Lall N. Isolation of flavonoids and flavonoid glycosides from *Myrsine africana* and their inhibitory activities against mushroom tyrosinase. J Nat Prod. 2018; 81(1): 49–56.
74. Op de Beck P, Cartier G, David B, Dijoux-Franca M, Mariotte A. Antioxidant flavonoids and phenolic acids from leaves of *Leea guineense* G. Don (Leeaceae). Phyther Res An Int J Devoted to Pharmacol Toxicol Eval Nat Prod Deriv. 2003; 17(4): 345-347.
75. Fukunaga T, Miura T, Furuta K, Kato A. Hypoglycemic effect of the rhizomes of *Smilax glabra* in normal and diabetic mice. Biol Pharm Bull. 1997; 20(1): 44–46.
76. Silva DHS, Yoshida M, Kato MJ. Flavonoids from *Iryanthera sagotiana*. Phytochem. 1997; 46(3): 579–582.
77. Zhang QF, Guo YX, Zheng G, Wang WJ. Chemical constituents comparison between *Rhizoma smilacis* Glabrae and *Rhizoma smilacis* Chinae by HPLC-DAD-MS/MS. Nat Prod Res. 2013; 27(3): 277–281.
78. Christianson DW. Structural and chemical biology of terpenoid cyclases. Chem Rev. 2017; 117(17): 11570–11648.
79. Ashour M, Wink M, Gershenzon J. Biochemistry of terpenoids: monoterpenes, sesquiterpenes and diterpenes. Annu. Rev. Plant Biol. 2010; 40: 258–303.
80. Vigneshwaran S, Maharani K, Sivasakthi P, Selvan PS, Saraswathy SD, Priya ES. Bioactive fraction of *Tragia involucrata* Linn leaves attenuates inflammation in Freund's complete adjuvant-induced arthritis in wistar albino rats via inhibiting NF- κ B. Inflammopharmacology. 2023; 31(2): 967–981.
81. Ren QL, Wang Q, Zhang XQ, Wang M, Hu H, Tang JJ, Yang XT, Ran YH, Liu HH, Song ZX, Liu JG, Li XL. Anticancer activity of diosgenin and its molecular mechanism. Chin J Integr Med. 2023; 29(8): 738-749.
82. Lerma-Torres JM, Navarro-Ocaña A, Calderón-Santoyo M, Hernández-Vázquez L, Ruiz-Montañez G, Ragazzo-Sánchez JA. Preparative scale extraction of mangiferin and lupeol from mango (*Mangifera indica* L.) leaves and bark by different extraction methods. J Food Sci Technol. 2019; 56(10): 4625-4631.
83. Majumder S, Ghosh A, Bhattacharya M. Natural anti-inflammatory terpenoids in *Camellia japonica* leaf and probable biosynthesis pathways of the metabolome. Bull Nat Res Cent. 2020; 44(1): 1–14.
84. Pometti C, Camps GA, Soldati MC, Sastre TV, Gavier G, Zelener N, Verga A, Ewens M, Saidman BO, Sérsic AN, Cosacov A. Species without current breeding relevance but high economic value: *Acacia caven*, *Acacia aroma*, *Acacia visco*, *Prosopis affinis*, *Prosopis caldenia* and *Gonopterodendron sarmientoi*. In: Low-Intensity Breeding of Native Forest Trees in Argentina: Genetic Basis for their Domestication and Conservation. 2020.
85. Starks CM, Williams RB, Norman VL, Lawrence JA, Goering MG, O'Neil-Johnson M, Hu JF, Rice SM, Eldridge GR. Abronione, a rotenoid from the desert annual *Abronia villosa*. Phytochem Lett. 2011; 4(2): 72–74.
86. Demets O V., Takibayeva A, Kassenov R, Aliyeva M. Methods of betulin extraction from Birch bark. Molecules. 2022; 27(11): 3621.
87. Wang J, Shi Y.-M. Recent updates on anticancer activity of betulin and betulinic acid hybrids (a review). Russ J Gen Chem. 2023; 93(3): 610–627.
88. Chen IH, Du YC, Lu MC, Lin AS, Hsieh PW, Wu CC, Chen SL, Yen HF, Chang FR, Wu YC. Lupane-type triterpenoids from *Microtropis fokienensis* and *Perrottetia arisanensis* and the apoptotic effect of 28-hydroxy-3-oxo-lup-20(29)-en-30-al. J Nat Prod. 2008; 71(8): 1352–1357.
89. El-Seedi HR, Ringbom T, Torrsell K, Bohlin L. Constituents of *Hypericum laricifolium* and their cyclooxygenase (COX) enzyme activities. Chem Pharm Bull. 2003; 51(12): 1439–1440.
90. Jiang W, Li X, Dong S, Zhou W. Betulinic acid in the treatment of tumour diseases: application and research progress. Biomed Pharmacother. 2021; 142: 111990.
91. Liu M, Liu X, He K, Jian Y, Li Y, Guo J, Yang J, Xu Z, Kang W. 3-Epi-betulinic acid 3-O- β -D-glucopyranoside (eBAG) induces autophagy by activation of AMP-activated protein kinase in hepatocellular carcinoma. Food Science and Human Wellness, 2024, 13(3): 1453-1464.
