Tropical Journal of Natural Product Research

Available online at https://www.tjnpr.org *Original Research Article*

Chemical Composition and Antifungal Activity of Essential Oils of *Rosmarinus officinalis* **L***.* **and** *Salvia officinalis* **L***.* **against** *Botrytis cinerea* **Pers**

Manal Kasmi^{1,2*}, Adama Diakite¹, Said Barrijal¹, Haiat Essalmani¹

¹Department of biology, Facultyof Sciences and Techniques, Abdelmalek-Essaadi University, Tangier, Morocco ²Department of biology, Faculty of Sciences, Mohammed V University, Rabat, Morocco.

Copyright: © 2024 Kasmi *et al.* This is an open-

Accepted 11 January 2024 Published online 01 February 2024

access article distributed under the terms of the [Creative Commons](https://creativecommons.org/licenses/by/4.0/) Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

safer alternatives. The aim of this study was to investigate the chemical composition and the antifungalactivity of essential oils of *Rosmarinus officinalis* L.(REO) and *Salvia officinalis* L*.*(SEO), against *B. cinerea* Pers.on tomato leaves and fruits. The EOs were extracted by hydrodistillation, and their chemical composition was analysed by gas chromatography-mass spectrometry (GC-MS). The antifungal activity of the EOs was evaluated on tomato leaves and fruitsinfested with *B. cinerea* Pers. using four treatment methods; simultaneous, preventive, curative, and combined preventive and curative treatments. GC-MS analysis showed that REO is composed mostly of α-pinene (29.01%), verbenone (21.59%), and camphor (7.32%), while SEO is predominantly composed of trans-thujone (29.01%), 1,8-cineole (22.78%), camphor (20.31%), and α -pinene (7.49%). REO and SEO demonstrated a concentration-dependent antifungal activity against *B. cinerea* Pers, especially in the combined preventive and curative treatment which exhibited 100% fungal growth inhibition for both EOs on tomatoleaves and fruits. On the tomato leaves, REO showed the least antifungal activity in the simultaneous treatment (IC $_{50}$ = 7.62 \pm 0.83µL/mL), while on tomato fruits, the curative treatment withSEOexhibited the least effective activity $(IC_{50} = 9.15 \pm 0.53 \mu L/mL)$. Therefore, the essential oils of *R. officinalis* and *S. officinalis*. have potential for use in the biocontrol of tomato grey mould under green house and postharvest storage.

*Keywords***:**Antifungal effect, *Botrytis cinerea* Pers, Chemical composition, Essential oil, *Rosmarinus officinalis* L., *Salvia officinalis* L.

Introduction

Botrytis cinerea Pers. the causative agent of grey mould, is among the most pathogenic fungi in a wide spectrum of plants. It causes loss during culture, transportation and storage in more than 500 plant species across the globe.¹ It attacks many crops of economic importance, including many fruits, vegetables and ornamental crops during pre- and post-harvest period, especially crops under greenhouse conditions.²Among the crops infected by this fungus are tomatoes, (*Lycopersicon esculentum* L.),³ the most important vegetable and main industrial crop around the world.⁴ With the exception of the root, *Botrytis cinerea* infects all plant organs, causing significant losses in yield.⁵ the difficulty in its control lies in its saprophytic nature (survival on plant debris) or in its sclerotia form.⁶

To fight against grey mould and to protect fruits postharvest and to keep them during storage, the synthetic fungicides are commonly used. However, the use of chemical fungicides is increasingly being discouraged due to their harmful effect on human health andenvironment.⁷⁻¹¹

*****Corresponding author. E mail[: manal_kasmi@yahoo.fr](mailto:manal_kasmi@yahoo.fr) Tel: +212679453616

Citation: Kasmi M, Diakite A, Barrijal S, Essalmani H. Chemical Composition and Antifungal Activity of Essential Oils of *Rosmarinus officinalis* L*.* and *Salvia officinalis* L*.* against *Botrytis cinerea* Pers*.* Trop J Nat Prod Res. 2024: 8(1):5897-5907. http://www.doi.org/10.26538/tjnpr/v8i1.27

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria

In addition to causing harm to the environment, human health, and the economy, the use of chemical fungicides induces the development of resistant strains of pathogens to one or more active ingredient.12-17The selection and rapid development of fungicide-resistant *B. cinerea* Pers. genotypes have already been reported.¹⁸⁻²² Thus, several studies have been carried out on eco-friendly and safer alternatives, namely aromatic and medicinal plants and have demonstrated their different biological activities, especially antifungal activity.²³ The interest in the study of the effectiveness of the aromatic and medicinal plants and their essences, particularly essential oils, has increased recently due to the ever-increasing demand in the international market.^{24,25} Essential oils (EOs) have always played an important role in the daily lives of people, they have been widely used for perfuming, flavouring food or even for treatment, because of their potential as an antiseptic agent against infectious diseases, and also, for their properties as cytotoxic agents.²⁶EOs contain a wide range of bioactive compounds, which are natural and biodegradable. EOs compounds can act alone or synergistically,27-30enhancing several activities including antimicrobial, herbicidal, insecticidal, antioxidants and fungicidal activities.³¹⁻³⁶EOs have been the subject of several studies validating its effectiveness as alternatives to commercial pesticides for ecological conservation,³⁷ and also as a natural alternative to the control of antibiotic-resistant microorganisms.³⁸ Recently, several studies have demonstrated that essential oils have antifungal effect against plant pathogenic fungi namely *Botrytis cinerea*Pers.³⁹ The aim of this study is to demonstrate the effectness of the essential oils of *Rosmarinus officinalis* L. and *Salvia officinalis* L. against *Botrytis cinerea* Pers., for possible biological control against the deterioration of tomato crops.

Materials and Methods

Fungal pathogen

Highly virulent and fungicide-resistant strain of *Botrytis cinerea* Pers. was used for the study.

Cultivation of Tomato Plant

The Campbell 33 variety of tomatoes (leaves and fruits) was used for the experiment. Healthy tomato plants were grown on sterile sand, as described by Mouria *et al*. ⁴⁰The plants were maintained in a culture chamber under controlled conditions (21°C, 16-hour photoperiod).

Extraction of Essential Oils

The essential oils were extracted by hydrodistillation as previously described.⁴¹Briefly, about 100 g each of the dried and crushed leaves of *Rosmarinus officinalis* L. and *Salvia officinalis* L. were placed in a 2 L flask to which 1 L of distilled water was added and placed on a heating mantle. After boiling for 3 h, the steam, after being cooled in the condenser, descends into the collector, where the essential oils and water separate by density difference. After removing the water, the oils were collected in a dark bottle and stored at 4°C in a refrigerator until use.

Gas Chromatographic-Mass Spectrometric (GC-MS) Analysis

The GC-MS analyses were carried out in Station d'Ionisation de Boukhalef (SIBO), of the Institut National de Recherche Agronomique (INRA) of Tangier, using gas chromatography coupled with electronically controlled mass spectrometry: GC-MS: Trace GC Ultra-ITQ900 thermoscientific, equipped with a TG-SQC column, (thermoscientific) with a length of more than 60 m. The starting temperature was set at 50°C, rising to 240°C at 5°C/min, with an automatic injection mode (TRIPLUS RSH- thermoscientific) by a 10µL micro syringe. The volume injected was 1µL (20 µL of HE diluted in 1000 of hexane). The carrier gas was helium with a fixed flow rate of 1 mL/min. The temperature of the injector was 250°C in split mode with ratio 1/20, with an Ion trap detector, at 230°C and hexane as eluent. The identification of the constituents was based on the comparison of their respective mass spectra (GC/MS) with the library spectra and on the basis of the calculation of the Kovats indices.

