



Tropical Journal of Natural Product Research


Available online at <https://www.tjnpr.org>

Review Article

Pharmacological Actions of Phytoconstituents on Neurodegenerative Disorders

Abolanle A. A. Kayode^{1*}, Omowumi T. Kayode², Damilare E. Rotimi³¹Department of Biochemistry, School of Basic Medical Sciences, Babcock University, Ilishan-Remo, Ogun State, Nigeria²Department of Biochemistry, College of Basic and Applied Sciences, Mountain Top University, Kilometer 12, Lagos-Ibadan Expressway, Prayer City, Ogun State, Nigeria³Department of Biochemistry, College of Pure & Applied Sciences, Landmark University, Omu-Aran, Kwara State, Nigeria

ARTICLE INFO

Article history:

Received 28 August 2021

Revised 17 June 2022

Accepted 17 July 2022

Published online 03 August 2022

Copyright: © 2022 Kayode *et al.* This is an open-access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Neurodegenerative diseases (NDs) have over the years become a growing cause for concern and it is increasingly becoming a major health problem associated with impairment, ageing of the brain and other neuropathological conditions. These include Parkinson's disease, Dementia, Huntington's disease, Alzheimer disease, and Depression. Due to increase in prevalence of various NDs over the years, there is growing need to provide suitable means by which the disease can be mitigated. Various pharmacological agents are being researched on to investigate the most suitable and effective means to curb these diseases. This review was aimed at perusing therapeutic potentials of various pharmacological agents in the treatment of NDs. Over a hundred research papers, from Pubmed, NCBI, BMC med, Neurosci etc published on NDs treatments via phytochemicals within the past two decades were analysed. This review will help guide research on pharmacological agents with a wider range treatment options for NDs. Curcumins were most frequently studied for all NDs management. Other commonly used pharmacological agents are resveratrol, epigallocatechin n-gallate (ECGC) and l-theanine. In the management of NDs, curcumin is found to normalize altered mechanisms associated with caspase-3 level, TNF- α , COX-2, NO, iNOS, PGE2, IL-6, Glial fibrillary acidic protein (GFAP), IL-1 β , insulin-degrading enzymes, apoptosis, inflammation, mitochondria dysfunction, inhibitory activity of AChE, and cell viability. It was surmised from the study that curcumin, resveratrol, ECGC and L-theanine are the most common and effective classes of phytochemicals for a wide range of NDs management.

Keywords: Neurodegenerative disease, pharmacological agents, phytochemicals, curcumin, resveratrol.

Introduction

The human brain is the most complex organ of the body. It possesses a vast array of billions of neurons which work intricately to carry out various vital functions. Such functions include receiving and sending nerve impulses. It also functions in ensuring optimum neuron functioning by providing support for glial cells. By carrying out all these functions it is able to perform behavioural and cognitive activity. Once there is alteration in the normal functioning of the brain, this will lead to disease that could affect the normal way of life of an individual.¹ This alteration could be due to inability to maintain a constant energy supply required for the brain to perform its function. It could also be as a result of damage to the supporting glia cells. Any of these can lead to brain dysfunction and also result in behavioural lapse. Any disease as a result of this is referred to as Neurodegenerative disease which includes Parkinson's disease, Alzheimer's disease (AD), seizure disorders, dementia and depression. Neurodegenerative disease has been a prominent cause of concern when it comes to neuropathological conditions and also brain ageing. It has a death rate of about 10% and has been identified as a prominent cause of death worldwide.¹

*Corresponding author. E mail: kayodeab@babcock.edu.ng;
naturalproductpharmacist@gmail.com
Tel: +2348038272670

Citation: Kayode AAA, Kayode OT, Rotimi DE. Pharmacological Actions of Phytoconstituents on Neurodegenerative Disorders. Trop J Nat Prod Res. 2022; 6(7):1019-1046. doi.org/10.26538/tjnpr/v6i7.2

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

Neurodegenerative diseases have been a major health problem involved in brain ageing and neuropathological conditions. They have been noted as a prominent cause of death worldwide, with a death rate of about 10%.² Some common examples of cognitive dysfunction and neurodegenerative disorders include Parkinson disease, Alzheimer's Disease (AD), seizure disorders, dementia and depression.³ This brings about the need for neuroprotection. Neuroprotection are ways or mechanisms by which the brain can be protected from neuronal injury due to neurodegenerative disorders and also ensure viability and optimum functioning of the brain.⁴ The need for neuroprotection has led to research in prospective candidates that could provide protection for the brain. Plants are one of the investigated candidates for neuroprotection. Plants have been an essential need for humans as they are not only important as food source alone but also serves as medicines. Previously, there have been intense interests in plant or herbal medicines in response to various health problems. Significant health benefits have been associated to vegetables and fruits rich diets as they boost the functionality of the body and brain due to the presence of phytochemicals present in them.⁵ Studies revealed that these phytochemicals can decrease the risk and symptoms of diseases including cancers, stroke, cardiovascular disease and neurodegenerative diseases.^{6,7} Over time, the intrinsic antioxidant properties of the phytochemical have been attributed to their therapeutic effects due to the fact that oxidative stress causes most of the chronic age-related diseases.⁵ These phytochemicals when consumed in adequate amount has been shown to have beneficial effects on health. This also can be applied to neurodegenerative diseases. Therefore, the risk of neuronal dysfunction can be reduced by the increased consumption of vegetables and fruits.^{8,9}

Parkinson's Disease

Parkinson's disease (PD) is the commonest neurodegenerative disease after Alzheimer disease and affects over 5.7 million people worldwide majorly aged adults.¹⁰ Parkinson's disease causes impairment of the cognitive function and motor skills due to the progressive deterioration of dopaminergic neurons of the substantia nigra (SN) pars compacta and of nerve fibres projecting to the striatum, significantly decreases the level of the neurotransmitter responsible for neural modulation which is dopamine.¹¹ Some symptoms associated with PD disorder include autonomic dysfunction, anxiety, resting tremor, bradykinesia, postural instability, cognitive dysfunction rigidity and depression.¹² Several pathogenic mechanisms that mediates damages to dopaminergic neurons include oxidative stress, inflammation, apoptosis, mitochondrial dysfunction and transition metal accumulation.^{11,13}

Till date, despite the effort to manage PD, the therapeutic approaches administrated are just able to alleviate symptoms. The pharmacological therapy for PD is dopamine replacement treatment which is effective for approximately 10 years and reduces efficiently depression, pain and motor symptoms. Although the long-term effect of the drug results in toxic metabolites and Reactive Oxygen Species (ROS) accumulation resulting from dopamine metabolism. Moreover, the disease progression cannot be slowed or halted by the drugs.^{14,15}

Although, the etiology of PD is unknown, numerous studies have revealed different risk factors associated with PD either through interaction or independently, they include environmental factors (pesticides, heavy metals, and herbicides) and genetic factors (leucine-rich repeat kinase 2, parkin and mutations in the α -synuclein).¹⁶ Studies about the neuropathology revealed that the two stages of the disease are dopamine reduction in the SN due to the development of Lewy neuritis (LNs) and Lewy bodies (LBs) and the selective death of dopaminergic neurons in the SN pars compacta.¹⁷ The pathogenesis of PD has been attributed to enhanced oxidative stress and mitochondria dysfunction.¹⁶ Mitochondrial respiratory chain activities are inhibited by oxidative stress resulting in overproduction of aggregated α syn

and ultimately increase ROS levels and mitochondrial dysfunction.¹⁸ A study showed that PD development can be reduced by promoting autophagy which is facilitated by placing aggregates of α syn into the autophagic vesicle and then degrading it.¹⁶ Previous PD animal and *in vitro* models showed that pathogenesis of PD are linked with the apoptosis pathways. Also, it has been observed that the apoptosis process of PD involves significantly the p53/Bcl-2 family members. The promotion of p53-dependent apoptosis activates p38 and c-Jun NH2-terminal kinase (JNK) by suppressing the anti-apoptotic agents (Bcl-2) and activating the caspase pathway which result in neurodegeneration of the disease.¹⁹

Different phytochemicals have been effective in cell and animal models of PD (tables 1 and 2). Some of these phytochemicals with already established anti-parkinsonian effects include polyphenols such as phenols, phenolic acids, flavonoids, flavones, stilbenes, and lignanes as well as terpenes. Others include alkaloids, amino acids, carbohydrates, cinnamates and fatty acid amides and they all had therapeutic effects in PD treatment. Different investigations revealed that majority of the compounds suppressed apoptosis and elevates the cell viability by reducing Bax/Bcl-2 ratio, caspase-3 level and accumulation of α -syn and expression of proinflammatory cytokines (such as NF- κ B, IL-6, IL-1 β , NO, TNF- α , PGE 2) and intracellular transduction and transcription pathways (SIRT and Nrf) and modulation of cellular and nuclear inflammatory signaling. In addition, there are increases in neurotrophic factors, protection of mitochondrial respiratory chain, improvement of antioxidant enzyme, increase in GSH level and decrease in ROS formation and lipid peroxidation, are cellular phytochemicals mechanisms of anti-parkinsonian. In animal-induced PD, therapeutic effect of numerous phytochemicals includes suppression of the apoptosis process and expression and decrease in dopaminergic neuronal loss and depletion of DA. Neuronal inflammation was reduced in TNF- α , iNOS, IL-1 β and COX-2 while increases in CAT, SOD and GPx and oxidative stress were reduced via decrease in lipid peroxidation, MDA and ROS.

Table 1: *in vitro* anti-parkinson activity of different phytochemicals²⁰

Authors	<i>In vitro</i> model	Phytochemical name	Mechanism
Chao <i>et al.</i> ²¹	Rotenone and dieldrin were exposed to SH-SY5Y neuroblastoma cells	L-theanine	↓ caspase-3 activity; ↓ nuclear damage; ↑ Cell viability; ↑ GDNF and BDNF
Kumar <i>et al.</i> ²²	MPTP was exposed to SH-SY5Y neuroblastoma cells	Gastrodin	↓ ROS; ↑ Cell viability; ↓ PARP proteolysis ↓ Bax/Bcl-2;
Du <i>et al.</i> ²³	MPTP was exposed to MES23.5 cells	Rosmarinic acid	↑ caspase-3 activity ↑ Cell viability; ↑ Bcl-2/Bax; ↓ ROS;
Qualls <i>et al.</i> ¹⁴	salsolinol and rotenone was exposed to SH-SY5Y neuroblastoma cells	Curcumin	↓ caspase-3 level ↑ Cell viability;
Ham <i>et al.</i> ²⁴	SH-SY5Y neuroblastoma cells	Moracenin D	↓ α -syn mRNA
Kim <i>et al.</i> ²⁵	MPTP was exposed to rat mesencephalic cells	Acacetin	↓ TNF- α , PGE2 and NO
Mu <i>et al.</i> ²⁶	6-OHDA was exposed to PC12 cells and SH-SY5Y neuroblastoma cells	Baicalein SH-	Improved morphological properties in PC12 cells ↑ cell viability SH-SY5Y cells; ↓ Apoptosis in

Li <i>et al.</i> ²⁷	rotenone was exposed to PC12 cells	Bu-7	↑ Cell viability; ↓ cleaved caspase-3 ↓ p53;
Tai and Truong ²⁸	DDT was exposed to SH-SY5Y neuroblastoma cells	Epigallocatechin 3-Gallate (EGCG)	↑ Cell viability
Tamilselvam <i>et al.</i> ²⁹	Rotenone was exposed to SK-N-SH neuroblastoma cells	Hesperidin	↑ Cell viability; ↑ Bcl-2 ↓ caspase-3 and -9; ↓ cytochrome c release; ↓ Bax; ↑ antiapoptotic performance: protection of MMP; ↑ GSH; ↑ antioxidant performance: ↓ LPO; ↓ ROS
Filomeni <i>et al.</i> ³⁰	SH-SY5Y neuroblastoma exposed to Rotenone	Kaempferol	↓ ROS and mitochondrial carbonyls;
Lee <i>et al.</i> ³¹	Primary cultured neurons exposed to MPTP	Silibinin	no antioxidant effect; ↑ Cell viability; protection of MMP
Park <i>et al.</i> ³²	MPTP was exposed to rat mesencephalic cells	6-Shogaol	↓ TNF- α and NO; ↑ Cell viability
Leal <i>et al.</i> ³³	6-OHDA was exposed to rat mesencephalic cells	Amburoside A	↓ nitrite; ↓ LPO; ↑ Cell viability
Wang and Xu ³⁴	MPTP was exposed to SH-SY5Y neuroblastoma cells	Salvianic acid A	↑ Cell viability; ↓ caspase-3 activity ↓ Cytochrome c release; ↓ Bax/ Bcl-2; ↓ nuclear damage; ↓ ROS; ↓ apoptosis;
Chao <i>et al.</i> ³⁵	6-OHDA was exposed to SH-SY5Y neuroblastoma cells	Resveratrol	↓ caspase-3 activity ↓ LDH;
Chen <i>et al.</i> ³⁶	6-OHDA was exposed to SH-SY5Y neuroblastoma cells	Carnosic acid	↓ cleaved caspase-3 and PARP; ↓ ROS; ↑ Cell viability; ↓ nuclear damage; ↑ Nrf2; ↑ GSH; ↓ JNK and p38 phosphorylation;

Dementia

Dementia is principally an age-related irreversible disease which progressively reduces cognitive functions by reducing the ability to perform daily activities, especially in millions of elderly people suffering from dementia and Alzheimer's disease (AD).³⁷ It has been recorded that there is a new case of dementia every 4 second and 8 million cases documented annually and about 50 million patients worldwide. Dementia is a multifactorial disease (Figure 1) and has been connected to aging and environmental impacts.³⁸

According to National Institute of Health, the different classification of dementia is Alzheimer's disease (AD), frontotemporal dementia (FTD), vascular dementia (VaD), mixed dementias and dementia with Lewy bodies (DLB).³⁹ The commonest dementia worldwide is AD and it accounts for roughly 55-60% of

dementia cases, followed by vascular dementia (VaD) (20%), then dementia with Lewy bodies (DLB) at almost 10% and frontotemporal dementia (FTD) at 2% .⁴⁰

Alzheimer's disease is described as a progressive deterioration of the cholinergic neurons in the cortex and hippocampus areas thereby resulting in damage to the cognitive function and results in symptoms including depression, mental deterioration, memory loss and impaired judgement. The pathological mechanism of Alzheimer's disease are senile plaques, resulting from the amyloid beta ($A\beta$) protein accumulation, aggregated Tau protein and the neurofibrillary tangles (NFTs).⁴¹ Oxidative stress and inflammation are events that results from accumulation of $A\beta$.⁴² In addition, $A\beta$ activates microglia by releasing Reactive Oxygen Species (ROS) and pro-inflammatory cytokines which leads to excitotoxic neuronal death. Typical treatment options for AD are primarily symptomatic among which are