92. Cheng YB, Liu FJ, Wang CH, Hwang TL, Tsai YF, Yen CH, Wang HC, Tseng YH, Chien CT, Chen YMA. Bioactive triterpenoids from the leaves and twigs of *Lithocarpus litseifolius* and *L. corneus*. Planta Med. 2018; 84(01): 49–58.
93. Raga DD, Cheng CLC, Lee KCIC, Olaziman WZP, de Guzman VJA, Shen CC, Franco FC, Ragasa CY. Bioactivities of triterpenes and a sterol from *Syzygium samarangense*. Zeitschrift fur Naturforsch - Sect C J Biosci. 2011; 66(5–6): 235–244.
94. Perveen S. Introductory Chapter: Terpenes and terpenoids. IntechOpen. 2018; 1-12.
95. Bathori M, Toth N, Hunyadi A, Marki A, Zador E. Phytoecdysteroids and anabolic-androgenic steroids - structure and effects on humans. Curr Med Chem. 2008; 15(1):75–91.
96. Wei JH, Yin X, Welander P V. Sterol synthesis in diverse bacteria. Front Microbiol. 2016; 7: 990.
97. Parvin T, Akhlas MB, Khatun F, Akter A, Al Amin M, Islam MT, Rouf R. Phytochemical screening and evaluation of

- pharmacological activities of aqueous extract of *Typhonium trilobatum* (L.) Schott. *Orient Pharm Exp Med*. 2019; 19(13): 1-11.
98. Previtera L, Merola D, Monaco P. Acetogenins from the aquatic plant *Elodea canadensis*. *Phytochemistry*. 1985; 24(8): 1838–1840.
99. Kamboj S, Soni SK, Shukla G. Preparation, characterization, and safety assessment of statistical optimized probiotic supplemented herbal wine from *Tinospora cordifolia*. *3 Biotech*. 2023; 13(4)(118): 1-14.
100. Ram H, Jaipal N, Charan J, Kashyap P, Kumar S, Tripathi R, Singh BP, Siddaiah CN, Hashem A, Tabassum B, Abd-Allah EF. Phytoconstituents of an ethanolic pod extract of *Prosopis cineraria* triggers the inhibition of HMG-CoA reductase and the regression of atherosclerotic plaque in hypercholesterolemic rabbits. *Lipids Health Dis*. 2020; 19(1)(6): 1–11.
101. Acebey-Castellon IL, Voutquenne-Nazabadioko L, Doan Thi Mai H, Roseau N, Bouthagane N, Muhammad D, Le Magrex Debar E, Gangloff SC, Litaudon M, Sevenet T, Hung N Van, Lavaud C. Triterpenoid saponins from *Symplocos lancifolia*. *J Nat Prod*. 2011; 74(2): 163–168.
102. Chen CY, Kao CL, Huang ST, Li HT. Secondary metabolites of *Dimocarpus longan*. *Chem Nat Compd*. 2022; 58(3): 578–580.
103. Aamer M, Iqbal S, Jabeen A, Ahmad MS, Asrar M, Yousuf S, Atia-tul-Wahab, Choudhary MI, Wang Y. Bioassay-guided isolation of anti-inflammatory constituents from *Erigeron canadensis*. *Chem Nat Compd*. 2021; 57: 749–751.
104. Rabbi F, Zada A, Nisar A, Sohail M, Khalil SK, Shah WA. Bioassay-guided isolation, identification of compounds from *Sterculia diversifolia* and investigation of their anti-glycation and antioxidant activities. *Pharm Chem J*. 2020; 53(12): 1102–1109.
105. Dai DN, Thang TD, Ogunwande IA. Chemical composition of essential oils from the leaves and stem barks of Vietnamese species of *Polyalthia harmandii*, *Polyalthia jucunda*, and *Polyalthia thorelii*. *Nat Prod Res*. 2014; 28(8): 555–562.
106. Jolanki R, Suhonen R, Henriks-Eckerman ML, Estlander T, Kanerva L. Contact allergy to salicyl alcohol in aspen bark. *Contact Dermatitis*. 1997; 37(6): 304–305.
107. O'Neil MJ. *The Merck index: an encyclopedia of chemicals, drugs, and biologicals*. RSC Publishing; 2013.
108. Lee HH, Shin JS, Lee WS, Ryu B, Jang DS, Lee KT. Biflorin, Isolated from the flower buds of *Syzygium aromaticum* L., Suppresses LPS-Induced inflammatory mediators via STAT1 inactivation in macrophages and protects mice from Endotoxin Shock. *J Nat Prod*. 2016; 79(4): 711–720.
109. De Vasconcellos MC, Bezerra DP, Fonseca AM, Pereira MRP, Lemos TLG, Pessoa ODL, Pessoa C, De Moraes MO, Alves APNN, Costa-Lotufu LV. Antitumor activity of biflorin, an *o*-naphthoquinone isolated from *Capraria biflora*. *Biol Pharm Bull*. 2007; 30(8): 1416–1421.