Determination of the antifungal activity of essential oils in vivo

The antifungal effects of different concentrations (0.25, 0.5, 0.75, 1, 10 and 20 µL/mL) of EOs of *S. officinalis* L. and *R. officinalis* L. were tested against *B. cinerea* Pers. on tomato leaves and fruits, using four types of treatments methods which include simultaneous treatment, preventive treatment, curative treatment, and a combination of preventive and curative treatment. The different concentrations of EOs were prepared in 1 mL of a previously sterilised mixture of tween 20 (0.1%) and ethanol (3%). For the simultaneous treatment, the treatment with EOs was carried out at the same time of inoculation with the pathogen. For the preventive treatment, the treatment with EOs was carried out 24 h before inoculation with the pathogen, for the curative treatment, treatment with EOs was done 24 h after inoculation with the pathogen, while for the combination of preventive and curative treatment, the EOs were applied 24 h before and 24 h after pathogen inoculation. Each test was carried out in seven replicates. The controls were treated with the mixture of tween 20 (0.1%) and ethanol (3%) only.

Leaf assays

Eight-week-old tomato seedlings were treated by spraying 1 mL of each concentration of the EOs on the leaves simultaneously, 24 h before, 24 h after, and both 24 h before and 24 h after inoculation with *B. cinerea* Pers. The treated leaves were inoculated with a mycelial explant (5mm) of seven-day-old pathogen. The seedlings were then placed in a growth chamber at 23±2°C and high humidity. After 5 days of incubation, the effects of SEO and REO was assessed by measuring the average diameter of the rot zone on the treated leaves.

Fruit assays

Uniform and healthy tomato fruits of the Campbell 33 variety were sterilised with 70% ethanol, rinsed and dried aseptically. Wounds of 3 mm depth were made with sterile needles (2 mm) at 3 different points

on the equatorial zone. The fruits were subjected to the four types of treatments mentioned above for the leaf assay. A 20 μL of each concentration of EOs were placed in the wounds simultaneously, before, after inoculation. The wounds were inoculated with a mycelial explant (2mm) of seven-day-old *B. cinerea* Pers. The fruits were then placed in the dark at 21±2°C and high humidity conditions. After 7 days of incubation, the mean diameters of the rot zone and the spore zone were measured.

Evaluation of inhibitory activity

The antifungal index (AFI) of each essential oil was determined by the following formula:

AFI (%) = $((d-di)/d) \times 100$.

Where; $d =$ Diameter of mycelial growth in the absence of essential oils. di = Diameter of mycelial growth in the presence of essential oils.

Statistical analysis

All experiments were performed twice under the same conditions with seven replicates. The results obtained were subjected to one-way analysis of variance (ANOVA), and differences between means were determined using Tukey's Post-hoc test. Significant difference was set at $P < 0.05$.

Results and Discussion

Chemical composition of the essential oils

GC/MS analysis of the essential oils identified thirty-six constituents in REO, which represent a total of 98.84% of the essence and twenty-one constituents in SEO representing 99.69% of the essence (Table 1). REO is composed of α-pinene (29.01%), verbenone (21.59%), camphor (7.32%), camphene (4.98%), 1,8-cineole (3.54%), β-copaene (3.49%), borneol (3.43%) and limonene (3.02%) and other minor compounds. The chemical composition of REO obtained in this study is very similar to those reported in several works, with camphor, α-pinene, 1,8-cineole, camphene, β-pinene, Myrcene, borneol and piperitone as major compounds.42-45 A studycarried out on the chemotaxonomy of Moroccan REO, identified three chemotypes: 1,8-Cineole (58-63%), camphor (41-53%) and α-pinene (37-40%).⁴⁶Flamini *et al.* (2002)⁴⁷ found the highest alpha-pinene content (30.3%) in the essential oil extracted from the dried leaves of *Rosmarinus officinalis*. Ainane *et al*. (2018)⁴⁸ found that REO contains camphor (31.16%), β-caryophyllene (18.55%), 2,4-hexadiene, 3,4 -dimethyl-, (Z, Z) (9.08%), α-fenchene (4.67%), cis-verbenone (4.33%) and bornyl acetate (3.4%) as the major compounds, while the rest of the constituents represented less than 1%. Other researchers have demonstrated that *R. officinalis* extract are mainly composed of aromatic compounds such as borneol, camphene, camphor and cineol, flavonoids, tannins, rosmarinic acid, diterpenes, and rosmarin.49-51In fact many previous studies, have shown that the chemical composition of REO can vary qualitatively and quantitatively depending on several factors. The SEO were predominantly composed of trans-thujone (29.01%), 1,8-cineole (22.78%), camphor (20.31%), αpinene (7.49%), camphene (4.99%) and myrcenol (2.96%), these compounds constituted more than three quarter of the total composition. In 2015, Rus *et al*., ⁵² identified camphor (20.4%), eucalyptol (11.7%), camphene (11.5%), α-pinene (9.5%) as major compounds of the SEO. Similarly, several previous studies have confirmed that SEOcontain αpinene, camphene, β-pinene, Myrcene, 1,8-cineole, camphor, borneol, bornyl acetate, (E)-caryophyllene, and viridiflorol as major compounds.^{44,53-56}Also, the study ofAlexa, $(2018)^{57}$ identified caryophyllene (25.364%), camphene (14.139%), eucalyptol (13.902%), and β-pinene (11.230%), thymol (8.073%), camphor (4.028%), and valancene (5.525%) as themain compounds of SEO, Camphor has been reported as the main compound of SEO in several studies,⁵⁸⁻⁶⁰ while other authors have reported α-thujone as the main compound of SEO, 58,61,62 which is in line with the results of this study which shows that REO and SEO are mainly composed of α-pinene, trans-thujone, 1,8-cinéole, and camphor.

Generally, despite the variations in the chemical composition, EOs are usually composed of two major chemical groups, the terpenoids, which are the most predominant (represented mainly by monoterpenes and sesquiterpenes), $63-66$ and the phenylpropanoids. $63, 67-69$ In fact, the

ISSN 2616-0692 (Electronic)

variations in the results obtained in the different studies cited explains the role that environmental and climatic factors (geographical origin, time of harvest, light exposure)play in the chemical composition and quality of EOs.⁷⁰ In addition to the environmental and climatic factors, genetic factor also have a considerable influence on the quantitative and qualitative composition of EOs.⁷¹

In vivo antifungal activity of essential oils against B. cinerea Pers. *on tomato leaves*

The essential oils of *R. officinalis* L. (REO) and *S. officinalis* L. (SEO) showed aconcentration-dependent antifungal effect against *B. cinerea* Pers. (Figure 1). A recent study has proven the effictiveness of several EOs against several fungal pathogens including *B. cinerea* Pers.³⁹ The antifungal activity of EOs could be attributed to the major phytochemical group of compounds which are the monoterpenes with proven antifungal activity, $2^{9,72-75}$ and the sesquiterpenes, which play a principal role in defence against bacterial and fungal attacks.⁶⁶ Generally, since terpenoids are non-polar with hydrophobic and lipophilic characters that facilitate their interaction with the membrane components of fungal cell, causing cell membrane damage and interferance in energy homeostasis. All substances rich in terpenoids are considered natural antifungals.76,77The compounds of EOs can act alone or in synergy with other compounds with a promise of greater effectiveness.30,78,79