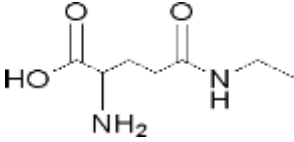
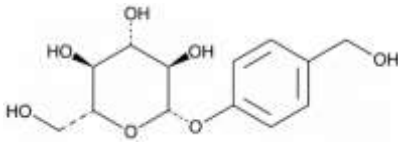
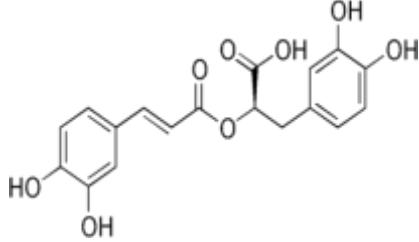
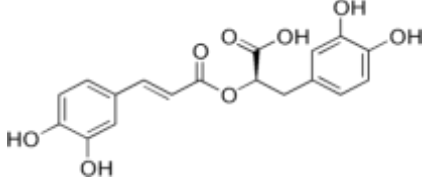
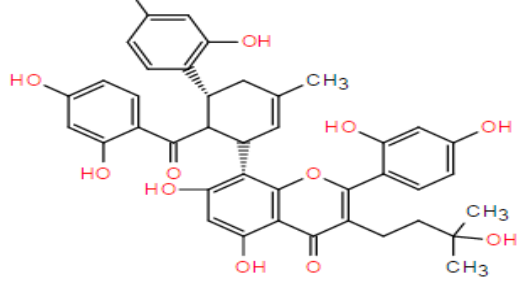
rivastigmine (RIV, galantamine (GAL, Reminyl®), donepezil (Aricept®) and Exelon®). These treatments consist of acetylcholinesterase inhibitors (AChEIs) which enhances cholinergic transmission and improved cognition functioning [43]. Other drug that have proven beneficial effects is memantine (Namenda®), acts as N-methyl-D-aspartate (NMDA) receptor's antagonist.⁴⁴ The VaD is a condition characterized by a cerebrovascular etiology (stroke) of a whole spectrum of cognitive dysfunctions. There have been reported cases of cholinergic deficits in VaD patients and cholinergic treatment shows therapeutic effects on cognitive function.⁴⁵ Treatment options for VaD are aimed at averting further vascular injury by reducing its risk factors such as diabetes mellitus and hypertension.⁴⁶ The DLB is characterized by the abnormal α -synuclein (α -Syn) protein aggregation in Lewy bodies (neuronal cells).⁴⁷ The DLB pathogenetic mechanisms are multifactorial, although formation of

Lewy bodies implicates the genetic mutations in the α -Syn family genes.⁴⁸ It is clinically characterized by slowed movements, sleep disturbances, recurrent visual hallucinations, tremors (Parkinsonism), stiff limbs, and fluctuations in alertness and cognition. Neurodegeneration can be caused by accumulation of α -Syn and therefore results in mitochondrial degeneration which induces oxidative stress.^{49, 50}

The FTD is characterized by progressive atrophy in the temporal or frontal lobes which causes a gradual decline in behavior. It is pathologically and genetically heterogeneous. So far, no therapy has been found but the symptoms can be managed using antipsychotics or antidepressants.⁵¹

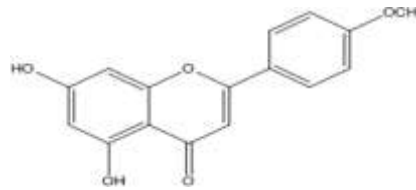
In preclinical studies, phytochemicals such as polyphenols have displayed properties including anti-oxidative, anti-inflammatory and anti-amyloidogenic properties in dementia treatment (table 3).

Table 2: Chemical structures of phytochemicals with anti-parkinson activity

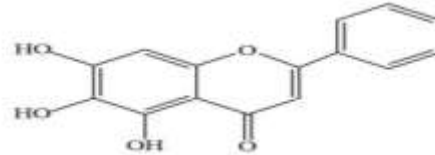
Photochemical name	Category	Structure
L-theanine	Amino acid	
Gastrodin	Benzyl alcohol	
Rosmarinic acid	Cinnamate	
Curcumin	Diarylheptanoid	
Moracenin D	Flavone	

Acacetin

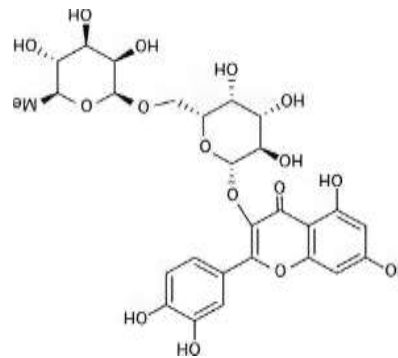
Flavonoid



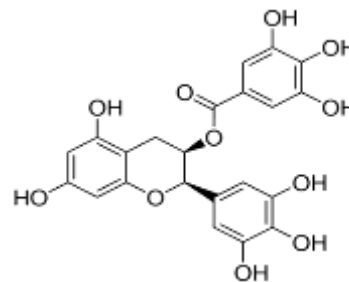
Baicalein SH-



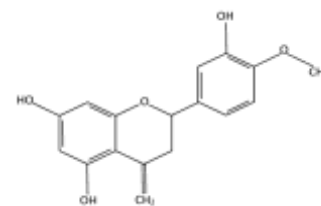
Bu-7



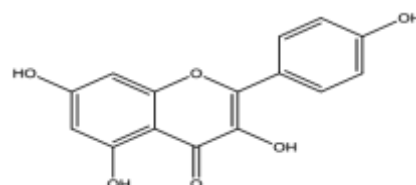
Epigallocatechin 3-Gallate (EGCG)



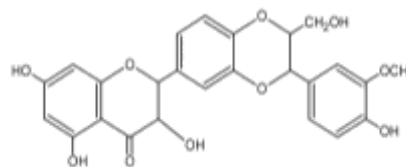
Hesperidin



Kaempferol

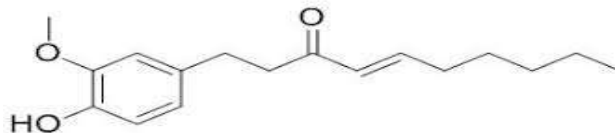


Silibinin



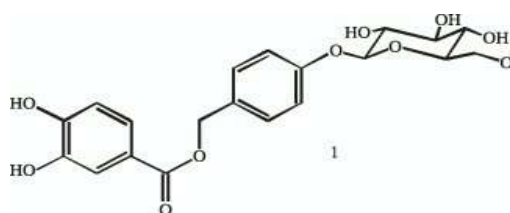
6-Shogaol

Phenol



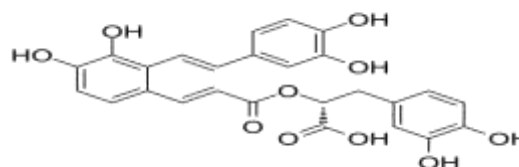
Amburoside A

Phenol glucoside



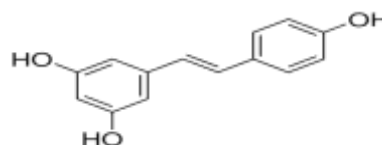
Salvianic acid A

Phenolic acid



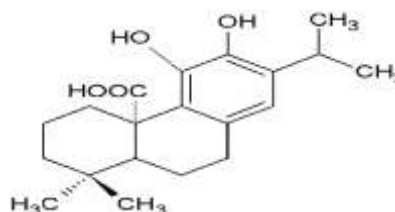
Resveratrol

Stilbene



Carnosic acid

Terpene

**Table 3:** Neuroprotective effects of phytochemicals on dementia in preclinical studies.⁵²

Phytochemical	Model	Proposed Mechanisms Involved (<i>In vitro/In vivo</i>)	Mediated Protective Effects	References
Curcumin	LPS-stimulated rat BV2 microglia	TNF- α , COX-2, IL-1 β , NO, iNOS, PGE2, IL-6	antioxidative, anti-inflammatory	Jin <i>et al.</i> ⁵³
	Mutant APPswe over expression in SH-SY5Y		anti-amyloidogenic	Durairajan <i>et al.</i> ⁵⁴
	Tg2576 mice expressing mutant APP	GFAP, IL-1 β , amyloid plaques	anti-inflammatory, anti-amyloidogenic	Lim <i>et al.</i> ⁵⁵
Resveratrol	PS1/APP double transgenic AD mice	insulin-degrading enzymes	anti-amyloidogenic	Wang <i>et al.</i> ⁵⁶
	A β -induced rat C6 glioma cells	iNOS, PGE2, COX-2, NO	anti-inflammatory	Debprasad <i>et al.</i> ⁵⁷

	A β -induced rat PC12 cells	JNK, Bax, ROS, NF κ B	anti-apoptotic anti-inflammatory	Jang and Surh ⁵⁸
	Healthy rats	MDA, SOD and CAT	Anti-oxidative	Mokni <i>et al.</i> ⁵⁹
	PS1/APP double transgenic AD mice	SIRT-1, AMPK	Anti-oxidative, anti-amyloidogenic	Porquet <i>et al.</i> ⁶⁰
	CCH rats	CREB phosphorylation and PKA	spatial learning and memory improvement	Lonze and Ginty ⁶¹
EGCG	IL-1 β /A β exposed U373MG cells	MAPK, COX2, VEGF, IL-8, PGE, IL-6, NF κ B, JNK	anti-inflammatory	(Kim <i>et al.</i> ⁶²
	icv-STZ rats	AChE, ROS	anti-amyloidogenic anti-oxidative	Biasibetti <i>et al.</i> ⁶³

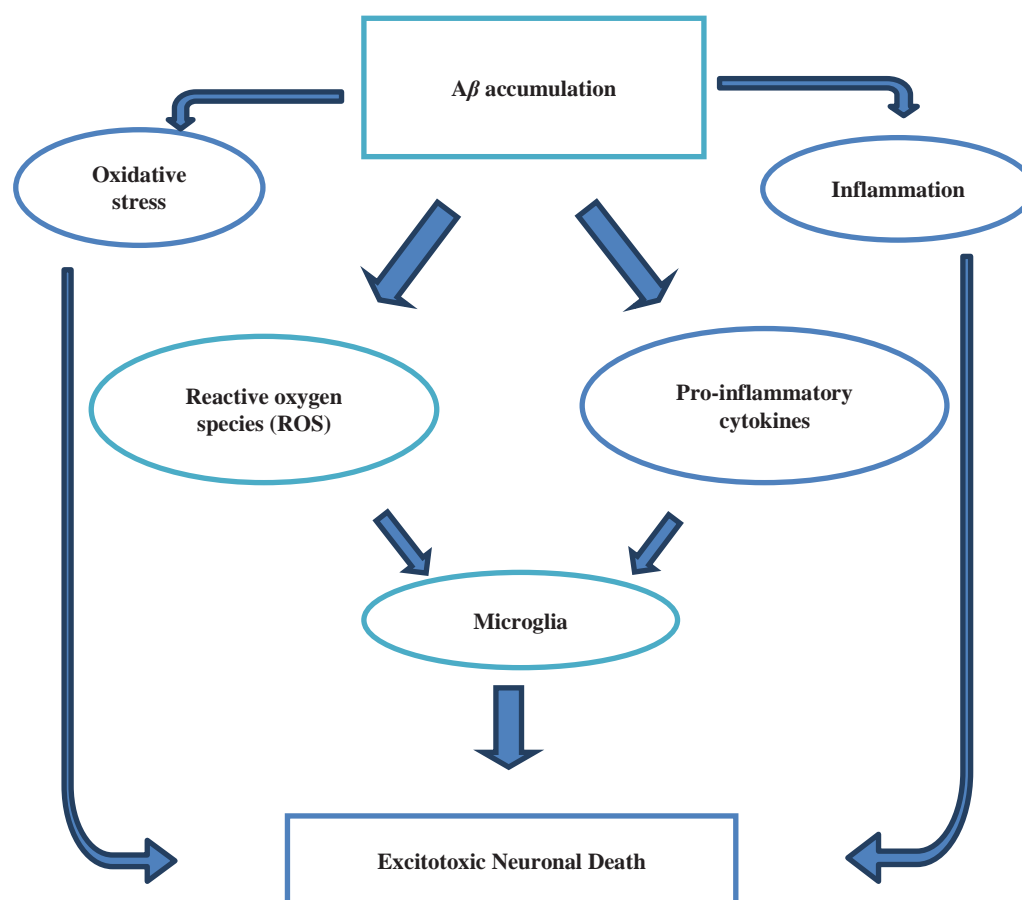


Figure 1: Mechanisms of action of dementia as it relates to inflammation and oxidative stress

Huntington's disease

Huntington's disease (HD) was first described by an Ohio physician, George Huntington. The HD also known as Huntington's chorea is an inherited autosomal dominant neurodegenerative disorder characterized by progressive motor dysfunction affecting motor ataxia, muscle coordination leading to chorea and dystonia, cognitive decline, emotional disturbances, memory, and weight loss.^{64, 65} The pathological alterations mainly involves enkephalin neurons of basal ganglia in HD, GABA loss, N-methyl-D-aspartate (NMDA) receptors modification and the Medium Spiny Neurons (MSNs) of striatum, and cortex.⁶⁶ Out of 100, 000 persons worldwide, HD occur in about 3-6 persons and approximately 20 persons as carriers.⁶⁷ Death of the patients usually occurs 16-20 years after the symptoms appear, these symptoms usually develop between the ages of 37 and 50 in humans.^{68, 69} So far, there has not been a known treatment procedure to check progressive neuronal dysfunction; symptom management has

been through conventional therapeutic approach.⁷⁰ Presently, non-pharmacological treatments of HD involve palliative care and gene therapy.⁶⁸

Early symptoms management reported as efficacious are mood stabilizers and Selective Serotonin Reuptake Inhibitors (SSRIs).^{66, 65} Series of cellular and animal models have been used to mimic this condition. According to WHO, there is reported over-dependence on traditional medicine. It has been popularized due to its lower cost, reported therapeutic efficacy against a number of ailments, its availability and lesser side effects.⁷¹ Characterization of active constituents of these herbs may bring about new drug discoveries.^{71, 72} Recently, considerable attention has been given to the use of herbs to treat neurodegenerative diseases. Some of the effective phytochemicals that may ameliorate the condition are listed in table 4 and there structures are shown in table 5.

However, further investigation is required to understand the safety, efficacy and tolerability of many herbal formulations.⁷³

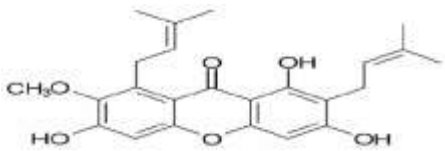
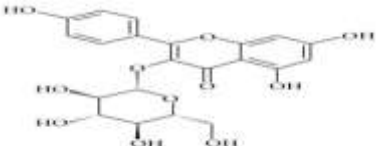
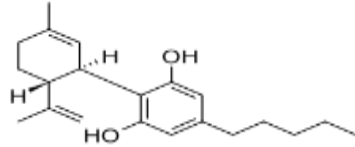
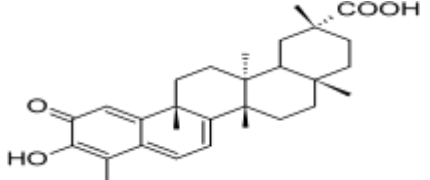
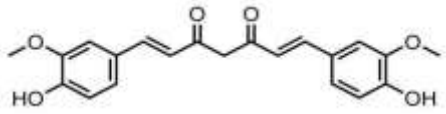
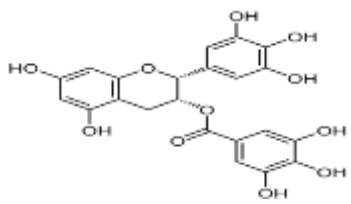
Table 4: Phytochemicals with anti-HD activity