With respect to the effects of the different types of treatments with REO against tomato leaf rot, the combined preventive and curative treatment was the most effective, resulting in 100% inhibition at concentration of 10 µL/mL (Figure 1A), followed by the curative treatment which gave 100% inhibition at concentration of 20 µL/mL, then the preventive treatment while the simultaneous treatment was the least effective producing slightly above 50% inhibition of the rot at 10 µL/mL (Figure 1A). In fact, the leaves treated with the simultaneous treatment at low concentrations showed no significant difference compared with the positive control leaves (inoculated with *B. cinerea* Pers. only), while the leaves treated with the other treatments were not significantly different from the negative control (healthy leaves) from a concentration of 10 µL/mL (Figure 2). On the other hand, SEO resulted in a significant inhibition of the rot (Figure 1B).The antifungal effect of SEO against *B. cinerea* Pers. and *Fusarium* sp. has been highlighted.80,81 Also, Rguez et al. (2018)⁸² found that SEO showed significant antifungal effect *in vitro* against *B. cinerea* Pers*.* With respect to the effectiveness of the type of treatment, the combined preventive and curative treatment consistently gave the most potent activity, resulting in 100% inhibition from a concentration of $1 \mu L/mL$ and no significant difference with the healthy control (Figure 2), this was followed by the preventive treatment and then the curative treatment, and lastly, the simultaneous treatment (Figure 1B).

Table 1: Chemical composition of *R. officinalis* L. and *S. officinalis* L. essential oils

Chemical group	Components	EO of R. officinalis	EO of S. officinalis
Terpenes	α -thujene		0.16%
	α -pinene	29.01%	7.49%
	Camphene	4.98%	4.99%
	β -pinene	0.88%	1.66%
	Myrcene	0.46%	0.94%
	α -terpinene	0.56%	$\bar{}$
	Limonene	3.02%	0.63%
	γ -terpinene	1.14%	0.28%
	Alloocimene	0.33%	$\overline{}$
	allo-Aromadendrene	\blacksquare	1.01%
	β -bourbonene	0.33%	\bar{a}
	α -cedrene	0.28%	
	β -copaene	3.49%	1.81%
	β -acoradiene	2.22%	$\overline{}$
	Valencene	0.54%	
	δ -cadinene	0.43%	
	γ -bisabolene E	1.26%	
Total *		48.93 % *	18.97*
Ketones	Camphor	7.32%	20.31%
	Menthone	$\overline{}$	0.33%
	Cryptone	1.32%	0.71%
	Dihydrocarvone	0.52%	÷.
	β -oplopenone	$\overline{}$	1.88%
	Verbenone	21.59%	$\overline{}$
	β -ionone,	0.58%	
	Trans-thujone	$\overline{}$	29.01%
	Cis-thujone	1.8%	
Total *		33.13%*	52.24*
Alcohols	Linalol	0.84%	0.26%

Trop J Nat Prod Res, January 2024; 8(1):5897-5907 **ISSN 2616-0684 (Print)**

(-): Absent

On comparison of the IC_{50} values of the two EOs studied (Table 2), it was observed that the combined preventive and curative treatment with REO was the most effective $(IC_{50} < 0.25 \mu L/mL)$, followed by the curative treatment and the preventive treatment with IC₅₀values of0.53µL/mL and 0.66 µL/mL, respectively, while the simultaneous treatment was the least effective (IC $50=7.62 \mu L/mL$). Like REO, the combined preventive and curative treatment of SEO was the most effective (IC₅₀< 0.25 μ L/mL), followed by the preventive treatment and the simultaneous treatment (IC₅₀= $0.48 \mu L/mL$ and IC₅₀= $0.96 \mu L/mL$, respectively), while the curative treatment was the least effective $(IC_{50} =$ 4.21 μ L/mL). On comparing the IC₅₀ values of the different treatments of the two EOs, the simultaneous treatment of REO appeared to be the least effective, followed by the curative treatment of SEO. Moreover, the preventive treatment and the curative treatment of REO as well as the simultaneous treatment and the preventive treatment of SEO were not significantly different. The determination of IC⁹⁰ allowed us to deduce that the combined preventive and curative treatment of SEO (IC₉₀= 0.61 µL/mL) was more effective than that of REO (IC₉₀= 7.64) µL/mL).The fact that the combined preventive and curative treatment, and the preventive treatment were more effective against the leaf rot, suggests that the EOs stimulate in the plant certain defense mechanisms against *B. cinerea* Pers. and consequently the accumulation of secondary metabolites in the plants. In this regard, differents studies revealed that EOs stimulate plant defense mechanisms against pathogenic infection on the basis of their antioxidant activity and ability to improve plant growth.⁸³⁻⁸⁵

The results obtained in the simultaneous treatments and curative treatments is justified by the fact that REO may contain substances that are capable of inhibiting the development of the disease at stages subsequent to infection, so if they are applied simultaneously with the infection by the pathogen, they evaporate before they act. Whereas, if they are applied after the infection (curative treatment), they act directly on the mechanisms of the disease development. On the other hand, SEO could be rich in substances that act at the earlier stages of the infection, so if they are applied simultaneously with the infection, they are effective, whereas when they are applied after the infection (curative treatment), the infection is not inhibited because the pathogen has

already developed. It could be deduced that the EOs studied act differently, due to their diverse chemical constituents. In fact, EOs have several functional agents that enhance the antifungal activity and deactivate the fungus by the interaction between hydrophobic compounds and lipids of the fungal cell membrane, $63,86,87$ interrupting its structure and function such as inhibiting nuclear material or protein synthesis,^{69,87}and may also cause membrane expansion, affect the morphogenesis and growth of the hyphae.88-90 Caccioni and Guzzardi $(2011)^{91}$ showed that monoterpenes such as α -pinene caused morphological changes in the fungal hyphae by disrupting cell membrane. They reveled that EOs with high concentrations of α-pinene had antifungal effects against differents pathogens. Similarly, Reguez *et al*. (2020)⁹² showed that the pure α-pinene of *Tetraclinis articulata* EOs caused bulge of *B. cinerea* Pers. conidia which were probably attributed to a disturbance in membrane permeability. On the other hand, α-pinene reduces germination of *B. cinerea* Pers. conidia and alters their morphology. Also, Fernando *et al.* (2013)⁶⁵ proved that αpinene targets the integrity of cell membrane, causing morphological alterations in the conidia such as rupture of membranes and vacuolization and disorganization of the cytoplasm. On the fungal mycelium, EOs act on the key proteins involved in the regulation of metabolism, growth and cell differentiation.93,94 They lead to loss of cell membrane integrity and rigidity, causing disturbance of mycelium,⁹⁵ and alterations of fungal morphology (rupture, peeling, dehydration and interference with energy homeostasis).29,76,77 According to Paula *et* al. (2020),⁹⁶ the antifungal effect of the EOs against *B. cinerea* Pers. could be attributed to the chemical composition of these EOs including monoterpenes, as α-pinene, α-thujene, limonene, α-terpinolene, 1,8 cineole, and α-terpinyl acetate.On the other hand, some authors suggest that the mechanism of EOs against fungal species, is probably attributed to the high content of lipid peroxides, such as the hydroxyl radicals, inhibition of cell membrane synthesis of ergosterol, mitochondrial fragmentation, and inhibition of respiratory enzymes.⁹⁷⁻⁹⁹ Several other studies confirmed that EOs can cause cell death and/or inhibition of sporulation and germination of fungi by interfering with the mitochondrial respiratory chain complexes, leading to mitochondrial