Authors	Models	Plant source	Compound	Mechanism of action
Pedraza	3-NP, CGNs	<i>Garcinia</i>	α -mangostin	Enhanced antioxidant activities, ↓ROS production
Chaverrí <i>et al.</i> ⁷⁴		<i>mangostana</i> L.		
Zhang <i>et al.</i> ⁷⁵	mutant poly Q, <i>C. elegans</i>	<i>Astragalus membranaceus</i>	astragalin	↑adult lifespan, ↑lifespan of daf-2 and age-↓polyQ aggregation
Sagredo <i>et al.</i> ⁷⁶	3-NP, rats	<i>Cannabissativa</i> L.	cannabidiol	antioxidant, ↓striatal atrophy
Zhang and Sarge ⁷⁷	mutant polyQ, HeLa and PC12 cells	<i>Tripterygium wilfordii</i>	celastrol	Modulation of HSP
Sandhir <i>et al.</i> ⁷⁸	3-NP, rats	<i>Curcuma longa</i> L.	curcumin encapsulated solid lipid nanoparticles (C-SLNs)	↓mitochondrial dysfunction
Ehrnhoefer <i>et al.</i> ⁷⁹	polyQ-mediated htt protein, HD yeast;	<i>Camellia sinensis</i> (L.) <i>Kuntze</i>	(-)- epigallocatehi n-gallate	↓photoreceptor degeneration, ↓cytotoxicity ↑motor function ↓mutant Httex1 protein aggregation
Maher <i>et al.</i> ⁸⁰	PC12 cells expressing mutant Httex1	<i>Many plants</i>	fisetin	↑ERK activation ↓mHtt,
Park <i>et al.</i> ⁸¹	3-NP, rats	<i>Galanthus sp.</i>	galantamine	↓striatal lesion, nAChR modulation, anti-apoptotic
Wu <i>et al.</i> ⁸²	Striatal MSNs from YAC128 HD mouse	<i>Panax ginseng</i> C.A.	ginsenosides (Rb1, Rc, and Rg5)	↓glutamate-induced Ca(2+) responses
Menze <i>et al.</i> ⁸³	3-NP, rats	<i>citrus fruits</i>	hesperidin	↑cortical, striatal and hippocampal MDA levels, antioxidant, anti-inflammatory, prevented change in locomotor activity
Lagoa <i>et al.</i> ⁸⁴	3-NP, rats	<i>many plants</i>	kaempferol	↓striatal lesions, antioxidant ↓mortality, ↓motor deficit,
Binawade and Jagtap ⁸⁵	3-NP, rats	<i>many plants</i>	lutein	↑neurobehavioral improvement, ↑body weight, ↑mitochondrial enzymes complex activities, antioxidant
Kumar <i>et al.</i> ⁸⁶	3-NP, rats	<i>tomatoes, other red fruits and vegetables</i>	lycopene	NO modulation, ↑behavioral and biochemical activities
Túnez <i>et al.</i> ⁸⁷	3-NP, rats	<i>many plants</i>	melatonin	antioxidant
Kumar and Kumar ⁸⁸	3-NP, rats	<i>citrus fruits and others</i>	naringin	↓mitochondrial enzymes complex dysfunction, antioxidant, NO modulation ↓behavioral alterations,
Tariq <i>et al.</i> ⁸⁹	3-NP, rats	<i>Nicotiana tabacum</i> L.	nicotine	↓depletion of striatal DA and GSH
Wu <i>et al.</i> ⁹⁰	mHtt, PC12 cells	<i>Radix Polygalae</i>	onjisaponin B	↑autophagy via the AMPK-mTOR signaling
Sandhir and Mehrotra ⁹¹	3-NP, rats	<i>many plants</i>	quercetin	↓oxidative stress, ↓neurobehavioral deficits ↓mitochondrial dysfunctions,
Maher <i>et al.</i> ⁸⁰	PC12 cells	<i>red grapes</i>	resveratrol	↑ERK activation

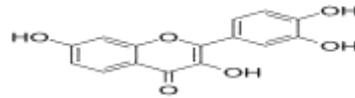
	expressing mutant Httex1;	<i>and others</i>		
Pérez-De La Cruz <i>et al.</i> ⁹²	3-NP, rat brain	<i>Allium sativum L.</i>	S-allylcysteine	↓mitochondrial dysfunction, ↓lipid peroxidation, antioxidant
Lam and Ko ⁹³	3-NP, rat PC12 cells	<i>Schisandra chinensis</i>	(-)-schisandrin B	anti-apoptotic, anti-necrotic
Kumar <i>et al.</i> ⁹⁴	3-NP, rats	<i>Sesamum indicum, L.</i>	sesamol	antioxidant, ↑free radical scavenging activity
Sarkar <i>et al.</i> ⁹⁵	COS-7 and PC12 cells expressing	<i>many plants</i>	trehalose	↑autophagy against mHtt

↑: increased/improved/enhanced ↓: decreased/declined/attenuated

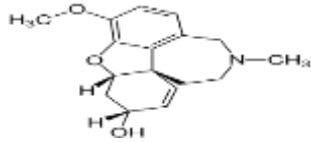
Table 5: Chemical structures of phytochemicals with anti-HD activity

Compound	Chemical Structure
α -mangostin	
Astragalin	
Cannabidiol	
Celastrol	
Curcumin	
epigallocatechin n-gallate	

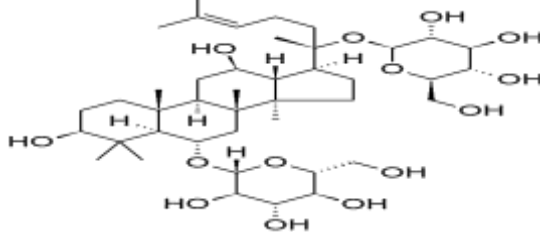
Fisetin



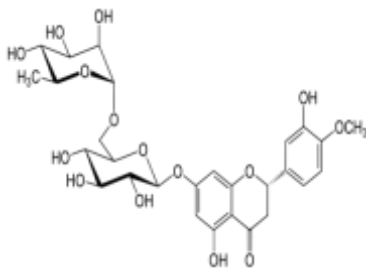
galantamine



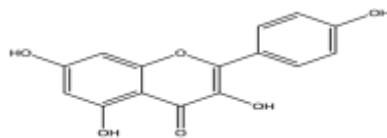
ginsenosides



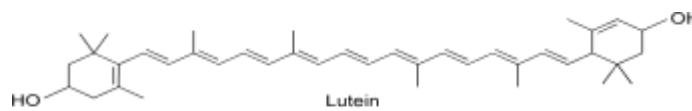
Hesperidin



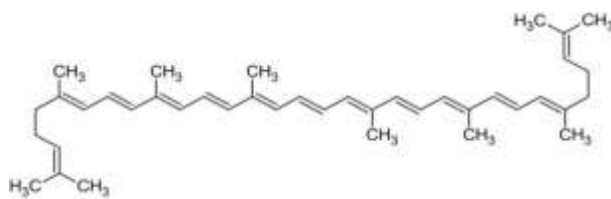
kaempferol



Lutein



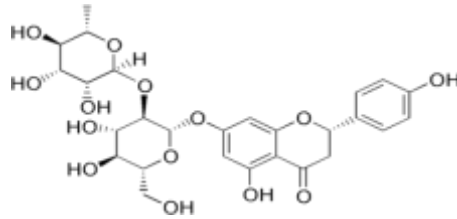
lycopene



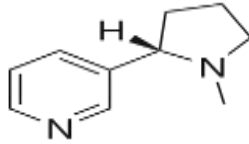
melatonin



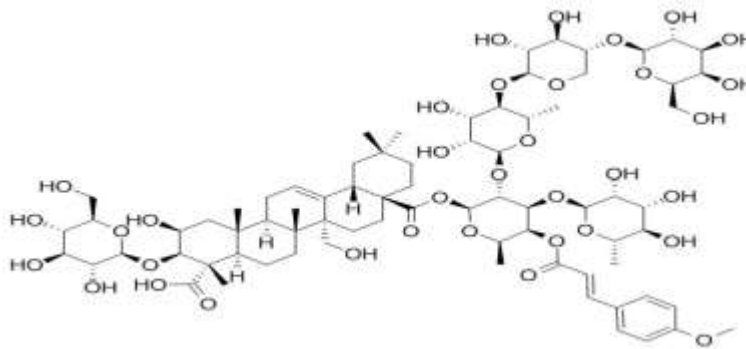
naringin



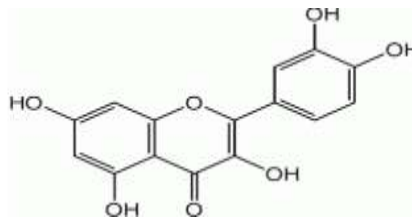
nicotine



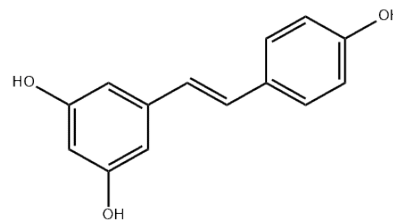
onjisaponin B



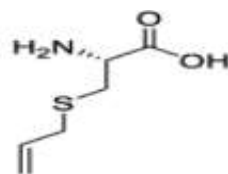
quercetin



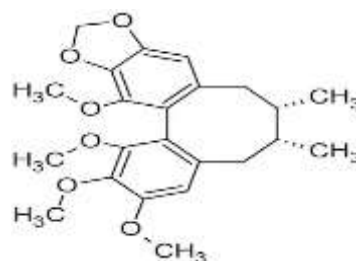
resveratrol



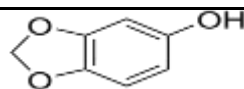
S-allylcysteine



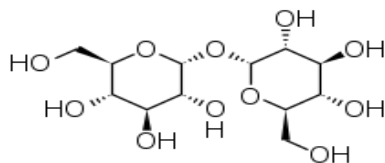
(-)-schisandrin B



sesamol



trehalose

**Alzheimer disease**

Alzheimer's disease (AD) is a complicated and severe neurodegenerative disease characterized by impairment of cognitive function and memory.⁹⁶ In 1906, a German physician named Alois Alzheimer was the first to describe and diagnose Alzheimer's disease.⁹⁷ According to a worldwide consensus, neurodegenerative diseases prevalence is on the increase.⁹⁸ It is commonly associated with dementia in aged people (> 65 years) and account for up to 70% cases of chronic dementia around the world.⁹⁹ The chances of AD occurrence after the age of 65 has been understood to double every five years.¹⁰⁰ Different epidemiological studies of AD also revealed that women over the age of 85 have a higher rate of developing AD than the men.¹⁰¹

It has three clinical phases categorized as dementia phase, pre-dementia phase and pre-symptomatic phase.¹⁰² Symptoms experienced include difficulty in word fluency, delusional symptoms, loss of short term memory and decreasing level of vocabulary along with urinary incontinence^{103, 104}

So far, AD is still irremediable but present treatment option only offers symptomatic relief by reducing cognitive impairment associated with AD through temporary palliative therapy.^{105, 1-6} Enzyme

inhibition of acetylcholinesterase (AChE) is a major therapeutic approaches providing symptomatic effects and in advanced stages of AD, Memantine can be successfully combined as treatment.¹⁰⁷ Although this combined treatment has been approved as anti-AD drugs but there are adverse side effects.⁹⁹ Lack of extensive knowledge on pathogenesis of AD, restricted therapeutic approaches and AD heterogeneity open up new searches for anti-AD treatment effective on AD.¹⁰⁸ Common conventional therapy include vitamin therapy and antioxidant, hormonal therapy, use of NSAIDs and selective phosphodiesterase (PDE) inhibitors.¹⁰⁹⁻¹¹¹

Plants have been proven to be enhancers and promoters of cognitive functions as they act as neuroprotectants either as phytochemicals or in crude extracts¹¹² Presently, rivastigmine and galantamine are plants products licensed as anti-AD drugs. Phytochemicals have been sources of anti-AD treatment and also candidates for synthetic drugs. Some phytochemicals that exhibited anti-AD properties such as MAO inhibitors and antioxidants¹¹³ include resveratrol, curcumin and catechins (Table 6 & 7) and their activity is due to their anti-inflammatory, anti-oxidative and anti-amyloidogenic properties; by triggering neurohormesis⁹⁹ (Figure 2)

Table 6: Phytochemicals with Anti-AD properties

Authors	Models	Source plant	Phytochemical Compound	Mode of action
Elgorashi <i>et al.</i> ¹¹⁴	AChE inhibition	Amaryllidaceae family members	1-O-acetyllycorine	↑ inhibitory activity of AChE
Heo <i>et al.</i> ¹¹⁵	Aβ-induced PC12 cells	<i>Artemisia asiatica</i>	4',5-dihydroxy-3',6,7-trimethoxyflavone	↓oxidative stress ↓ Aβ toxicity
Park <i>et al.</i> ¹¹⁶	Aβ (1-42)-induced BV-2 murine microglia cell lines and murine primary microglia	<i>Schisandra chinensis</i> (Turcz.) Baill. (Schisandraceae) fruit	<i>α</i> -iso-cubebenol	↓neuroinflammation ↓NF-κB/inhibitor of κBα and MAPK
Orhan <i>et al.</i> ¹¹⁷	AChE inhibition	<i>Lycopodium clavatum</i> L.	<i>α</i> -onocerin (atriterpenoid)	↑ inhibitory activity of AChE
Wang <i>et al.</i> ¹¹⁸	Aβ (25-35)- induced SHSY5Y cells	many plants	Acteoside (phenylethanoid glycoside)	↓ROS ↓Aβ toxicity,
Zhao <i>et al.</i> ¹¹⁹	PS1/APP double transgenic AD mice	many plants	apigenin (flavone)	↓oxidative stress ↓Aβ burden, ↑ERK/CREB/BDNF pathway ↑learning and memory,
Wei <i>et al.</i> ¹²⁰	PC12 cells and AβPP/PS1 mice	an active component of the Acorigramineirhizome (AGR)	<i>β</i> -asarone	↑CaMKII- <i>α</i> /p CREB/Bcl-2 pathway ↓apoptosis,
Heo <i>et al.</i> ¹²¹	Aβ-induced PC12 cells	<i>Scutellaria baicalensis</i> (Lamiaceae)	baicalein and baicalin (flavonoids)	↓Aβ-induced toxicity
Urbain <i>et al.</i> ¹²²	AChE inhibition	<i>Gentiana campestris</i> L. (Gentianaceae)	bellidifolin, bellidin	↑ inhibitory activity of AChE
Jiang <i>et al.</i> ¹²³	Aβ (1-42)-induced cortical neuron-glia cultures	<i>Rehmannia glutinosa</i>	catalpol (iridoid glycoside)	↓ Aβ toxicity
Mei <i>et al.</i> ¹²⁴	Aβ42-insulted SH-SY5Y cells	<i>Salvia miltiorrhiza</i>	cryptotanshinone (diterpene)	↓ apoptosis ↓ cytotoxicity
Durairajan	Aβ40 and Aβ42 in N2a	root of <i>Salvia</i>	cryptotanshinone	↓production of Aβ,