ISSN 2616-0692 (Electronic)

dysfunction, an imbalance in cellular metabolic activity, and consequently a drop in energy availability.86,89,100-103

The antifungal activity of Eos is attributed not only to a single compound,^{104,105} but also to sevearal compounds which act synergistically.^{90,106} Minor compounds of EOs such as alcohols, aldehydes and esters have been shown to have antimicrobial activity.¹⁰⁷ In fact, plants rich in phenolics seem also to be more effective against phytopathogens.⁶⁷⁻⁶

In vivo antifungal activity of essential oils against B. cinerea Pers. *on tomato fruits*

On fruit rot, REO and SEO demonstrated a remarkable inhibitory activity against *B. cinerea* Pers. especially with the combined preventive and curative treatment which resulted in a 100% inhibition of the rot at concentrations of 0.25 µL/mL and 0.75 µL/mL for REO and SEO, respectively (Figures 3 and 4).The effect of the combined preventive and curative treatment was followed by that of the preventive treatment which produced slightly above 50% inhibition at 0.25 µL/mL and 0.5 µL/mL for REO and SEO, respectively. However, the simultaneous treatment and the curative treatment with SEO and REO were less effective, producing approximately 50% inhibition at 1 uL/mL

Figure 1: Antifungal effect of differents treatments against *B. cinerea* Pers. on tomato leaves. A: Treatments with *R. officinalis* L. essential oils, B: Treatments with *S. officinalis* L. essential oils

Figure 2: Effect of preventive and curative treatments with 10µL/mL of EOs against the rot of tomato leaves after 5 days of incubation. A: Leaf treated with *R. officinalis* L. essential oils, B: Leaf treated with *S. officinalis* L. essential oils, C: Positive control.

Determination of the IC⁵⁰ of REO and SEO against *B. cinerea* Pers. growth on the fruits confirmed that the combined preventive and curative treatment was the most effective against fruit rot $(IC_{50} < 0.25)$ µL/mL), while the curative treatment with SEO was the least effective with IC_{50} of 9.15 $\mu L/mL$ (Table 2). Although, the range of concentrations studied did not allow for the determination of IC⁹⁰ for all the treatments methods, but the IC⁹⁰ of only the combined preventive and curative treatments with the two essential oils were determined, and it was found out that the combined preventive and curative treatment of rosemary (IC $90 < 0.25$ µL/mL) was more effective than that of sage (IC90= 0.39 µL/mL).

The results obtained are in agreement with the findings of other studies which have proven that 60% of the EOs, or of their derivatives, are effective against a wide spectrum of fungi. Thus these EOscan be used to control fungal diseases in fruits and vegetables, ensuring an extension of their storage life.108,109 In fact EOs have been shown to be effective against *B. cinerea* Pers. Infection on fruit and against several other fungal infections. For example, on apple, *Satureja hortensis* and *Thymus vulgaris*, EOs have been shown to be effective against *B. cinerea* Pers. during postharvest.⁸⁵

The fact that the combined preventive and curative treatment, and the preventive treatment were more effective against the fruit rot, suggests that the essential oils stimulated in the fruits certain defense mechanisms against *B. cinerea* Pers. According to Gholamnezhad (2019) ,¹¹⁰ the treatment of apple fruits with plant extracts induced POX, PAL, β-1,3-glu and PPO and total phenolic contents in plants infected by *B. cinerea* Pers. The induction of phenolic compounds by plant extracts, suggests the roles of these compounds in resistance induction against fungal pathogens. Plant extracts, elicits production in apple fruits molecules that act against apple gray mould, especially phenol compounds which have a strong effect on the induction of POX, PPO, and PAL enzymes. Thus these plant extracts can be useful in the control of apple grey mould.

With regards to sporulation on fruits, the results obtained indicated that the combined preventive and curative treatment, as well as the preventive treatment with REO and SEO remain the most effective in reducing the growth of the sporulation zone, especially the combined preventive and curative treatment which caused 100% inhibition of sporulation even at the lowest concentration of the two essential oils (Table 3). On the other hand, for the curative treatment and the simultaneous treatment with low concentrations of the essential oils, it was found that the diameter of the sporulated zone on the treated fruits

significantly exceeds that of the positive control, which means that these two treatments stimulated sporulation at low concentration, especially the curative treatment which only showed a slight inhibition at the 20 µL/mL concentration (Figure 5A). Figure 5B shows that during the simultaneous treatment, the antifungal effect was only observed from concentration of 10µL/mL (Table 3). This was confirmed by the fact that the REO and SEO at low concentrations are unable to inhibit fungal growth when applied simultaneously or curatively, but they create stressful conditions that stimulate sporulation of the fungus to ensure its dissemination and thus its survival. Comparing the inhibitory effect of the simultaneous treatment of the two EOs, it appeared that REO are more effective than SEO. On the basis of the results obtained for the tomato fruits, the REO and SEO hold promise for the biocontrol and the management of postharvest pathogen of tomatoes, especially *B. cinerea* Pers. which has been described as the most important postharvest pathogen.111,112 However, the effectiveness of these biocontrol agents can be inflenced by the mode of their application which is a fundamental part of biocontrol. From the results of the present study, it appears that the application of EOs as biocontrol agents of *B. cinerea* Pers. should be both preventive and curative in order to stimulate the defence mechanisms of the leaves and/or fruit before infection and to act on the pathogen after infection.

Conclusion

The antifungal activity of the essential oils of *R. officinalis* L. and *S. officinalis* L. against *B. cinerea* Pers. on tomato leaves or fruits is concentration-dependent, the activity increases with increase in concentration. According to these results, the high antifungal activity observed in the essential oils of *S. officinalis* L. and *R. officinalis* L. could be attributed to the efficacy of its components which may act singly or synergistically.The preventive treatment as well as the combined preventive and curative treatment on tomato leaves and fruits with EOs of *R. officinalis* L. and *S. officinalis* L. reduced the severity of gray mould disease. There was no significant reduction in the severity of *B. cinerea* Pers. with the simultaneous treatment. Based on the findings from this study, postharvest losses of stored crops may be reduced if crops are protected with EOsin a preventive manner. Thus the essential oils of *R. officinalis*L. and *S. officinalis* L. have a great potential as antifungal compounds that could be used in the biocontrol of grey mould disease. Additionally EOs of *R. officinalis* L. and *S. officinalis* L. may be safer alternatives to synthetic fungicides in the control of both fungal diseases in the field and during storage. Further studies is therefore encouraged to explore the mechanism of action of isolated and/or combined essential oil components and the formulation of biofungicides based on these components.

Figure 4: Effect of preventive and curative treatment with 0.25 µL/mL of EOs, against the rot of tomato fruits after 7 days of incubation. **A:** Tomato fruit treated with *S. officinalis* L. essential oils, **B:** Tomato fruit treated with *R. officinalis* L. essential oils, **C:** Positive control.

Conflict of Interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

Acknowledgments

The authors would like to thank Mr. Said Zentar, coordinator of SIBO, for the use of his facility for the GC-MS analysis of the essential oils.

Table 2: Antifungal activity of *Rosmarinus officinalis* L. and *Salvia officinalis* L. essential oils against *Botrytis cinerea* Pers*.*

Values are means ±SD. Values in the columns followed by the same superscript letter are not significantly different.

Table 3: Diameter (cm) of the spore zone on tomato fruits after treatment with *Rosmarinus officinalis* L. and *Salvia officinalis* L. essential oils

Values are means ±SD. Values in the columns followed by the same superscript letter are not significantly different.

Figure 5: Effect of *R. officinalis* L. and *S. officinalis* L. essential oils on sporulation. **A:** Curative treatmnt, **B:** Simultanous treatmnt Bars for each concentration, with the same letters represent values that are not significantly different.

References

- 1. FillingerS and Elad Y. Botrytis the Fungus, the Pathogen and Its Management in Agricultural Systems. Springer International Publishing., Switzerland. 2016.
- 2. Elad Y. Effect of filtration of solar light on the production of conidia by field isolates of *Botrytis cinerea* and on several diseases of green house‐grown vegetables. Crop Prot. 1997; 16(7): 635-642
- 3. Dulger B and Hacioglu N. Antifungal activity of Endemic *Salvia tigrina* in Turkey. Trop J Pharm Res. 2008; 7:1051- 1054.
- 4. Abdallah RAB, Jabnoun-Khiareddine H, Nefzi A, Mokni-Tlili S, Daami-Remadi M. Endophytic Bacteria from *Datura stamonium* for Fusarium wilt suppression and tomato growth promotion. J Microb Biochem Sci Technol. 2016; 26(8):1- 51.
- 5. Fiume F and Fiume F. Biological control of *Botrytis cinerea* gray mould on tomato cultivated in greenhouse. Comm Appl Biol Sci. 2006; 71:897.
- 6. Wang K, Xion Q, Kan Jianquan K, Cao S, Zheng Y. Induction of director priming resistance against *Botrytis cinerea* in strawberries by β-aminobutyric acid and their effects on sucrose metabolism. J Agric Food Chem. 2016; 64:5855-5865.
- 7. Yan D, Zhang Y, Liu L, Yan H. Pesticide exposure and risk of Alzheimer's disease: A systematic review and metaanalysis. Sci Rep. 2016; 6:1-9.
- 8. Cimino AM, Boyles AL, Thayer KA, Perry MJ. Effects of neonicotinoid pesticide exposure on human health: A systematic review. Environ Health Perspect. 2017; 125(2):155-162.
- 9. Sun C, Lin M, Fu D, Yang J, Huang Y, Zheng X, Yu T. Yeast cell wall induces disease resistance against *Penicillium expansum* in pear fruit and the possible mechanisms involved. Food Chem. 2017; 241:301-307.
- 10. Brandhorst TT and Klein BS. Uncertainty surrounding the mechanism and safety of the postharvest fungicide fludioxonil. Food Chem Toxicol. 2018; 123:561-565.
- 11. Catto C, de Vincenti L, Borgonovo G, Bassoli A, Marai S, Villa F, Cappitelli F, Saracchi M. Sub-lethal concentrations of Perilla frutescens essential oils affect phytopathogenic fungal biofilms. J Environ Manag. 2019; 245:264-272.
- 12. Ali A, Bordoh PK, Singh A, Siddiqui Y, Droby S. Postharvest development of anthracnose in pepper (*Capsicum spp*): Etiology and management strategies. Crop Protect. 2016; 90:132-141.
- 13. Mari M, Bautista-Banos S, Sivakumar D. Decay control in the postharvest system: Role of microbial and plant volatile organic compounds. Postharv Biol Technol. 2016; 122:70- 81.
- 14. Munir M, Amsden B, Dixon E, Vaillancourt L, Ward Gauthier NA. Characterization of *Colletotrichum* species causing bitter rot of apple in Kentucky orchards. Plant Dis. 2016; 100(11):2194-2203.
- 15. Gonzalez-Estrada RR, Chalier P, Ragazzo-Sanchez JA, Konuk D, Calderon-Santoyo M. Antimicrobial soy protein based coatings: Application to Persian lime (*Citrus latifolia* Tanaka) for protection and preservation. Postharv Biol Technol. 2017; 132:138-144.
- 16. Zhang L, Song L, Xu X, Zou X, Duan K, Gao Q. Characterization and Fungicide Sensitivity of *Colletotrichum* Species Causing Strawberry Anthracnose in Eastern China. Plant Dis. 2020, Article PDIS-10-19-2241.
- 17. Sarkhosh A, Schaffer B, Vargas AI, Palmateer AJ, Lopez P, Soleymani A, Farzaneh M. Antifungal activity of five plantextracted essential oils against anthracnose in papaya fruit. Biol Agric Horticult. 2017; 34(1):18–26.
- 18. Leroch M, Kretschmer M, Hahn M. Fungicide resistance phenotypes of *Botrytis cinerea* isolates from commercial vineyards in South West Germany. J Phytopathol. 2011; 159:63–65.
- 19. Leroch M, Plesken C, Weber RWS, Kauff F, Scalliet G, Hahn M. Gray mold populations in German strawberry fields show multiple fungicide resistance and are dominated by a novel clade close to *Botrytis cinerea*. Appl Environ Microbiol. 2013: 79:159-167.
- 20. Grabke J, Fernandez-Ortuno A, Amiri D, Li A, Peres X, Smith NA, Schnabel P. Characterization of iprodione resistance in *Botrytis cinerea* from strawberry and blackberry. Phytopathol. 2014; 104:396-402.
- 21. Lopes UP, Zambolim L, Capobiango NP, Gracia NAO, Freitas-Lopes RL. Resistance of *Botrytis cinerea* to fungicides controlling gray mold on strawberry in Brazil. Bragantia. 2017; 76(2):266-272.
- 22. Avenot HF, QuattriniJ, Puckett R, Michailides TJ. Different levels of resistance to cyprodinil and iprodione and lack of fludioxonil resistance in *Botrytis cinerea* isolates collected from pistachio, grape and pomegranate fields in California. Crop Prot. 2018; 112:274-281.
- 23. El Ouadi Y, Manssouri M, Bouyanzer A, Majidi L, Lahhit N, Bendaif H, Costa J, Chetouani A, Elmsellem H, Hammouti B. Essential oil composition and antifungal activity of *Salvia officinalis* originating from North-East Morocco, against postharvest phytopathogenic fungi in apples. Der Pharm Chem. 2015; 7(9):95e102.
- 24. González-Tejero MR, Casares-Porcel M, Sánchez-Rojas CP, Ramiro-Gutiérrez JM, Molero-Mesa A, Pieroni D. Paraskeva-Hadijchambi. J Ethnopharmacol. 2008; 116:341- 357.
- 25. Jamila F and Mostafa E. Ethnobotanical survey of medicinal plants used by people in Oriental Morocco to manage various ailments. J Ethnopharmacol. 2014; 154:76- 87.
- 26. Ali B, Al-wabel NA, ShamsS, Ahamad A, Khan SA, Anwar F.Essential oils used in aromatherapy: A systemic review. Asian Pac J Trop Biomed. 2015; 5:601-611.
- 27. Issouffou C, Suwansri S, Salaipeth L, Domig KJ, Hwanhlem N. Synergistic effect of essential oils and enterocin KT2W2G on the growth of spoilage microorganisms isolated from spoiled banana peel. Food Contr. 2018; 89:260-269.
- 28. Oliveira J, Parisi MCM, Baggio JS, Silva PPM, Paviani B, Spoto MHF, Gloria EM. Control of *Rhizopus stolonifer* in strawberries by the combination of essential oil with carboxymethylcellulose. Int J Food Microbiol. 2019; 292:150-158.
- 29. Oliveira J, Gloria EM, Parisi MCM, Baggio JS, Silva PPM, Ambrosio CMS, Spoto MHF. Antifungal activity of essential oils associated with carboxymethylcellulose against Colletotrichum acutatum in strawberries. Sci Horticult. 2019a; 243:261-267.
- 30. OuYang Q, Okwong RO, Chen Y, Tao N. Synergistic activity of cinnamaldehyde and citronellal against green mold in citrus fruit. Postharv Biol Technol. 2020; 162:111095.
- 31. Zhang X, Guo Y, Guo L, Jiang H, Ji Q. *In vitro* evaluation of antioxidant and antimicrobial activities of *Melaleuca alternifolia* essential oil. BioMed Res Int. 2018; 2018:2396109.
- 32. Feng G, Zhang XS, Zhang ZK, Ye HC, Liu YQ, Yang GZ, Chen C, Chen M, Yan C, Wang LY, Zhang JX, Zhang J. Fungicidal activities of camptothecin semisynthetic derivatives against *Colletotrichum gloeosporioidesin vitro*and in mango fruit. Postharv Biol Technol. 2019; 147:139-147.
- 33. Jahani M, Pira M, Aminifard MH. Antifungal effects of essential oils against *Aspergillus nigerin vitro* and *in vivo* on pomegranate (*Punica granatum*) fruits. Sci Horticult. 2020; 264:109188.
- 34. Matos Ferreira L, Douglas Rafael e Silva B, Emanuel da Cruz L, Dutra KA, Daniela Maria do Amaral Ferraz N, Jessica Lafaiete Ribeiro A, Gutierres Nelson S. Chemical composition and insecticidal effect of essential oils from