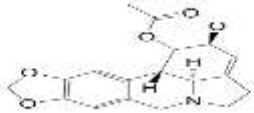
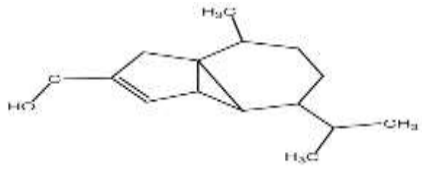
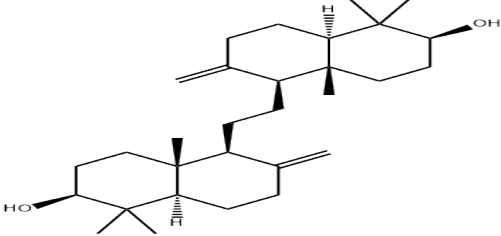
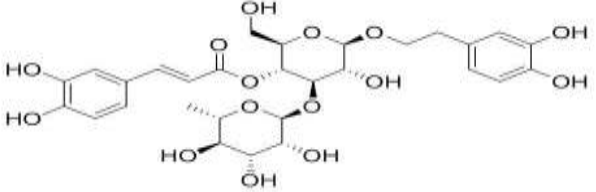
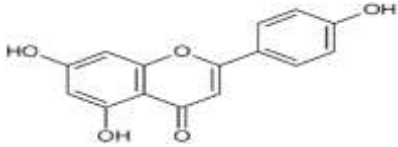
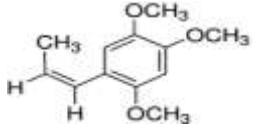
<i>et al.</i> ¹²⁵	mouse neuroblastoma cells	<i>miltiorrhiza</i>		↑activation and translocation of a disintegrin and metalloproteinase-10 (ADAM10)
Park and Kim ¹²⁶	A β -induced PC12 cells	<i>Curcuma longa</i> L. (Zingiberaceae)	curcumin, calebin-A, demethoxycurcumin, 1,7-bis (4-hydroxyphenyl)-1-heptene-3,5-dione	↓ A β toxicity
Wang <i>et al.</i> ¹²⁷	A β (1-40)-induced rats	<i>Curcuma longa</i> L. (Zingiberaceae)	curcumin (a diarylheptanoid)	↑GFAP (Glial fibrillary acidic protein) ↑spatial memory
Ahmed and Gilani ¹²⁸	ex vivo and in-vitro (hippocampus and frontal cortex)	<i>Curcuma longa</i> L. (Zingiberaceae)	curcuminoids	↑ inhibitory activity of AChE ↑memory
Ahmed and Gilani ¹²⁹	A β (1-40) + IBA-infused rat model	<i>Curcuma longa</i> L. (Zingiberaceae)	demethoxycurcumin, curcuminoid mixture and demethoxycurcumin	↓ apoptosis ↓inflammation,
Lee <i>et al.</i> ¹³⁰	AChE inhibition, scopolamine induced mice	<i>Cynanchum atratum</i> (Apocynaceae)	cynatoside B	↑anti-amnesic activities ↑anti-AChE,
Yan <i>et al.</i> ¹³¹	A β (1-40)-induced mice	<i>Angelica gigas</i>	decursinol, a coumarin	↓ impairment of memory
Li <i>et al.</i> ¹³²	A β (25-35)-induced PC12 cells	<i>Angelica gigas</i> Nakai	Decursinol	↓A β -induced toxicity ↑MAPK signal, ↑Nrf2 activation ↑free radical scavenging activity, ↑efficacious against AD
Aronson <i>et al.</i> ¹³³	patients with AD	<i>Galanthus</i> sp.	galantamine (hydrobromide), alkaloid	
Suh <i>et al.</i> ¹³⁴	Korean population with mild to moderate AD	<i>Galanthus</i> sp.	galantamine (hydrobromide), alkaloid	↑behavioral symptoms ↑cognitive function
Zeng <i>et al.</i> ¹³⁵	A β (25-35)-induced cultured hippocampal neurons	<i>Glycine max</i> (L.) Merr. (Fabaceae)	genistein (isoflavone)	↓ apoptosis, ↑antioxidation
Bate <i>et al.</i> ¹³⁶	amyloid-beta1-42 induced SH SY5Y neuroblastoma cells	EGb 761 extract from <i>Ginkgo biloba</i> L. leave	ginkgolides A or B	↓neurotoxicity ↓caspase-3 ↓prostaglandin E2
Wang <i>et al.</i> ¹³⁷	A β (25-35)-induced PC12 cells	<i>Valeriana amurensis</i> P.	heishuixiecaoline A-C	↓A β -induced toxicity
Xiao <i>et al.</i> ¹³⁸	A β (25-35)-induced cells	<i>Huperzia serrata</i>	huperzine A (sesquiterpene)	↓caspase-3, ↓ apoptosis, ↓ROS,
Zeng <i>et al.</i> ¹³⁹	A β (25-35)-induced PC12 cells	<i>Epimedium brevicornum</i>	icariin (flavonoid)	↓GSK-3 β , ↑PI3K/Akt, ↓tau protein hyperphosphorylation
Xian <i>et al.</i> ¹⁴⁰	A β (25-35)-induced rats	<i>Uncaria rhynchophylla</i>	isorhynchophylline	↓tau protein hyperphosphorylation ↓apoptosis, and ↑phosphorylation of phosphatidylinositol 3-kinase

				(PI3K) substrate Akt (PI3K/Akt) ↓glycogen synthase kinase 3β (GSK-3β) activity, ↓Aβ toxicity, ↓ERK, ↓ NF-κB ↓p38 ↓BACE1 expression
Kim <i>et al.</i> ¹⁴¹	Aβ (1-42)- induced mice	<i>Camellia sinensis (L.) many plants</i>	l-theanine (an amino acid)	
Zheng <i>et al.</i> ¹⁴²	SH-SY5Y cells		luteolin (bioflavonoid)	
Heo <i>et al.</i> ¹⁴³	Aβ induced PC12 cells, scopolamine induced mice	<i>Citrus junos</i>	naringenin (flavanone)	↑anti-amnesic activity, ↑antioxidation ↓oxidative stress, ↓ Aβ toxicity,
Ma <i>et al.</i> ¹⁴⁴	PC12 neuronal cells incubated with Aβ(25-35)	<i>Panax notoginseng</i>	notoginsenoside R1	↑cell viability, ↓apoptosis, ↓ MAPK signaling ↑ mitochondrial membrane potential,
Kim <i>et al.</i> ¹⁴⁵	Aβ-induced PC12 cells	many plants	piceatannol	↑antioxidation, ↓PARP cleavage ↓activation of caspase-3 ↓DNA fragmentation ↑anti apoptotic
Yoon <i>et al.</i> ¹⁴⁶	Aβ (25-35)-induced PC12 cells	<i>Cornus officinalis</i>	p-coumaric acid	↓NF-κB activity ↓ c-Jun N terminal kinase (JNK) phosphorylation, ↓ERK1/2
Chonpatho mpikunlert <i>et al.</i> ¹⁴⁷	AD rat models induced by AF64A	<i>Piper nigrum L. (Piperaceae)</i>	piperine (alkaloid)	↑neurotrophic effect ↓lipid peroxidation, ↑anti-AChE
Li <i>et al.</i> ¹⁴⁸	fAβ 1-40 insulted hBMECs	many plants	quercetin (flavonoid)	↑cell viability, ↑SOD, ↓LDH, ↓ROS
Zhang <i>et al.</i> ¹⁴⁹	Aβ (25-35)- induced SH SY5Y human neuroblastoma cells	<i>Rhodiola rosea L. (Crassulaceae)</i>	salidroside (a glucoside of tyrosol)	↓phosphorylation of JNK and p38 MAPK ↑antioxidant enzymes, ↓oxidative stress, ↓Bax, ↑Bcl-X(L), ↑ MMP
Peng <i>et al.</i> ¹⁵⁰	Aβ (25-35)- induced PC12 cells	<i>Allium sativum L. (Amaryllidace ae)</i>	s-allyl cysteine (SAC)	↓Aβ toxicity ↓caspase-3, ↓ROS, ↓PARP cleavage, ↓memory impairments
Chauhan ¹⁵¹	AD transgenic Swedish double mutant mouse model Tg2576	<i>Allium sativum L.</i>	di-allyl-disulfide (DADS) and s-allyl-cysteine (SAC)	↑anti-amyloidogenic, ↑anti-inflammatory, ↑anti-tangle
Lu <i>et al.</i> ¹⁵²	Aβ (25-35)- induced mice	<i>Silybum marianum (L.)</i>	silibinin (flavonoid)	↓overexpression of iNOS and TNF-α mRNA ↓oxidative stress ↑memory, ↑anti-inflammatory
Ingkaninan <i>et al.</i> ¹⁵³	AChE inhibition	<i>Stephania venosa</i>	stephanine, cyclanoline and N methyl stepholidine	↑ inhibitory activity of AChE
Urbain <i>et al.</i> ¹⁵⁴	AChE inhibition	<i>Gentianella amarelle</i>	triptexanthoside C and other xanthenes	↑ inhibitory activity of AChE
Pan <i>et al.</i> ¹⁵⁵	Aβ (1-42)- induced microglial cells	<i>Tripterygium wilfordii Hook.F</i>	tripchlorolide (T4), an extract	↓neuroinflammation, NF-kappaB and JNK modulation
Hong <i>et al.</i> ¹⁵⁶	Aβ(25-35)- induced PC12 cell.	<i>Cornus officinalis</i>	gallic acid, p coumaric acid, and ursolic acid	↑anti apoptotic ↓DNA fragmentation ↑antioxidation,
Chi <i>et al.</i> ¹⁵⁷	Aβ (25-35)- induced SHSY5Y cells	<i>Xanthoceras sorbifolia</i>	xanthoceraside (triterpene)	↓caspase-3 activity ↓ROS, ↓apoptosis
Yu <i>et al.</i> ¹⁵⁸	Aβ-induced SH	<i>Itoa orientalis</i>	xylocoside G (XG)	↓Aβ-

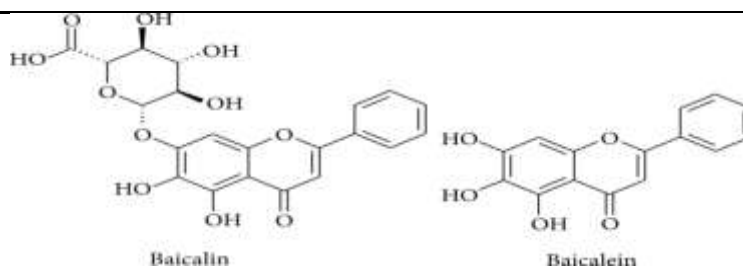
	SY5Y cells	<i>Hemsl.</i>		induced NF- κ B p65 translocation, \downarrow prostaglandin E2 \downarrow interleukin-1 β , \downarrow TNF- α , \downarrow JNK phosphorylation \uparrow anti-AChE
Heo <i>et al.</i> ¹⁵⁹	AChE inhibition in PC12 cells	<i>Fiatoua villosa</i>	zeatin	

\uparrow : increased/improved/enhanced \downarrow : decreased/declined/attenuated

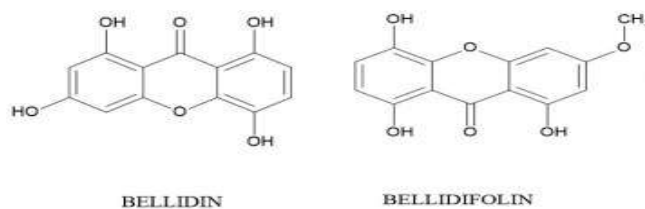
Table 7: Chemical structures of phytochemicals with Anti-AD properties

Phytochemical Compound	Chemical structure
1-O-acetyllycorine	
α -iso-cubebenol	
α -onocerin (a triterpenoid)	
acteoside (phenylethanoid glycoside)	
apigenin (flavone)	
β -asarone	

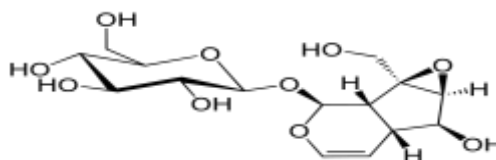
baicalein and baicalin
(flavonoids)



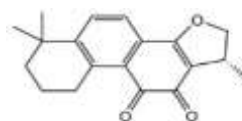
bellidifolin, bellidin



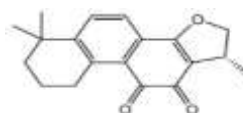
catalpol (iridoid
glycoside)



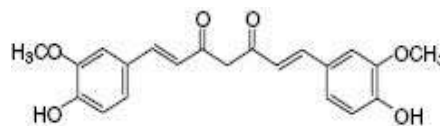
cryptotanshinone
(diterpene)



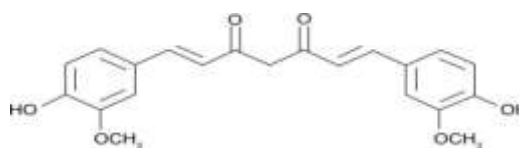
cryptotanshinone



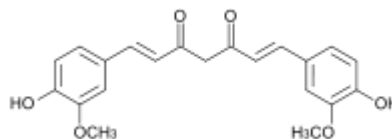
curcumin, calebin-A, demethoxycurcumin,
1,7-bis (4-hydroxyphenyl)-1-heptene-3,5 dione



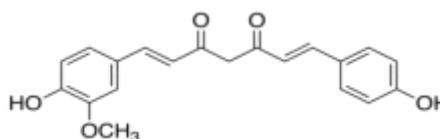
curcumin (a diarylheptanoid)



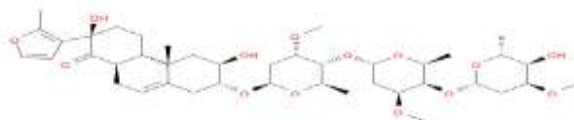
curcuminoids



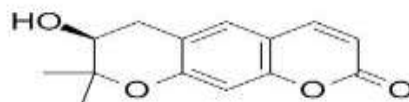
demethoxycurcumin, curcuminoid mixture
and demethoxycurcumin



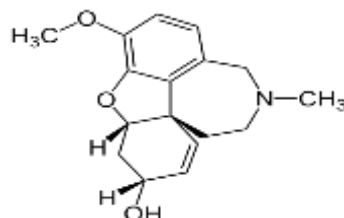
cynatroside B



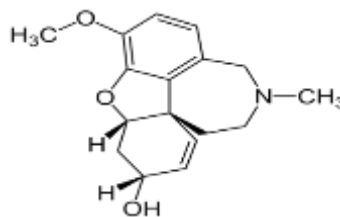
Decursinol



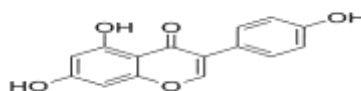
Galantamine (hydrobromide), alkaloid



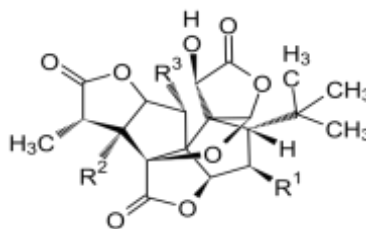
Galantamine (hydrobromide), alkaloid



genistein (isoflavone)

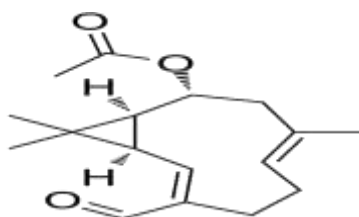


ginkgolides A or B

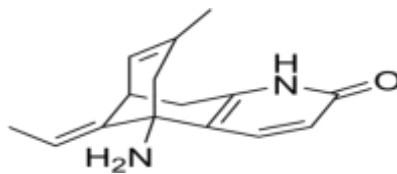


heishuixiecaoline

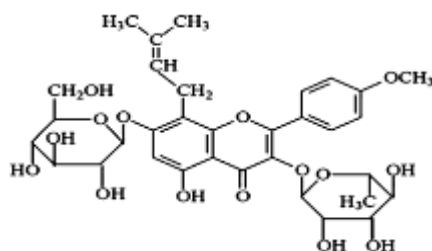
A-C



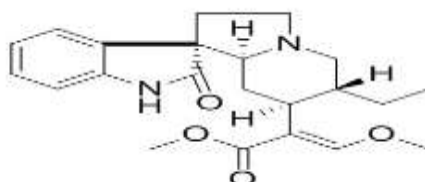
huperzine A
(sesquiterpene)



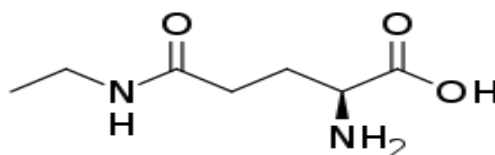
icariin (flavonoid)



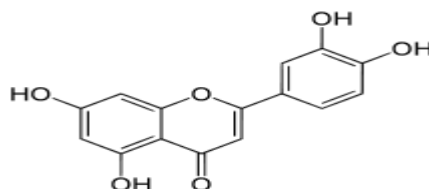
isorhynchophylline



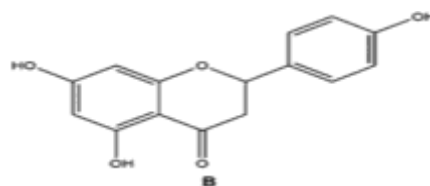
l-theanine (an amino acid)