Illicium verum and *Eugenia caryophyllus* on *Callosobruchus maculatus* in cowpea. Ind Crops Prod. 2020; 145:112088.

- 35. Mehdizadeh L, Taheri P, Ghasemi PA, Moghaddam M. Phytotoxicity and antifungal properties of the essential oil from the *Juniperus polycarpos* var. turcomanica (B. Fedsch.) R.P. Adams leaves. Physiol Mol Biol Plants. 2020; 26(4):759-771.
- 36. Raspo MA, Vignola MB, Andreatta AE, Juliani HR. Antioxidant and antimicrobial activities of citrus essential oils from Argentina and the United States. Food Biosci. 2020l; 36:100651.
- 37. Asgar E, Masumeh Z, Franco P. Molecules. 2020; 25(7):1- 15.
- 38. Chavez-Gonzalez ML, Rodriguez R, Aguilar CN. Essential Oils: A Natural Alternative to Combat Antibiotics Resistance. In: Kateryna Kon; Mahendra Rai (Eds), Antibiotic Resistance. Mechanisms and New Antimicrobial Approaches, Chapter 11; Elsevier, 2016; 227-237 p.
- 39. Zatla AT, Dib MA, Djabou N, Ilias F, Costa J, Muselli A. Antifungal activities of essential oils and hydrosol extracts of *Daucus carota* subsp sativus for the control of fungal pathogens, in particular gray rot of strawberry during storage. J Essent Oil Res. 2017; 29:391-399.
- 40. Mouria A, Ouazzani-Touhami A, Douira A. Mise en évidence d"une variation intra spécifique chez *Botrytis cinerea* et lutte biologique *in vitro* par l"extrait de compost. J Appl Biosci. 2013; 64:4797-4812.
- 41. Kasmi M and Essalmani H.Antifungal Effect of *Rosmarinus officinalis*L. and*Salvia officinalis* L. Extracts against Tomato Grey Mould.Trop J Nat Prod Res, February 2022, 6(2), 180-187.
- 42. Djeddi S, Bouchenah N, Settar I, Skaltsa HD. Composition and antimicrobial activity of the essential oil of *Rosmarinus officinalis*from Algeria. Chem Nat Comp. 2007; 43(4):487- 490.
- 43. Rasooli I. Antimycotoxigenic characteristics of *Rosmarinus officinalis*and *Trachyspermum copticum* L. essential oils; Int J Food Microbiol. 2008; 122:135-139.
- 44. Hussain AI, Anwar F, Iqbal T, Bhatti IA. Antioxidant attributes of fourlamiaceae essential oils. Pak J Bot. 2011; 43(2):1315-1321.
- 45. Jiang Y, Wu N, Fu YJ, Wang W, Luo M, Zhao CJ, Zu YG, Liu XL. Chemical composition and antimicrobial activity of the essential oil of Rosemary. Environ Toxicol Pharmacol. 2011; 32(1):63-68.
- 46. Elamrani A, Zrira S, Benjilali B. A study of Moroccan rosemary oils. J Ess Oil Res. 2000; 12:487-495.
- 47. Flamini G, Cioni PL, Morelli I, Macchia M, Ceccarini L. Main agronomicproductive characteristics of two ecotypes of *Rosmarinus officinalis*L. and chemical composition of their essential oils. J Agric Food Chem. 2002; 50:3512-3517.
- 48. Ainane A, Benhima R, Khammour F, Elkouali M, Talbi M, Abba E, Cherroud S, Ainane T. Composition chimique et activité insecticide de cinq huiles essentielles: *Cedrus atlantica*, *Citrus limonum*, *Eucalyptus globules*, *Rosmarinus officinalis*et *Syzygium aromaticum*. BIOSUNE'1. 2018; 67- 79 p.
- 49. Akbari J, Saeedi M, Farzin D, Morteza-Semnani K, Esmaili Z.Transdermal absorption enhancing effect of the essential oil of *Rosmarinus officinalis*on percutaneous absorption of Na diclofenac from topical gel.Pharm Biol. 2015; 53:1442- 1447.
- 50. Tak JH, Jovel E, Isman MB.Comparative and synergistic activity of *Rosmarinus officinalis*L. essential oil constituents against the larvae and an ovarian cell line of the cabbage looper, Trichoplusia ni (Lepidoptera: Noctuidae).Pest Manag Sci. 2016; 72:474-480.
- 51. Mekonnen A, Yitayew B, Tesema A, Taddese S. "*In Vitro* Antimicrobial Activity of Essential Oil of *Thymus schimperi*, *Matricaria chamomilla*, *Eucalyptus globulus*,

and *Rosmarinus officinalis*". Int J Microbiol. 2016; 2016:Article ID 9545693. 8 pages.

- 52. Rus CF, Alexa E, Pop G, Sumalan RM, Copolovici M. Antifungal activity and chemical composition of *Salvia officinalis* L. essential oil. Res J Agric Sci. 2015; 47:186-194.
- 53. Radulescu V, Chiliment S, Oprea E. Capillary gas chromatography-mass spectrometry of volatile and semivolatile compounds of *Salvia officinalis*. J Chromatogr A. 2004; 1027)1-2):121-126.
- 54. Vukovic-Gacic B, Nikcevic S, Beric-Bjedov T, Knezevic-Vukcevic J, Simic D. Antimutagenic effect of essential oil of sage (*Salvia officinalis* L.) and its monoterpenes against UVinduced mutations in *Escherichia coli* and *Saccharomyces cerevisiae*. Food Chem Toxicol. 2006; 44:1730-1738.
- 55. Pierozan MK, Pauletti GF, Rota L, Santos ACA, Lerin L, Di Luccio M, Mossi AJ, Atti-Serafini L, Cansian RL, Oliveira JV. Chemical characterization and antimicrobial activity of essential oils of Salvia L. species. Ciência Tecnol Alimen. 2009; 29(4):764-770.
- 56. Bouaziz M, Yangui T, Sayadi S. Disinfectant properties of essential oils from *Salvia officinalis* L. cultivated in Tunisia. Food Chem Toxicol. 2009; 47(11):2755-2760.
- 57. Alexa E, Sumalan RM, Danciu C, Obistioiu D, Negrea M, Poiana MA, Rus C, Radulov I, Pop G, Dehelean C. Synergistic Antifungal, Allelopatic and Anti-Proliferative Potential of *Salvia officinalis* L., and *Thymus vulgaris* L. Essential Oils. Molecules. 2018; 23:185.
- 58. Schmidt E, Wanner J, Hiiferl M, Jirovetz L, Buchbauer G, Gochev V, Girova T, Stoyanova A, Geissler M. Chemical composition, olfactory analysis and antibacterial activity of *Thymus vulgaris* chemotypes geraniol, 4-thujanol/terpinen-4-ol, thymol and linalool cultivated in southern France. Nat Prod Commun. 2012; 7:1095-1098.
- 59. Rguez S, Daami-remadi M, Cheib I, Laarif A, Hamrouni I. Composition Chimique, Activité Antifongique et Activité Insecticide de l'Huile Essentielle de *Salvia officinalis.* Tunis J Med Plants Nat Prod. 2013; 9(2):65-76.
- 60. Adrar N, Oukil N, Bedjou F. Antioxidant and antibacterial activities of *Thymus numidicus* and *Salvia officinalis* essential oils alone or in combination. Ind Crops Prod. 2016; 88:112-119.
- 61. Santos-Gomes PC and Fernandes-Ferreira M. Organ- and season-dependent variation in the essential oil composition of *Salvia officinalis* L. cultivated at two different sites. J Agric Food Chem. 2001; 49:2908-2916.
- 62. Baranauskiene R, Venskutonis PR, Viškelis P, Dambrauskiene E. Influence of nitrogen fertilizers on the yield and composition of Thyme (*Thymus vulgaris*). J Agric Food Chem. 2003; 51:7751-7758.
- 63. Vianna TC, Marinho CO, Júnior LM, Ibrahim SA, Vieira RP. Essential oils as additives in active starch-based food packaging films: A review. Int J Biol Macromol. 2021; 182(1):1803–1819.
- 64. Vintila I. Basic Structure, Nomenclature, Classification and Properties of Organic Compounds of Essential Oil. In: Seyed Hashemi, M.B. (Ed.), Essential Oils in Food Processing: Chemistry, Safety and Applications. First Edition. John Wiley & Sons Ltd. Chap. 5. 2018.
- 65. Fernando P, Monteiro I, Larissa C, Ferreira I, Jhonata L, Silva I, Leandro P, Pacheco E, Paulo E. Influence of Plant extracts and essential Oils against Panama Disease (*Fusarium oxysporum* f. sp. cubense) in Banana Seedlings. J Agric Sci. 2013; 5:4–14.
- 66. Buckle, J Clin Aromather. 2015; 37-72.
- 67. Freiesleben S and Jager AK. Correlation between Plant Secondary Metabolites and Their Antifungal Mechanisms–A Review. Med Arom Plants. 2014; 03(02):1–6.
- 68. Gutierrez-del-Río I, Fernandez J, Lombo F. Plant nutraceuticals as antimicrobial agents in food preservation: Terpenoids, polyphenols and thiols. Int J Antimicrob Agents. 2018; 52(3):309–315.
- 69. Tariq S, Wani S, Rasool W, Shafi K, Bhat MA, Prabhakar A, Shalla AH, Rather MA. A comprehensive review of the antibacterial, antifungal and antiviral potential of essential oils and their chemical constituents against drug-resistant microbial pathogens. Microb Pathogen. 2019; 134:103580.
70. Fernandes RV, Borges SV, Botrel DA.Gum
- RV, Borges SV, Botrel DA.Gum arabic/starch/maltodextrin/inulin as wall materials on the microencapsulation of rosemary essential oil.Carbohydr Polym. 2014; 101:524-532.
- 71. Eugénia P, Salgueiro L, Cavaleiro C, Pameira A, Conçalves M. *In vitro* susceptibility of certain species of yeast and filamentous fungi with essential oil. Ind Crops Prod. 2007; 26:135- 141.
- 72. Marei GIK and Abdelgaleil SAM. Antifungal Potential and Biochemical Effects of Monoterpenes and Phenylpropenes on Plant Pathogenic Fungi. Plant Prot Sci. 2018; 54:9–16.
- 73. Zillo RR, da Silva PPM, de Oliveira J, da Gloria EM, Spoto MHF. Carboxymethylcellulose coating associated with essential oil can increase papaya shelf life. Sci Horticult. 2018; 239:70–77.
- 74. Wang H, Yang Z, Ying G, Yang M, Nian Y, Wei F, Kong W. Antifungal evaluation of plant essential oils and their major components against toxigenic fungi. Ind Crops Prod. 2018; 120:180–186.
- 75. Xing C, Qin C, Li X, Zhang F, Linhardt RJ, Sun P, Zhang A. Chemical composition and biological activities of essential oil isolated by HS-SPME and UAHD from fruits of bergamot. Lwt-Food Sci Technol. 2019; 104:38–44.
- 76. Mello AM, Gomes RT, Lara SR, Gomes Silva L, Alves JB, Cort´es ME, Abreu SL, Santos VR. The effect of brazilian propolis on the germ tube formation and cell wall of *Candida albicans*. Pharmacol Online. 2006; 3:352–358.
- 77. Viriato A. Terpenoids with antifungal activity for Candida Berkhout, causing nosocomial infections. World Health. 2014; 38:40–50.
- 78. Hossain F, Follett P, Dang Vu K, Harich M, Salmieri S, Lacroix M. Evidence for synergistic activity of plant-derived essential oils against fungal pathogens of food. Food Microbiol. 2016; 53:24–30.
- 79. Nikkhah M and Hashemi M. Boosting antifungal effect of essential oils using combination approach as an efficient strategy to control postharvest spoilage and preserving the jujube fruit quality. Postharv Biol Technol. 2020; 164:111159.
- 80. Daferera DJ, Ziogas BN, Polissiou MG. The effectiveness of plant essential oils on the growth of *Botrytis cinerea*, *Fusarium* sp. and Clavibacter michiganensis subsp. michiganensis. Crop Prot. 2003; 22:39–44.
- 81. Sumalan RM, Alexa E, Poiana MA. Assessment of inhibitory potential of essential oils on natural mycoflora and Fusarium mycotoxins production in wheat. Chem Cent J. 2013; 7:32.
- 82. Rguez S, Msaada K, Daami-Remadi M, Chayeb I, Rebey IB, Hammami M, Laarif A, Hamrouni-Sellami I. Chemical composition and biological activities of essential oils of *Salvia officinalis* aerial parts as affected by diurnal variations. Plant Biosyst. - Int J Deal With All Asp Plant Biol. 2018; 1126:1724–5575.