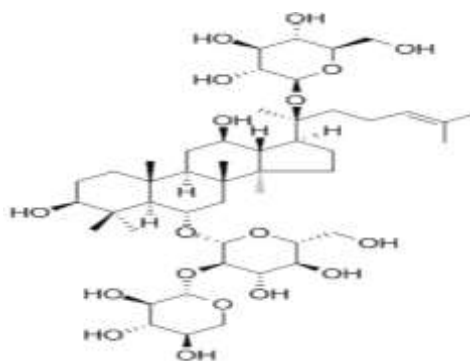
Luteolin (bioflavonoid)



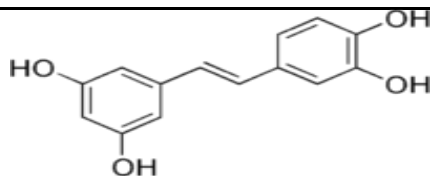
Naringenin (flavanone)



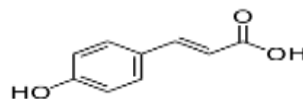
notoginsenoside R1



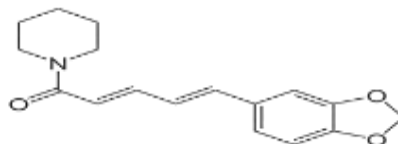
piceatannol



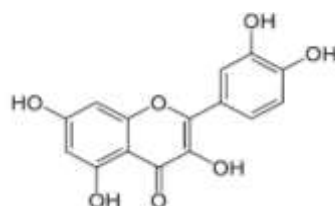
p-coumaric acid



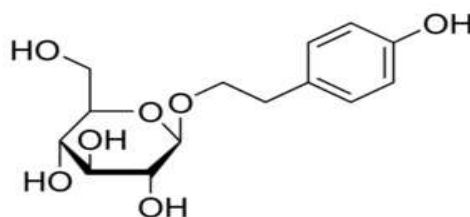
piperine (alkaloid)



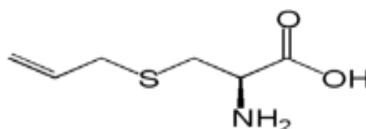
quercetin (flavonoid)



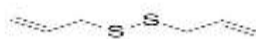
salidroside (aglycoside of tyrosol)



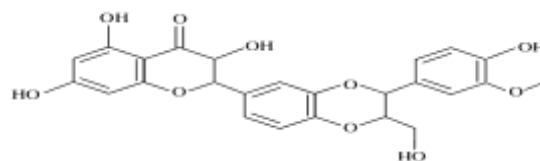
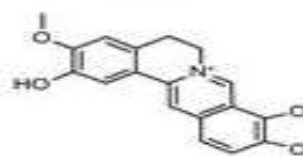
s-allyl cysteine (SAC)



di-allyl-disulfide (DADS) and s-allyl-cysteine (SAC)

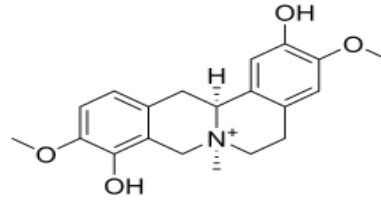
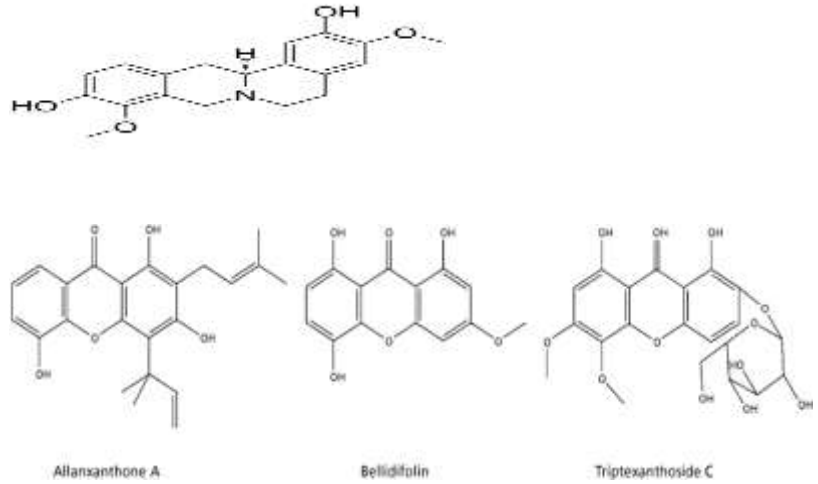


silibinin (flavonoid)

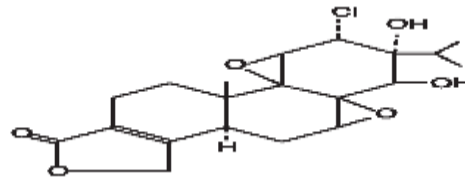
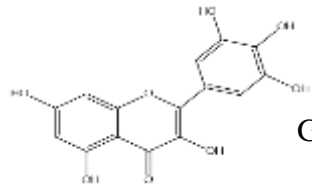
stepharanine, cyclanoline and N
methyl stepholidine

Cyclanoline

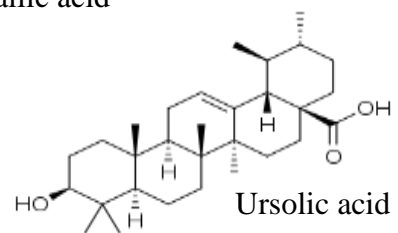
N-methyl stepholidine

triptexanthoside C
and other xanthones

tripchlorolide (T4), an extract

gallic acid,
coumaric acid,
and ursolic acid

Gallic acid

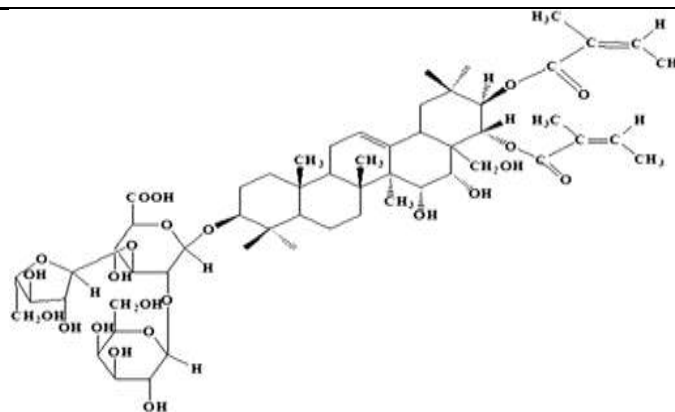


Ursolic acid

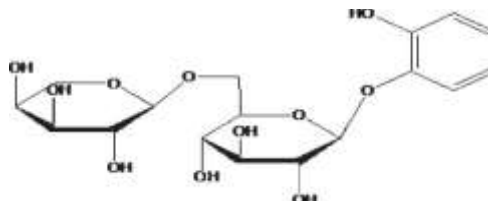


Coumaric acid

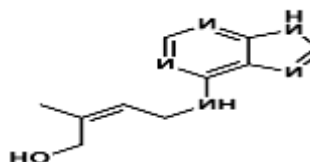
xanthoceraside
(triterpene)



xylocoside G (XG)



zeatin



Depression

Depression is a severe and recurrent mental ailment connected to anorexia, insomnia, incessant sadness, and loss of interest in activities.¹⁶⁰

The etiology of depression involves several factors such as psychological, environmental, social, genetic and biochemical factors. Environment and gene interaction jointly project a person at a greater risk. Common risk factors include trauma and viral infections¹⁶¹ Monoamine hypothesis of depression in which decrease in the brain neurotransmitters level (norepinephrine and serotonin) is assumed as the leading cause of depression; however, recently, the pathophysiology of the disease has now been associated with multiple hormonal and neuronal systems. In general, depression incidence is about 3–10% but higher in patients with chronic disorders.¹⁶²

World Health Organization ranked depression as the fourth leading cause of disability.¹⁶³ Women are twice more probable to suffer from depression during their lifetime. There are three different main therapeutic agents for the disease: monoamine oxidase inhibitors, second-generation and tricyclic antidepressants. Monoamine oxidase (MAO) inhibitors inhibit the enzymatic activities of monoamine oxidase. Phenelzine, isocarboxazid, tranylcypromine and moclobemide are examples of MAO that act as first-line therapy.¹⁶⁴ MAO inhibitors block neurotransmitter's transporter which increase the level of synaptic c and thus enhances neurotransmission.¹⁶⁵

Tricyclic antidepressants are however being replaced by novel antidepressants with fewer adverse effects. Norepinephrine reuptake inhibitors, the selective serotonin reuptake inhibitors, and the serotonin norepinephrine reuptake inhibitors, are examples of new second-generation antidepressants. Despite the development of these conventional drugs, depression treatment still fails to achieve clinical remission in lots of cases.¹⁶⁶

Moreover, many patients still display intolerant or refractory responses with these drugs.¹⁶⁷ Indeed, use of these agents is limited by unexpected side effects and some of them show contradictive outcomes.^{168, 169} On the other hand, conventional antidepressants have a long list of side effects, categorized into sexual dysfunction, central nervous system disorders (fatigue, headaches, sedation, agitation, insomnia), serotonin syndrome, parkinsonism, postural hypotension, blurred vision, gastrointestinal (vomiting and nausea), and weight gain, which cause poor patient compliance that could result in treatment failure.¹⁷⁰⁻¹⁷³ The risk of neurodegenerative diseases and other disorders like autoimmune and cardiovascular diseases have been decreased significantly by the use of phytochemicals obtained from herbs overtime (Table 8). Phytochemicals that showed potent anti-inflammatory and antioxidant properties such as curcumin, carvacrol, ferulic acid, L-Teanine quercetin, proanthocyanidin, and resveratrol have demonstrated their neuroprotective effects thereby improving the symptoms of depression.

Table 8: Phytochemicals effective against Depression

Phytochemical	Treatment	Study design	Effects and mechanisms	Reference
Carvacrol	12.5–50 mg/kg	Oral administration in mice	Induce antidepressant effects that seem to be dependent on an interaction with the dopaminergic brain pathways	Melo <i>et al.</i> ¹⁷⁴
	12.5 mg/kg	Oral administration in rats	Raise 5-HT and dopamine ranges in the hippocampus and prefrontal cortex Influence neuronal activity through modulation of neurotransmitters	Zotti <i>et al.</i> ¹⁷⁵

Curcumin (<i>Curcuma longa</i>)	1.25–10 mg/kg	Oral administration in rats	Reduce immobility time in the forced swimming test Reverse bilateral olfactory bulbectomy-induced hyperactivity in the open field and deficits in step-down passive avoidance	Xu <i>et al.</i> ¹⁷⁶
	20–40 mg/kg	Intraperitoneal injection in mice	Restore biochemical and behavioral changes induced by the chronic stress Reverse the decreased immobility period and MAO activity induced chronic stress	Bhutani <i>et al.</i> ¹⁷⁷
	10 mg/kg	Oral administration in mice	Reduce duration of immobility in forced swimming test May be associated with 5-HT _{1A/1B} and 5-HT _{2C} subtypes	Wang <i>et al.</i> ¹⁷⁸
	10–20 mg/kg	Oral administration in rats	Attenuate the stress-induced hippocampus 5-HT _{1A} mRNA	Xu <i>et al.</i> ¹⁷⁹
Ferulic acid	100–250 mg/kg	Oral administration in mice	Attenuate stress-induced behavior Increase CREB phosphorylation and brain-derived neurotropic factor mRNA level in the hippocampus	Yabe <i>et al.</i> ¹⁸⁰
L-Theanine (<i>Camellia sinensis</i>)	1–20 mg/kg	Oral administration in mice	Reduce immobility time in the forced swimming test and tail suspension test without ambulation in the open field test Antagonize reserpine-induced ptosis and hypothermia	Yin <i>et al.</i> ¹⁸¹
Proanthocyanidin	25–50 mg/kg	Oral administration in mice	Reduce immobility period in the forced swimming test and tail suspension test Enhance 5-HT levels in hypothalamus, hypothalamus, and frontal cortex	Xu <i>et al.</i> ¹⁸²
Quercetin	20–40 mg/kg	Oral administration in mice	Prevent hyperactivation of the HPA axis	(Bhutada <i>et al.</i> ¹⁸³
Resveratrol	20–80 mg/kg	Oral administration in mice	Decrease immobility period in the despair tests without influence on locomotor activity Enhance 5-HT and noradrenaline concentrations in the brain	Xu <i>et al.</i> ¹⁸²
	40–80 mg/kg	Oral administration in rats	Reverse less weight gain, reduce sucrose preference and deficits in the shuttle box Raise 5-HT, dopamine, and noradrenaline concentrations in brain Reduce MAO activity	Yu <i>et al.</i> ¹⁸⁴

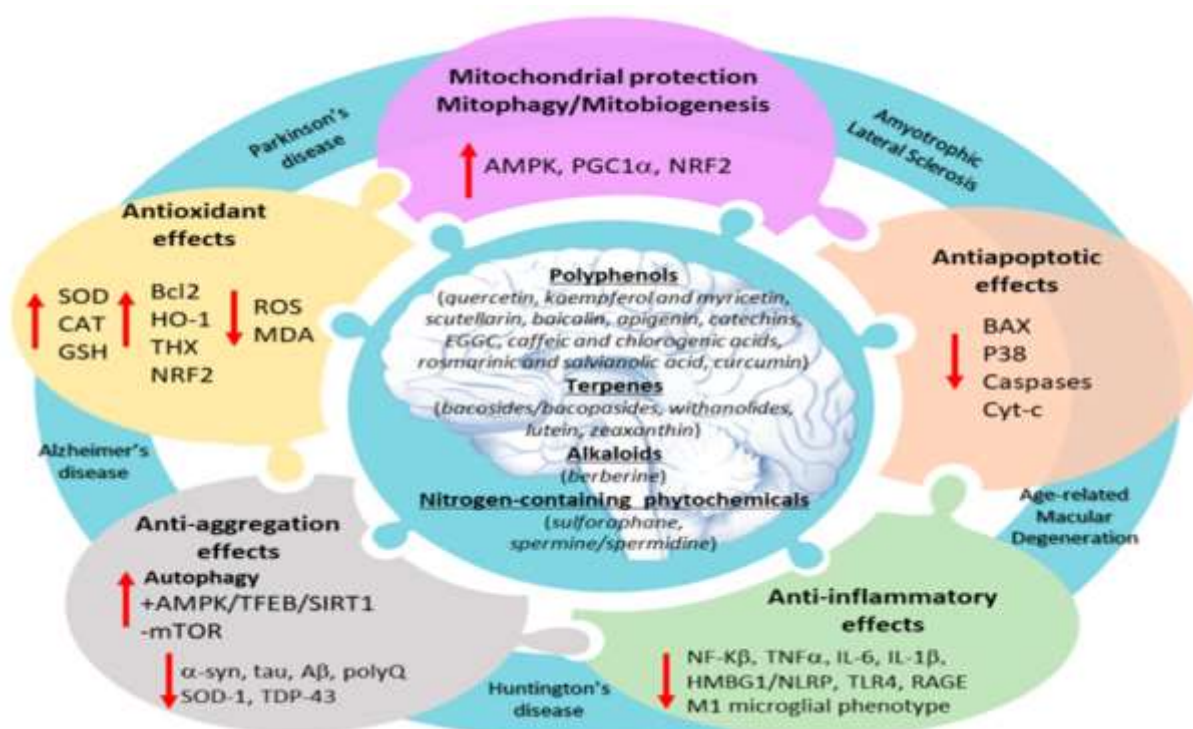


Figure 2: Mechanism of action of phytochemical constituents' interactions with Huntington's disease and other neurodegenerative disorders¹⁸⁵

Conclusion

In this review, it can be deduced that certain phytochemicals possess ameliorative effect towards the NDs management. From this study, curcumin was observed to be effective in the management of a wider range of NDs, normalizing altered mechanisms associated with caspase-3 level, TNF- α , COX-2, IL-1 β , NO, iNOS, PGE2, IL-6, Tau hyperphosphorylation, APP and GSK3 β activity, Glial fibrillary acidic protein (GFAP), IL-1 β , amyloid plaques, insulin-degrading enzymes, apoptosis, inflammation, mitochondria dysfunction, inhibitory activity of AChE, and cell viability. Curcumins, resveratrol, ECGC and L-theanine are found to be the most common and effective classes of phytochemicals for a wide range of NDs management. However, gap in knowledge abound in extensive elucidation of rate-limiting and protein conformational studies of the use of these phytochemicals in NDs management. Therefore it is highly imperative that these promising phytochemicals be researched on extensively in order to come up with more effective treatment for these neurodegenerative diseases.