- 83. Stangarlin JR and Pascholati SF. Activities of ribulose-1,5 bisphosphate Carboxylaseoxygenase (rubisco), chlorophyllase, β-1,3 glucanase and Chitinase and Chlorophyll Content in Bean Cultivars (*Phaseolusvulgaris*) infected with *Uromycesappendiculatus*. Sum. Phytopathol. 2000; 26 (1):34–42.
- 84. Lucas GC, Alves E, Pereira RB, Perina FJ, Magela de Sauza R. Antibacterial activity of essential oils on *Xanthomonas vesicatoria* and control of bacterial spot in tomato. Pesq AgropecBras. 2012; 47(3):351-359.
- 85. Banani H, Olivieri L, Santoro K, Garibaldi A, Gullino ML, Spadaro D. Thyme and savory essential oil efficacy and induction of resistance against *Botrytis cinerea* through priming of defense responses in apple. Foods. 2018; 7(2):11.
- 86. Hu Y, Zhang J, Kong W, Zhao G, Yang M. Mechanisms of antifungal and anti-aflatoxigenic properties of essential oil derived from turmeric (*Curcuma longa* L.) on *Aspergillus flavus*. Food Chem. 2017; 220:1–8.
- 87. Raveau R, Fontaine J, Sahraoui LA. Essential oils as potential alternative biocontrol products against plant pathogens and weeds: A review. Foods. 2020; 9(365):31.
- 88. Lagrouh F, Dakka N, Bakri Y. The antifungal activity of Moroccan plants and the mechanism of action of secondary metabolites from plants. J Mycol Med. 2017; 27(3):303–311.
- 89. Nazzaro F, Fratianni F, Coppola R, De Feo V. Essential oils and antifungal activity. Pharm. 2017; 10(86):1–20.
- 90. Grande-Tovar CD, Chaves-Lopez C, Serio A, Rossi C, Paparella A. Chitosan coatings enriched with essential oils: Effects on fungi involve in fruit decay and mechanisms of action. Trends Food Sci Technol. 2018; 78:61–71.
- 91. Caccioni RLD and Guizzardi M. Inhibition of germination and growth of fruit and vegetable postharvest pathogenic fungi by essential oil components. J Essential Oil Res. 2011; 6(2):173–179.
- 92. Rguez S, Ben Slimene I, Abid G, Hammemi M, Kefid A, Elkahoui S, Ksouri R, Hamrouni Sellami I, Djébali N. *Tetraclinis articulata* essential oil reduces *Botrytis cinerea* infections on tomato. Sci Horticul. 2020; 266:109291.
- 93. Bevilacqua A, Speranza B, Perricone M, Sinigaglia M, Corbo MR. Bioactivity of essential oils towards fungi and bacteria: mode of action and mathematical tools. Essential Oils in Food Proc: Chem Saf Applic. 2017; 231–246.
- 94. He J, Wu D, Zhang Q, Chen H, Li H, Han Q, Lai X, Wang H, Wu Y, Yuan J, Dong H, Qin W. Efficacy and mechanism of cinnamon essential oil on inhibition of *Colletotrichum acutatum* isolated from "Hongyang" kiwifruit. Front Microbiol. 2018: 9:1–12.
- 95. Sharma N and Tripathi A. Fungitoxicity of the essential oil of *Citrus sinensis* on post-harvest pathogens. World J Microbiol Biotechnol. 2006: 22:587–593.
- 96. PaulaP, Moreira S, Jacqueline O, Anaíle dos Mares B, Marise MP, Eduardo Micoti G, Marta HFS.Essential oils from *Eucalyptus staigeriana* F. Muell. ex Bailey and *Eucalyptus urograndis* W. Hill ex Maiden associated to carboxymethylcellulose coating for the control of *Botrytis cinerea* Pers. Fr. and *Rhizopus stolonifer* (Ehrenb.: Fr.) Vuill. in strawberries.Ind Crops Prod. 2020; 156:112884.
- 97. Lucini EI, Zunino MP, Lopez ML, Zygadlo JA. Effect of monoterpenes on lipid composition and sclerotial development of *Sclerotium cepivorum* Berk. J Phytopathol. 2006: 154:441–446.
- 98. Zhang Z, Vriesekoop F, Yuan Q, Liang H. Effects of nisin on the antimicrobial activity of d-limonene and its nanoemulsion. Food Chem. 2014; 150:307–312.
- 99. Escamilla-García M, Calderon-Domínguez G, Chanona-Perez JJ, MendozaMadrigal AG, Di Pierro P, García-Almend´arez BE. Physical, Structural, Barrier, and Antifungal Characterization of Chitosan–Zein Edible Films with Added Essential Oils. Int J Mol Sci. 2017: 18 (11).
- 100. Yu D, Wang J, Shao X, Xu F, Wang H. Antifungal modes of action of tea tree oil and its two characteristic components against *Botrytis cinerea*. J Appl Microbiol. 2015; 119(5):1253–1262.
- 101. Swamy MK, Akhtar MS, Sinniah UR. Antimicrobial properties of plant essential oils against human pathogens and their mode of action: An updated review. Evid-Based Compl Altern Med.2016;2016:3012462.
- 102. Atares L and Chiralt A. Essential oils as additives in biodegradable films and coatings for active food packaging. Trends Food Sci Technol. 2016; 48:51–62.
- 103. Mbili NC, Opara UL, Lennox CL, Vries FA. Citrus and lemongrass essential oils inhibit *Botrytis cinerea* on 'Golden Delicious', 'Pink Lady' and 'Granny Smith' apples. J Plant Dis Prot. 2017; 124(5):499–511.
- 104. Sharifi R, Kiani H, Farzaneh M, Ahmadzadeh M. Chemical Composition of Essential Oils of Iranian *Pimpinella anisum* L. and *Foeniculum vulgare* Miller and their Antifungal Activity Against Postharvest Pathogens. J Essent Oil-Bear Plants. 2008; 11:514–522.
- 105. Vilela GR, de Almeida GS, D'Arce MABR, Moraes MHD, Brito JO, da Silva MFDG. Activity of essential oil and its major compound, 1,8-cineole, from *Eucalyptus globulus* Labill., against the storage fungi *Aspergillus flavus* Link and *Aspergillus parasiticus* Speare. J Stored Prod Res. 2009; 45:108–111.
- 106. Kazemi M. Phytochemical Composition, Antioxidant, Antiinflammatory and Antimicrobial Activity of *Nigella sativa* L. Essential Oil. J Essent Oil Bear Plants. 2014; 17:1002–1011.
- 107. Derwich E, Benziane Z, Taouil R, Senhaji O, Touzani M. Aromatic Plants of Morocco: GC/MSAnalysis of the Essential Oils of Leaves of *Mentha piperita*. Adv Environ Biol. 2010; 4(1):80-85.
- 108. Bertoli A, Çirak C, Teixeira da Silva J. Hypericum Species as Sources of Valuable Essential Oils. In: Medicinal and

Aromatic Plant Science and Biotechnology, Global Sciece Books, 2011.

- 109. Meepagala KM, Sturtz G, Wedge DE. Antifungal constituents of the essential oil fraction of *Artemisia dracunculus* L. var. dracunculus. J Agric Food Chem. 2020; 50:6989–6992.
- 110. Gholamnezhad J. Effect of plant extracts on activity of some defense enzymes of apple fruit in interaction with *Botrytis cinerea*. J Integr Agric. 2019; 18(1):115–123.
- 111. Hua L, Yong C, Zhahnquan Z, Boqiang L, Guozheng Q, Shiping T. Pathogenic mechanisms and control strategies of *Botrytis cinerea* causing postharvest decay in fruits and vegetables. Food Qual Saf. 2018; 2:111–119. 112. Bolívar-Anillo HJ, Garrido C, Collado IG. Endophytic microorganisms for biocontrol of the phytopathogenic fungus *Botrytis cinerea*. Phytochem Rev. 2019; 19: 721-740.

© 2024 the authors. This work is licensed under the Creative Commons Attribution 4.0 International License