Conflict of Interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

References

1. Youdim KA and Joseph JA. A Possible Emerging Role of Phytochemicals in Improving Age-Related Neurological Dysfunctions: A Multiplicity of Effects. *Free Rad Biol Med.* 2001; 30:583–594.
2. Kolominsky-Rabas PL, Sarti C, Heuschmann PU, Graf C, Siemonsen S, Neundoerfer B, Katalinic A, Lang E, Gassmann KG, von Stockert TR. A prospective community based study of stroke in Germany. The Erlangen Stroke Project (ESPro): Incidence and case fatality at 1, 3, and 12 months. *Stroke.* 1998; 29:2501-2506.
3. Commenges D, Scotet V, Renaud S, Jacqmin-Gadda H, Barberger Gateau P, Dartigues JF. Intake of flavonoids and risk of dementia. *Eur J Epidemiol.* 2000; 6:357-363.
4. Kumar V. Potential medicinal plants for CNS disorders: An overview. *Phytother Res.* 2006; 20:1023-1035.
5. Lee J, Jo DG, Park D, Chung HY, Mattson MP. Adaptive cellular stress pathways as therapeutic targets of dietary phytochemicals: focus on the nervous system. *Pharmacol Rev.* 2014; 66:815–868.
6. Mattson MP, Son TG, Camandola S. Viewpoint: Mechanisms of action and therapeutic potential of neurohormetic phytochemicals. *Dose-Response.* 2007; 5:174-186.
7. Slavin JL and Lloyd B. Health benefits of fruits and vegetables. *Adv Nutr.* 2012; 3:506-516.
8. Selvam AB. Inventory of Vegetable Crude Drug samples housed in Botanical Survey of India, Howrah. *Pharmacogn Rev.* 2008; 2:61-94.
9. Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacogn Rev.* 2010; 4:118-126.
10. Gopalakrishna A and Alexander SA. Understanding Parkinson disease: a complex and multifaceted illness. *J Neurosci Nurs.* 2015; 47:320-326.
11. Moon HE and Paek SH. Mitochondrial dysfunction in Parkinson's disease. *Exp Neurobiol.* 2015; 24:103-116.
12. Bassani TB, Vital MA, Rauh LK. Neuroinflammation in the pathophysiology of Parkinson's disease and therapeutic evidence of anti-inflammatory drugs. *Arq Neuropsiquiatr.* 2015; 73:616–623.
13. Qualls Z, Brown D, Ramlochansingh C, Hurley LL, Tizabi Y. Protective effects of curcumin against rotenone and salsolinol-induced toxicity: implications for Parkinson's disease. *Neurotox Res.* 2014; 25:81-89.
14. Le Douaron G, Schmidt F, Amar M, Kadar H, Debortoli L, Latini A., Séon-Méniel B, Ferrié L, Michel, PP, Touboul, D, Brunelle, A, Raisman-Vozari R, Figadère B. Neuroprotective effects of a brain permeant 6-aminoquinoxaline derivative in cell culture conditions that model the loss of dopaminergic neurons in Parkinson disease. *Eur J Med Chem.* 2015; 89:467-479.
15. Orme RP, Bhargal MS, Fricker RA. Calcitriol imparts neuroprotection in vitro to midbrain dopaminergic neurons by upregulating GDNF expression. *PLoS ONE.* 2013; 8:e62040.
16. Blesa J, Trigo-Damas I, Quiroga-Varela A, Jackson-Lewis VR. Oxidative stress and Parkinson's disease. *Front Neuroanat.* 2015; 9:91.
17. Agim ZS and Cannon JR. Dietary factors in the etiology of Parkinson's disease. *BioMed Res Int.* 2015. Retrieved from: <https://doi.org/10.1155/2015/672838> (Accessed March 3rd, 2021).
18. Cheung ZH and Ip NY. The emerging role of autophagy in Parkinson's disease. *Mol Brain.* 2009; 2:29.
19. Ren P, Jiang H, Li R, Wang J, Song N, Xu HM, Xie JX. Rosmarinic acid inhibits 6-OHDA-induced neurotoxicity by anti-oxidation in MES23.5 cells. *J Mol Neurosci.* 2009; 39:220–225.
20. Shahpiri Z, Bahramsoltani R, Farzaei MH Farzaei F, Rahimi R. Phytochemicals as future drugs for Parkinson's disease: a comprehensive review. *Rev Neurosci.* 2016. Retrieved from: <https://doi.org/10.1515/revneuro-2016-0004> (Accessed March 6th, 2021).
21. Chao J, Yu MS, Ho YS, Wang M, Chang RC. Dietary oxyresveratrol prevents parkinsonian mimetic 6-hydroxydopamine neurotoxicity. *Free Radic Biol Med.* 2008; 45:1019–1026.
22. Kumar H, Kim IS, More SV, Kim BW, Bahk YY, Choi DK. Gastrodin protects apoptotic dopaminergic neurons in a toxin-induced Parkinson's disease model. *Evid-Based Compl Altern Med.* 2013; 514095:13.
23. Du T, Li L, Song N, Xie J, Jiang H. Rosmarinic acid antagonized 1-methyl-4-phenylpyridinium (MPP+)-induced neurotoxicity in MES23.5 dopaminergic cells. *Int J Toxicol.* 2010; 29:625–633.
24. Ham A, Lee HJ, Hong SS, Lee D, Mar W. Moracenin D from Mori cortex radices protects SH-SY5Y cells against dopamine-induced cell death by regulating nurr1 and α -synuclein expression. *Phytother Res.* 2012; 26:620–624.
25. Kim KH, Song K, Yoon SH, Shehzad O, Kim YS, Son JH. Rescue of PINK1 protein null-specific mitochondrial complex IV deficits by ginsenoside reactivation of nitric oxide signaling. *J Biol Chem.* 2012; 287:44109–44120.
26. Elyasi L, Eftekhari-Vaghefi SH, Esmaili-Mahani S. Morphine protects SH-SY5Y human neuroblastoma cells against 6-hydroxydopamine-induced cell damage: involvement of anti-oxidant, calcium blocking, and anti-apoptotic properties. *Rejuvenation Res.* 2014; 17:255-263.
27. Li BY, Yuan YH, Hu JF, Zhao Q, Zhang DM, Chen NH. Protective effect of Bu-7, a flavonoid extracted from *Clausena lansium*, against rotenone injury in PC12 cells. *Acta Pharmacol Sin.* 2011; 32:1321-1326.

28. Tai KK and Truong DD. (-)-Epigallocatechin-3-gallate (EGCG), a green tea polyphenol, reduces dichlorodiphenyltrichloroethane (DDT)-induced cell death in dopaminergic SHSY-5Y cells. *Neurosci Lett*. 2010; 482:183-187.
29. Tamilselvam K, Braidy N, Manivasagam T, Essa MM, Prasad NR, Karthikeyan S, Thenmozhi AJ, Selvaraju S, Guillemin GJ. Neuroprotective effects of hesperidin, a plant flavanone, on rotenone-induced oxidative stress and apoptosis in a cellular model for Parkinson's disease. *Oxid Med Cell Longev*. 2013; 2013: 102741.
30. Filomeni G, Graziani I, De Zio D, Dini L, Centonze D, Rotilio G, Ciriolo MR. Neuroprotection of kaempferol by autophagy in models of rotenone-mediated acute toxicity: possible implications for Parkinson's disease. *Neurobiol Aging*. 2012; 33:767-785.
31. Lee Y, Park HR, Chun HJ, Lee J. Silibinin prevents dopaminergic neuronal loss in a mouse model of Parkinson's disease via mitochondrial stabilization. *J Neurosci Res*. 2015; 93:755-765.
32. Park G, Kim HG, Ju MS, Ha SK, Park Y, Kim SY, Oh MS. 6-Shogaol, an active compound of ginger, protects dopaminergic neurons in Parkinson's disease models via anti-neuroinflammation. *Acta Pharmacol Sin*. 2013; 34:1131-1139.
33. Leal LKAM, Júnior HVN, Cunha GMA, Moraes MO, Pessoa C, Oliveira RA, Silveira ER, Canuto KM, Viana GSB. Amburoside A, a glucoside from *Amburana cearensis*, protects mesencephalic cells against 6-hydroxydopamine-induced neurotoxicity. *Neurosci Lett*. 2005; 388:86-90.
34. Wang XJ and Xu JX. Salvianic acid A protects human neuroblastoma SH-SY5Y cells against MPP⁺-induced cytotoxicity. *Neurosci Res*. 2005; 51:129-138.
35. Chao J, Yu MS, Ho YS, Wang M, Chang RC. Dietary oxyresveratrol prevents parkinsonian mimetic 6-hydroxydopamine neurotoxicity. *Free Radic Biol Med*. 2008; 45:1019-1026.
36. Chen JH, Ou HP, Lin CY, Lin FJ, Wu CR, Chang SW, Tsai CW. Carnosic acid prevents 6-hydroxydopamine-induced cell death in SH-SY5Y cells via mediation of glutathione synthesis. *Chem Res Toxicol*. 2012; 25:1893-1901.
37. Mendiola-Precoma J, Berumen LC, Padilla K, Garcia-Alcocer G. Therapies for Prevention and Treatment of Alzheimer's Disease. *Biomed Res Int*. 2016; 2016:2589276.
38. Edwards Iii GA, Gamez N, Escobedo G Jr, Calderon O, Moreno-Gonzalez I. Modifiable Risk Factors for Alzheimer's Disease. *Front Aging Neurosci*. 2019; 11:146.
39. Raz L, Knoefel J, Bhaskar K. The neuropathology and cerebrovascular mechanisms of dementia. *J Cereb Blood Flow Metab*. 2016; 36:172-186.
40. Libro R, Giacoppo S, Soundara Rajan T, Bramanti P, Mazzone E. Natural Phytochemicals in the Treatment and Prevention of Dementia: An Overview. *Mol*. 2016; 21(4):518.
41. Serrano-Pozo A, Frosch MP, Masliah E, Hyman BT. Neuropathological alterations in Alzheimer disease. *Cold Spring Harb Perspect Med*. 2011; 1:a006189.
42. Teixeira JP, de Castro AA, Soares FV, da Cunha EFF, Ramalho TC. Future Therapeutic Perspectives into the Alzheimer's Disease Targeting the Oxidative Stress Hypothesis. *Mol*. 2019; 24:4410.
43. Colović MB, Krstić DZ, Lazarević-Pašti TD, Bondžić AM, Vasić VM. Acetylcholinesterase inhibitors: pharmacology and toxicology. *Curr Neuropharmacol*. 2013; 11:315-335.
44. Olivares D, Deshpande VK, Shi Y, Lahiri DK, Greig NH, Rogers JT, Huang X. N-methyl D-aspartate (NMDA) receptor antagonists and memantine treatment for Alzheimer's disease, vascular dementia and Parkinson's disease. *Curr Alzheimer Res*. 2012; 9:746-758.
45. Liu J, Chang L, Song Y, Li H, Wu, Y. The role of NMDA receptors in Alzheimer's disease. *Front Neurosci*. 2019; 13:1-22.
46. Kuang H, Zhou ZF, Zhu YG, Wan, ZK, Yang, MW, Hong, FF, Yang, SL. Pharmacological Treatment of Vascular Dementia: A Molecular Mechanism Perspective. *Aging Dis*. 2021; 12:308-326.
47. Lastra G, Syed S, Kurukulasuriya LR, Manrique C, Sowers JR. Type 2 diabetes mellitus and hypertension: an update. *Endocrinol Metab Clin North Am*. 2014; 43:103-122.
48. Baba M, Nakajo S, Tu PH, Tomita T, Nakaya K, Lee VM, Trojanowski JQ, Iwatsubo T. Aggregation of alpha-synuclein in Lewy bodies of sporadic Parkinson's disease and dementia with Lewy bodies. *Am J Pathol*. 1998; 152:879-884.
49. Orme T, Guerreiro R, Bras J. The Genetics of Dementia with Lewy Bodies: Current Understanding and Future Directions. *Curr Neurol Neurosci Rep*. 2018; 18:67.
50. Zhang G, Xia Y, Wan F, Ma, K, Guo X, Kou L, Yin S, Han C, Liu L, Huang J, Xiong N, Wang, T. New Perspectives on Roles of Alpha-Synuclein in Parkinson's Disease. *Front Aging Neurosci*. 2018; 10:370.
51. Jicha GA and Nelson PT. Management of frontotemporal dementia: targeting symptom management in such a heterogeneous disease requires a wide range of therapeutic options. *Neurodegener Dis Manag*. 2011; 1:141-156.
52. Libro R, Giacoppo S, Soundara Rajan T, Bramanti P, Mazzone E. Natural Phytochemicals in the Treatment and Prevention of Dementia: An Overview. *Mol*. 2016; 21:518.
53. Jin CY, Lee JD, Park C, Choi YH, Kim GY. Curcumin attenuates the release of pro-inflammatory cytokines in lipopolysaccharide-stimulated BV2 microglia. *Acta Pharmacol Sin*. 2007; 28:1645-1651.
54. Durairajan SSK, Liu LF, Lu JH, Chen LL, Yuan Q, Chung SK, Huang L, Li XS, Huang JD, Li M. Berberine ameliorates β -amyloid pathology, gliosis, and cognitive impairment in an Alzheimer's disease transgenic mouse model. *Neurobiol Aging*. 2012; 33:2903-2919.
55. Lim GP, Chu T, Yang F, Beech W, Frautschy SA, Cole GM. The Curry Spice Curcumin Reduces Oxidative Damage and Amyloid Pathology in an Alzheimer Transgenic Mouse. *J Neurosci*. 2001; 21:8370-8377.
56. Wang Z, Xiong L, Wan W, Duan L, Bai X, Zu H. Intranasal BMP9 Ameliorates Alzheimer Disease-Like Pathology and Cognitive Deficits in APP/PS1 Transgenic Mice. *Front Mol Neurosci*. 2017; 10:1-14.
57. Debrasad C, Hemanta M, Paromita B, Durbadal O, Kumar KA, Shanta D, Kumar HP, Tapan C, Ashoke S, Sekhar C. Inhibition of NO(2), PGE(2), TNF- α , and iNOS EXpression by *Shorea robusta* L.: An Ethnomedicine Used for Anti-Inflammatory and Analgesic Activity. *Evid-Based Compl Altern Med*. 2012; 2012:254849.
58. Jang JH and Surh YJ. Protective effect of resveratrol on beta-amyloid-induced oxidative PC12 cell death. *Free Radic Biol Med*. 2003; 34:1100-1110.
59. Mokni M, Elkahoui S, Limam F, Amri M, Aouani E. Effect of resveratrol on antioxidant enzyme activities in the brain of healthy rat. *Neurochem Res*. 2007; 32:981-987.
60. Porquet D, Griñán-Ferré C, Ferrer I, Camins A, Sanfeliu C, del Valle J, Pallàs M. Neuroprotective role of trans-resveratrol in a murine model of familial Alzheimer's disease. *J Alzheimers Dis*. 2014; 42:1209-1220.
61. Lonze BE and Ginty DD. Function and Regulation of CREB Family Transcription Factors in the Nervous System. *Neuron*. 2002; 35:605-623.
62. Kim SJ, Jeong HJ, Lee KM, Myung NY, An NH, Yang WM, Park SK, Lee HJ, Hong SH, Kim HM. Epigallocatechin-3-gallate suppresses NF-kappaB activation and phosphorylation of p38 MAPK and JNK in human astrocytoma U373MG cells. *J Nutr Biochem*. 2007; 18:587-596.

63. Biasibetti R, Tramontina AC, Costa AP, Dutra MF, Quincozes-Santos A, Nardin P, Bernardi CL, Wartchow KM, Lunardi PS, Gonçalves CA. Green tea (–) epigallocatechin-3-gallate reverses oxidative stress and reduces acetylcholinesterase activity in a streptozotocin-induced model of dementia. *Behav Brain Res.* 2013; 236:186–193.
64. Krobitch S and Kazantsev AG. Huntington's disease: From molecular basis to therapeutic advances. *Int J Biochem Cell Biol.* 2011; 43:20-24.
65. Kumar P, Kalonia H, Kumar A. Huntington's disease: Pathogenesis to animal models. *Pharmacol Rep.* 2010; 62:1-14.
66. Zadori D, Geisz A, Vamos E, Vecsei L, Klivenyi P. Valproate ameliorates the survival and the motor performance in a transgenic mouse model of Huntington's disease. *Pharmacol Biochem Behav.* 2009; 94:148-153.
67. Nayak A, Ansar R, Verma SK, Bonifati DM, Kishore U. Huntington's disease: An immune perspective. *Neurol Res Int.* 2011; 2011: 563784.
68. Walker FO. Huntington's disease. *Lancet.* 2007; 369:218-228.
69. Margolis RL and Ross CA. Diagnosis of Huntington disease. *Clin Chem.* 2003; 49:1726-1732.
70. Phillips W, Shannon KW, Barker RA. The current clinical management of Huntington's disease. *Mov Disord.* 2008; 23:1491-504.
71. Dey A and De JN. Anti-snake venom botanicals used by the ethnic groups of Purulia District, West Bengal, India. *J Herbs Spices Med Plants.* 2012; 18:152-165.
72. Dey A and De JN. Ethnomedicinal plants used by the tribals of Purulia district, West Bengal, India against gastrointestinal disorders. *J Ethnopharmacol.* 2012; 143:68-80.
73. Man SC, Durairajan SS, Kum WF, Lu JH, Huang JD, Cheng CF, Chung V, Xu M, Li M. Systematic review on the efficacy and safety of herbal medicines for Alzheimer's disease. *J Alzheimers Dis.* 2008; 14:209-223.
74. Pedraza-Chaverrí J, Reyes-Fermín LM, Nolasco-Amaya EG, Orozco-Ibarra M, Medina-Campos ON, González-Cuahutencos O, Rivero-Cruz I, Mata R. ROS scavenging capacity and neuroprotective effect of alpha-mangostin against 3-nitropropionic acid in cerebellar granule neurons. *Exp Toxicol Pathol.* 2009; 61:491-501.
75. Zhang J, Li CY, Xu MJ, Wu T, Chu JH, Liu SJ, Ju WZ. Oral bioavailability and gender-related pharmacokinetics of celastrol following administration of pure celastrol and its related tablets in rats. *J Ethnopharmacol.* 2012; 144:195-200.
76. Sagredo O, Ramos JA, Decio A, Mechoulam R, Fernández-Ruiz J. Cannabidiol reduced the striatal atrophy caused 3-nitropropionic acid in vivo by mechanisms independent of the activation of cannabinoid, vanilloid TRPV1 and adenosine A2A receptors. *Eur J Neurosci.* 2007; 26:843-851.
77. Zhang YQ and Sarge KD. Celastrol inhibits polyglutamine aggregation and toxicity though induction of the heat shock response. *J Mol Med (Berl).* 2007; 85:1421-1428.
78. Sandhir R, Yadav A, Mehrotra A, Sunkaria A, Singh A, Sharma S. Curcumin nanoparticles attenuate neurochemical and neurobehavioral deficits in experimental model of Huntington's disease. *Neuromolecular Med.* 2014; 16:106-118.
79. Ehrnhoefer DE, Duennwald M, Markovic P, Wacker JL, Engemann S, Roark M, Legleiter J, Marsh JL, Thompson LM, Lindquist S, Muchowski PJ, Wanker EE. Green tea (–) epigallocatechin-gallate modulates early events in huntingtin misfolding and reduces toxicity in Huntington's disease models. *Hum Mol Genet.* 2006; 15:2743-2751.
80. Maher P, Dargusch R, Bodai L, Gerard PE, Purcell JM, Marsh JL. ERK activation by the polyphenols fisetin and resveratrol provides neuroprotection in multiple models of Huntington's disease. *Hum Mol Genet.* 2011; 20:261-270.
81. Park JE, Lee ST, Im WS, Chu K, Kim M. Galantamine reduces striatal degeneration in 3-nitropropionic acid model of Huntington's disease. *Neurosci Lett.* 2008; 448:143-147.
82. Wu J, Jeong HK, Bulin SE, Kwon SW, Park JH, Bezprozvany I. Ginsenosides protect striatal neurons in a cellular model of Huntington's disease. *J Neurosci Res.* 2009; 87:1904-1912.
83. Menze ET, Tadros MG, Abdel-Tawab AM, Khalifa AE. Potential neuroprotective effects of hesperidin on 3-nitropropionic acid-induced neurotoxicity in rats. *Neurotoxicol.* 2012; 33:1265-1275.
84. Lagoa R, Lopez-Sanchez C, Samhan-Arias AK, Ganan CM, Garcia-Martinez V, Gutierrez-Merino C. Kaempferol protects against rat striatal degeneration induced by 3-nitropropionic acid. *J Neurochem.* 2009; 111:473-487.
85. Binawade Y and Jagtap A. Neuroprotective Effect of lutein against 3-nitropropionic acid-induced Huntington's disease-like symptoms: possible behavioral, biochemical, and cellular alterations. *J Med Food.* 2013; 16:934-943.
86. Kumar P, Kalonia H, Kumar A. Lycopene modulates nitric oxide pathways against 3-nitropropionic acid-induced neurotoxicity. *Life Sci.* 2009; 85:711-718.
87. Túnez I, Montilla P, Del Carmen Muñoz M, Feijóo M, Salcedo M. Protective effect of melatonin on 3-nitropropionic acid-induced oxidative stress in synaptosomes in an animal model of Huntington's disease. *J Pineal Res.* 2004; 37:252-256.
88. Kumar P and Kumar A. Protective effect of hesperidin and naringin against 3-nitropropionic acid induced Huntington's like symptoms in rats: possible role of nitric oxide. *Behav Brain Res.* 2010; 206:38-46.
89. Tariq M, Khan HA, Elfaki I, Al Deeb S, Al Moutaery K. Neuroprotective effect of nicotine against 3-nitropropionic acid (3-NP)-induced experimental Huntington's disease in rats. *Brain Res Bull.* 2005; 67:161-168.
90. Wu AG, Wong VK, Xu SW, Chan WK, Ng CI, Liu L, Law BY. Onjisaponin B derived from *Radix Polygalae* enhances autophagy and accelerates the degradation of mutant α -synuclein and huntingtin in PC-12 cells. *Int J Mol Sci.* 2013; 14:22618-22641.
91. Sandhir R and Mehrotra A. Quercetin supplementation is effective in improving mitochondrial dysfunctions induced by 3-nitropropionic acid: implications in Huntington's disease. *Biochim Biophys Acta.* 2013; 1832:421-430.
92. Pérez-De La Cruz V, González-Cortés C, Pedraza-Chaverrí J, Maldonado PD, Andrés-Martínez L, Santamaría A. Protective effect of S-allylcysteine on 3-nitropropionic acid-induced lipid peroxidation and mitochondrial dysfunction in rat brain synaptosomes. *Brain Res Bull.* 2006; 68:379-383.
93. Lam PY and Ko KM. Beneficial effect of (–)schisandrin B against 3-nitropropionic acid-induced cell death in PC12 cells. *Biofact.* 2012; 38:219-225.
94. Kumar P, Kalonia H, Kumar A. Huntington's disease: Pathogenesis to animal models. *Pharmacol Rep.* 2010; 62:1-14.
95. Sarkar S, Davies JE, Huang Z, Tunnacliffe A, Rubinsztein DC. Trehalose, a novel mTOR-independent autophagy enhancer, accelerates the clearance of mutant huntingtin and alpha-synuclein. *J Biol Chem.* 2007; 282:5641-5652.
96. Moya-Alvarado G, Gershoni-Emek N, Perlson E, Bronfman FC. Neurodegeneration and Alzheimer's disease (AD). What Can Proteomics Tell Us About the Alzheimer's Brain?. *Mol Cell Proteomics.* 2016; 15:409-425.
97. Berchtold NC and Cotman CW. Evolution in the Conceptualization of Dementia and Alzheimer's Disease: Greco-Roman Period to the 1960s. *Neurobiol Aging.* 1998; 19:173-189.
98. Venkatesan R, Ji E, Kim SY. Phytochemicals That Regulate Neurodegenerative Disease by Targeting Neurotrophins: A

- Comprehensive Review. Biomed Res Int. 2015;2015:814068.
99. Davinelli S, Sapere N, Zella, D, Bracale R, Intrieri M, Scapagnini G. Pleiotropic protective effects of phytochemicals in Alzheimer's disease. *Oxid Med Cell Longev*. 2012; 2012:386527.
 100. Li M, Guo K, Ikehara S. Stem Cell Treatment for Alzheimer's Disease. *Int J Mol Sci*. 2014; 15:19226-19238.
 101. De-Paula VJ, Radanovic M, Diniz BS, Forlenza OV. Alzheimer's disease. *Subcell Biochem*. 2012; 65:329-352.
 102. Förstl H and Kurz A. Clinical features of Alzheimer's disease. *Eur Arch Psychiatry Clin Neurosci*. 1999; 249:288-290.
 103. Taler V and Phillips NA. Language Performance in Alzheimer's Disease and Mild Cognitive Impairment: a comparative review. *J Clin Exp Neuropsychol*. 2008; 30:501-556.
 104. Shakir T, Coulibaly AY, Kehoe PG. An exploration of potential mechanisms and translational potential of five medicinal plants for applications in Alzheimer's disease. *Am J Neurodegener Dis*. 2013; 2:70-88.
 105. Khairallah MI and Kassem LA. Alzheimer's disease: current status of etiopathogenesis and therapeutic strategies. *Pak J Biol Sci*. 2011; 14:257-272.
 106. Fang L, Gou S, Fang X, Cheng L, Fleck C. Current progresses of novel natural products and their derivatives/analogs as anti-Alzheimer candidates: an update. *Mini Rev Med Chem*. 2013; 13:870-887.
 107. Kovács T. Therapy of Alzheimer disease. *Neuropsychopharmacol Hung*. 2009; 11:27-33.
 108. Kim J, Lee HJ, Lee KW. Naturally occurring phytochemicals for the prevention of Alzheimer's disease. *J Neurochem*. 2010; 112:1415-1430.
 109. Honjo H, Iwasa K, Fushiki S, Hosoda T, Tatsumi H, Mihara M, Hirasugi Y, Oida M, Kariya K, Kikuchi N, Kawata M. Estrogen and non-feminizing estrogen for Alzheimer's disease. *Endocr J*. 2003; 50:361-367.
 110. Ghavami A, Hirst WD, Novak TJ. Selective phosphodiesterase (PDE)-4 inhibitors: a novel approach to treating memory deficit? *Drugs R D*. 2006; 7:63-71.
 111. Engelborghs S, Gilles C, Ivanou A, Vandewoude M. Rationale and clinical data supporting nutritional intervention in Alzheimer's disease. *Acta Clin Belg*. 2014; 69:17-24.
 112. Kennedy DO and Wightman EL. Herbal extracts and phytochemicals: Plant secondary metabolites and the enhancement of human brain function. *Adv Nutr*. 2011; 2:32-50.
 113. Fang X, Lin X, Liang S, Zhang WD, Feng Y, Ruan KF. Phytochemical study of *Hicriopteris glauca*. *Chem Nat Compd*. 2013; 49:514-515.
 114. Elgorashi EE, Stafford GI, Van Staden J. Acetylcholinesterase enzyme inhibitory effects of amaryllidaceae alkaloids. *Planta Med*. 2004; 70:260-262.
 115. Heo HJ, Cho HY, Hong B, Kim HK, Kim EK, Kim BG, Shin DH. Protective effect of 4',5-dihydroxy-3',6,7-trimethoxyflavone from *Artemisia asiatica* against Abeta-induced oxidative stress in PC12 cells. *Amyloid*. 2001; 3:194-201.
 116. Park SH, Lee H, Kim HS, Kim YR, Noh SH. Optimum conditions for S- allyl-(L)-cysteine accumulation in aged garlic by RSM. *Food Sci Biotechnol*. 2014; 23:717-722.
 117. Orhan I, Terzioglu S, Sener B. Alpha-onocerin: an acetylcholinesterase inhibitor from *Lycopodium clavatum*. *Planta Med*. 2003; 69:265-267.
 118. Wang H, Xu Y, Yan J, Zhao X, Sun X, Zhang Y, Guo J, Zhu C. Acteoside protects human neuroblastoma SH-SY5Y cells against beta-amyloid-induced cell injury. *Brain Res*. 2009; 1283:139-147.
 119. Zhao L, Wang JL, Liu R, Li XX, Li JF, Zhang L. Neuroprotective, anti-amyloidogenic and neurotrophic effects of apigenin in an Alzheimer's disease mouse model. *Mol*. 2013; 18:9949-9965.
 120. Wei G, Chen, YB, Chen D F, Lai XP, Liu DH, Deng RD, Zhou JH, Zhang SX, Li YW, Lii H, Liu LF, Wang Q, Nie H. β -Asarone inhibits neuronal apoptosis via the CaMKII/CREB/Bcl-2 signaling pathway in an in vitro model and A β PP/PS1 mice. *J Alzheimers Dis*. 2013; 33:863-880.
 121. Heo HJ, Kim DO, Shin SC, Kim MJ, Kim BG, Shin DH. Effect of antioxidant flavanone, naringenin, from *Citrus junoson* neuroprotection. *J Agric Food Chem*. 2004; 52:1520-1525.
 122. Urbain A, Marston A, Queiroz EF, Ndjoko K, Hostettmann K. Xanthenes from *Gentiana campestris* as new acetylcholinesterase inhibitors. *Planta Med*. 2004; 70:1011-1014.
 123. Jiang B, Du J, Liu JH, Bao YM, An LJ. Catalpol attenuates the neurotoxicity induced by beta-amyloid (1-42) in cortical neuron-glia cultures. *Brain Res*. 2008; 1188:139-147.
 124. Mei Z, Yan P, Situ B, Mou Y, Liu P. Cryptotanshinone inhibits β -amyloid aggregation and protects damage from β -amyloid in SH-SY5Y cells. *Neurochem Res*. 2012; 37:622-628.
 125. Durairajan SS, Liu LF, Lu JH, Koo I, Maruyama, K, Chung SK, Huang JD, Li M. Stimulation of non-amyloidogenic processing of amyloid- β protein precursor by cryptotanshinone involves activation and translocation of ADAM10 and PKC- α . *J Alzheimers Dis*. 2011; 25:245-262.
 126. Park SY and Kim DS. Discovery of natural products from *Curcuma longa* that protect cells from beta-amyloid insult: a drug discovery effort against Alzheimer's disease. *J Nat Prod*. 2002; 65:1227-1231.
 127. Wang J, Cao F, Su E, Wu C, Zhao L, Ying R. Improving flavonoid extraction from *Ginkgo biloba* leaves by prefermentation processing. *J Agric Food Chem*. 2013; 61:5783-5791.
 128. Ahmed T and Gilani AH. Inhibitory effect of curcuminoids on acetylcholinesterase activity and attenuation of scopolamine-induced amnesia may explain medicinal use of turmeric in Alzheimer's disease. *Pharmacol Biochem Behav*. 2009; 91:554-559.
 129. Ahmed T and Gilani AH. A comparative study of curcuminoids to measure their effect on inflammatory and apoptotic gene expression in an A β plus ibotenic acid-infused rat model of Alzheimer's disease. *Brain Res*. 2011; 1400:1-18.
 130. Lee KY, Yoon JS, Kim ES, Kang SY, Kim YC. Anti-acetylcholinesterase and anti-amnesic activities of a pregnane glycoside, cynatroside B, from *Cynanchum atratum*. *Planta Med*. 2005; 71:7-11.
 131. Yan JJ, Kim DH, Moon YS, Jung JS, Ahn EM, Baek NI, Song DK. Protection against beta-amyloid peptide-induced memory impairment with long-term administration of extract of *Angelica gigas* or decursinol in mice. *Prog Neuropsychopharmacol Biol Psychiatry*. 2004; 28:25-30.
 132. Li L, Zhang X, Cui L, Wang L, Liu H, Ji H, Du Y. Ursolic acid promotes the neuroprotection by activating Nrf2 pathway after cerebral ischemia in mice. *Brain Res*. 2013; 1497:32-39.
 133. Aronson S, Van Baelen B, Kavanagh S, Schwalen S. Optimal dosing of galantamine in patients with mild or moderate Alzheimer's disease: post Hoc analysis of a randomized, double-blind, placebo-controlled trial. *Drugs Aging*. 2009; 26:231-239.
 134. Suh GH, Jung HY, Lee CU, Oh BH, Bae JN, Jung HY, Ju YS, Yeon BK, Park J, Hong I, Choi S, Lee JH. A prospective, double-blind, community-controlled comparison of three doses of galantamine in the treatment of mild to moderate Alzheimer's disease in a Korean population. *Clin Ther*. 2004; 26:1608-1618.

135. Zeng H, Chen Q, Zhao B. Genistein ameliorates beta-amyloid peptide (25-35)-induced hippocampal neuronal apoptosis. *Free Radic Biol Med.* 2004; 36:180-188.
136. Bate C, Salmons M, Williams A. Ginkgolide B inhibits the neurotoxicity of prions or amyloid- β 1-42. *J Neuroinflamm.* 2004;1:4.
137. Wang J, Li PT, Du H, Hou JC, Li WH, Pan YS, Chen HC. Tong Luo Jiu Nao injection, a traditional Chinese medicinal preparation, inhibits MIP-1 β expression in brain microvascular endothelial cells injured by oxygen-glucose deprivation. *J Ethnopharmacol.* 2012; 141:151-157.
138. Xiao XQ, Zhang HY, Tang XC. Huperzine A attenuates amyloid beta-peptide fragment 25-35-induced apoptosis in rat cortical neurons via inhibiting reactive oxygen species formation and caspase-3 activation. *J Neurosci Res.* 2002; 67:30-36.
139. Zeng KW, Ko H, Yang HO, Wang XM. Icaritin attenuates β -amyloid-induced neurotoxicity by inhibition of tau protein hyperphosphorylation in PC12 cells. *Neuropharmacol.* 2010; 59:542-550.
140. Xian YF, Mao QQ, Wu, JC, Su ZR, Chen JN, Lai XP, Ip SP, Lin ZX. Isorhynchophylline treatment improves the amyloid- β -induced cognitive impairment in rats via inhibition of neuronal apoptosis and tau protein hyperphosphorylation. *J Alzheimers Dis.* 2014; 39:331-346.
141. Kim TI, Lee YK, Park SG, Choi IS, Ban JO, Park HK, Nam SY, Yun YW, Han SB, Oh KW, Hong J. T l-Theanine, an amino acid in green tea, attenuates beta-amyloid-induced cognitive dysfunction and neurotoxicity: reduction in oxidative damage and inactivation of ERK/p38 kinase and NF-kappaB pathways. *Free Radic Biol Med.* 2009; 47:1601-1610.
142. Zheng N, Yuan P, Li C, Wu J, Huang J. Luteolin reduces BACE1 expression through NF- κ B and through estrogen receptor mediated pathways in HEK293 and SH-SY5Y cells. *J Alzheimers Dis.* 2015; 45:659-671.
143. Heo HJ, Kim DO, Shin SC, Kim MJ, Kim BG, Shin DH. Effect of antioxidant flavanone, naringenin, from Citrus junoson neuroprotection. *J Agric Food Chem.* 2004; 52:1520-1525.
144. Ma B, Meng X, Wang J, Sun J, Ren X, Qin M, Sun J, Sun G, Sun X. Notoginsenoside R1 attenuates amyloid- β -induced damage in neurons by inhibiting reactive oxygen species and modulating MAPK activation. *Int Immunopharmacol.* 2014; 22:151-159.
145. Kim SJ, Jeong HJ, Lee KM, Myung NY, An NH, Yang WM, Park SK, Lee HJ, Hong SH, Kim HM, Um JY. Epigallocatechin-3-gallate suppresses NF-kappaB activation and phosphorylation of p38 MAPK and JNK in human astrocytoma U373MG cells. *J Nutr Biochem.* 2007; 18:587-596.
146. Yoon JH, Youn K, Ho CT, Karwe MV, Jeong WS, Jun M. p-Coumaric acid and ursolic acid from Corni fructus attenuated β -amyloid(25-35)-induced toxicity through regulation of the NF- κ B signaling pathway in PC12 cells. *J Agric Food Chem.* 2014; 62:4911-4916.
147. Chonpathompikunlert P, Wattanathorn J, Muchimapura S. Piperine, the main alkaloid of Thai black pepper, protects against neurodegeneration and cognitive impairment in animal model of cognitive deficit like condition of Alzheimer's disease. *Food Chem Toxicol.* 2010; 48:798-802.
148. Li F, Gao B, Dong H, Shi J, Fang D. Icaritin induces synoviolin expression through NFE2L1 to protect neurons from ER stress-induced apoptosis. *PLoS One.* 2015; 10:0119955.
149. Zhang L, Yu H, Zhao X, Lin X, Tan C, Cao G, Wang Z. Neuroprotective effects of salidroside against betaamyloid-induced oxidative stress in SH-SY5Y human neuroblastoma cells. *Neurochem Int.* 2010; 57:547-555.
150. Peng Q, Buz'Zard AR, Lau BH. Neuroprotective effect of garlic compounds in amyloid-beta peptideinduced apoptosis in vitro. *Med Sci Monit.* 2002; 8:328-337.
151. Chauhan NB. Effect of aged garlic extract on APP processing and tau phosphorylation in Alzheimer's transgenic model Tg2576. *J Ethnopharmacol.* 2006; 108:385-394.
152. Lu P, Mamiya T, Lu LL, Mouri A, Niwa M, Hiramatsu M, Zou LB, Nagai T, Ikejima T, Nabeshima TI. Silibinin attenuates amyloid beta (25-35) peptide-induced memory impairments: implication of inducible nitric-oxide synthase and tumor necrosis factor-alpha in mice. *J Pharmacol Exp Ther.* 2009; 331:319-326.
153. Ingkaninan K, Changwijit K, Suwanborirux K. Vobasinyliboga bisindole alkaloids, potent acetylcholinesterase inhibitors from *Tabernaemontana divaricata* root. *J Pharm Pharmacol.* 2006; 58:847-852.
154. Urbain A, Marston A, Grilo LS, Bravo J, Purev O, Puresuren B, Batsuren D, Reist M, Carrupt PA, Hostettmann K. Xanthones from *Gentianaella amarella* ssp. *acuta* with acetylcholinesterase and monoamine oxidase inhibitory activities. *It Good.* 2008; 71:895-897.
155. Pan XD, Chen XC, Zhu YG, Chen LM, Zhang J, Huang TW, Ye QY, Huang HP. Tripchlorolide protects neuronal cells from microglia-mediated beta-amyloid neurotoxicity through inhibiting NF-kappaB and JNK signaling. *Glia.* 2009; 57:1227-1238.
156. Hong SY, Jeong WS, Jun M. Protective effects of the key compounds isolated from Corni fructus against β -amyloid-induced neurotoxicity in PC12 cells. *Mol.* 2012; 17:10831-10845.
157. Chi TY, Wang LH, Ji XF, Shen L, Zou LB. Protective effect of xanthoceraside against β -amyloid-induced neurotoxicity in neuroblastoma SH-SY5Y cells. *J Asian Nat Prod Res.* 2013; 15:1013-1022.
158. Yu Y, Zhou L, Sun M, Zhou T, Zhong K, Wang H, Liu Y, Liu X, Xiao R, Ge J, Tu P, Fan DS, Lan Y, Hui C, Chui D. Xylocoside G reduces amyloid- β induced neurotoxicity by inhibiting NF- κ B signaling pathway in neuronal cells. *J Alzheimers Dis.* 2012; 30:263-275.
159. Heo HJ, Cho HY, Hong B, Kim HK, Heo TR, Kim EK, Kim SK, Kim CJ, Shin DH. Ursolic acid of *Origanum majorana* L. reduces Abeta-induced oxidative injury. *Mol Cells.* 2002; 13:5-11.
160. O'Keane V, Frodl T, Dinan TG. A review of atypical depression in relation to the course of depression and changes in HPA axis organization. *Psychoneuroendocrinology.* 2012; 37:1589-1599.
161. Berton O and Nestler EJ. New approaches to antidepressant drug discovery: beyond monoamines. *Nat Rev Neurosci.* 2006; 7:137-151.
162. Mitchelmore C and Gede L. Brain derived neurotrophic factor: epigenetic regulation in psychiatric disorders. *Brain Res.* 2014; 1586:162-172.
163. Evelyn B, Laura HA, Irving H, Nancy AS, Jordi A, Giovanni DG, Ron DG, Koen D, Chiya H, Noboru I, Aimee NK, Jagdish K, Stanislav K, Jean-Pierre L, Daphna L, Herbert M, Maria EMM, Mark OB, Jose P, Maria CV, David RW, Ronald CK. Cross-national epidemiology of DSM-IV major depressive episode. *BMC Med.* 2011; 9:90.
164. Shulman KI, Herrmann N, Walker SE. Current place of monoamine oxidase inhibitors in the treatment of depression. *CNS Drugs.* 2013; 27:789-797.
165. Gillman PK. Tricyclic antidepressant pharmacology and therapeutic drug interactions updated. *Br J Pharmacol.* 2007; 151:737-748.
166. Nemeroff CB. Prevalence and management of treatment resistant depression. *J Clin Psychiatry.* 2007; 68:17-25.
167. Trivedi MH, Fava M, Wisniewski SR, Thase ME, Quitkin F, Warden D, Ritz L, Nierenberg AA, Lebowitz BD, Biggs MM, Luther JF, Shores-Wilson K, Rush AJ, STAR*D

- Study Team . Medication augmentation after the failure of SSRIs for depression. *The New Eng J Med.* 2006; 354:1243-1252.
168. Baker CB, Johnsrud MT, Crismon ML, Rosenheck RA, Woods SW. Quantitative analysis of sponsorship bias in economic studies of antidepressants. *Br J Psychiatry.* 2003; 183:498-506.
169. Meijer WEE, Heerdink ER, Nolen WA, Herings RMC, Leufens HGM, Egberts ACG. Association of risk of abnormal bleeding with degree of serotonin reuptake inhibition by antidepressants. *Arch Int Med.* 2004; 164:2367-2370.
170. Vanderkooy JD, Kennedy SH, Aladeokin AC. Antidepressant side effects in depression patients treated in a naturalistic setting: a study of bupropion, moclobemide, paroxetine, sertraline, and venlafaxine. *Can J Psychiatry.* 2002; 47:174-180.
171. Khawam EA, Laurencic G, Malone Jr DA. Side effects of antidepressants: an overview. *Cleveland Clin J Med.* 2006; 73:351-361.
172. Papakostas GI. Tolerability of modern antidepressants. *J Clin Psychiatry.* 2008; 69:8-13.
173. Linde K. St. John's wort—an overview. *Forschende Komplementarmedizin.* 2009; 16:146-155.
174. Melo FH, Moura BA, de Sousa DP, de Vasconcelos SM, Macedo DS, Fonteles MM, Viana GS, de Sousa FC. Antidepressant-like effect of carvacrol (5-Isopropyl- 2-methylphenol) in mice: involvement of dopaminergic system. *Fund Clin Pharmacol.* 2011; 25:362-367.
175. Zotti M, Colaianna M, Morgese MG, Tucci P, Schiavone S, Avato P, Trabace L. Carvacrol: from ancient flavoring to neuromodulatory agent. *Mol.* 2013; 18:6161-6172.
176. Xu Y, Ku BS, Yao HY, Ma X, Zhang YH, Li XJ. Antidepressant effect of curcumin in mice. *Chin J Clin Rehab.* 2005; 9:162-164.
177. Bhutani MK, Bishnoi M, Kulkarni SK. Anti-depressant like effect of curcumin and its combination with piperine in unpredictable chronic stress-induced behavioral, biochemical and neurochemical changes. *Pharmacol Biochem Behav.* 2009; 92:39-43.
178. Wang R, Xu Y, Wu HL, Li YB, Li YH, Guo JB, Li XJ. The antidepressant effects of curcumin in the forced swimming test involve 5-HT1 and 5-HT2 receptors. *Eur J Pharmacol.* 2008; 578:43-50.
179. Xu Y, Ku B, Cui L, Li X, Barish PA, Foster TC, Ogle WO. Curcumin reverses impaired hippocampal neurogenesis and increases serotonin receptor 1A mRNA and brain-derived neurotrophic factor expression in chronically stressed rats. *Brain Res.* 2007; 116:9-18.
180. Yabe T, Hirahara H, Harada N, Ito N, Nagai T, Sanagi T, Yamada H. Ferulic acid induces neural progenitor cell proliferation *in vitro* and *in vivo*. *Neurosci.* 2010; 165:515-524.
181. Yin C, Gou LLY, Liu Y, Yin X, Zhang L, Jia G, Zhuang X. Antidepressant-like effects of L-theanine in the forced swim and tail suspension tests in mice. *Phytother Res.* 2011; 25:1636-1639.
182. Xu C, Teng J, Chen W, Ge Q, Yang Z, Yu C, Yang Z, Jia W. 20(S)-protopanaxadiol, an active ginseng metabolite, exhibits strong antidepressant-like effects in animal tests. *Prog. Neuropsychopharmacol Biol Psychiatry.* 2010; 34:1402-1411.
183. Bhutata P, Mundhada Y, Bansod K, Ubgade A, Quazi M, Umathe S, Mundhada D. Reversal by quercetin of corticotrophin releasing factor induced anxiety- and depression-like effect in mice. *Progr Neuro-Psychopharmacol Biol Psychiatry.* 2010; 34:955-960.
184. Yu Y, Wang R, Chen C, Du X, Ruan L, Sun J, Li J, Zhang L, O'Donnell JM, Pan J, Xu Y. Antidepressant-like effect of trans-resveratrol in chronic stress model: behavioral and neurochemical evidences. *J Psychiatr Res.* 2013; 47:315-322.
185. Limanaqi F, Biagioni F, Mastroiacovo F, Polzella M, Lazzeri G, Fornai F. Merging the Multi-Target Effects of Phytochemicals in Neurodegeneration: From Oxidative Stress to Protein Aggregation and Inflammation. *Antioxid.* 2020; 9(10):1022.