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**Original Research Article** 



## Antioxidant Activity of Uncaria Gambir (Hunter) Roxb Extracts

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## ARTICLE INFO

ABSTRACT

Article history: Received 27 May 2022 Revised 27 July 2022 Accepted 29 August 2022 Published online 02 September *Uncaria Gambir* is a popular plant in Southeast Asia, used traditionally for healing various diseases. Pharmacologically, *U. Gambir* has many potentials. This study aimed to investigate the antioxidant activities of *U. Gambir* stem extracts (methanol, ethyl acetate and n-hexane) using DPPH (2,2-diphenyl-2-2-picrylhydrazyl) and ABTS (2,2'-azinobis (3-ethylbenzothiazole-6-sulfonic acid) methods. The results of this study show that methanol extract has the highest antioxidant activity in the DPPH assay with IC<sub>50</sub> value of 9.71 µg/mL. In addition, it has the highest ABTS activity with the IC<sub>50</sub> value of 6.63 µg/mL. This study showed that the methanol extract of *U. Gambir* can potentially be used as an antioxidant.

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Keywords: Antioxidant, ABTS, DPPH, Uncaria gambir (Hunter) Roxb, Stem Extracts.

## Introduction

Free radical formation occurs continuously as a consequence of oxidation reaction of stable compounds in the body's metabolism. Free radicals are unstable and highly reactive due to the unpaired electron in their atomic orbital.<sup>1</sup> In addition, it can either donate an electron or accept an electron from other molecules. Furthermore, free radicals such as hydroxyl radical, superoxide anion radical, hydrogen peroxide, oxygen singlet, hypochlorite, nitric oxide radical and peroxinitrite radical cause many diseases.<sup>2</sup> They can attack important macromolecules such as DNA, protein, carbohydrate, and lipids. This condition leads to the damage of macromolecules, and this can induces human diseases like Alzheimer's disease and cancer.<sup>3</sup> Hence, chemical substances are needed for the prevention of free radicals.

One of the chemical substances that play a role in inhibiting free radicals is antioxidant. This compound works by neutralizing free radicals through electron donors. The neutralization process forms more stable compound, thereby inhibiting the mechanism of oxidative reaction.<sup>4</sup> Antioxidant compounds can be obtained from natural or synthetic sources. Butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA), tert-butylhydroquinone (TBHQ), propyl gallate (PG) and octyl gallate (OG) are popular synthetic antioxidant compounds which are commonly used as antioxidants in food, cosmetic and industrial products.<sup>5,6</sup> However, synthetic antioxidants pose high risks on human body since they are potentially carcinogenic and chemically unstable. This justifies the need for natural antioxidant sources for humans. The sole alternative to synthetic antioxidant is natural antioxidant.<sup>4,7</sup> One of the natural antioxidant sources is Uncaria gambir (Hunter) Roxb, which is ubiquitous in Southeast Asia. Uncaria gambir (Hunter) Roxb, which is locally known as Gambir, is a climbing shrub native to Southest Asia, particularly in Sumatra and Borneo. This plant is traditionally used for treating various illnesses like diarrhea, dysentery, and sore throat.<sup>8</sup> U. gambir also can be used as mouthwash<sup>9</sup> and wound dressing.<sup>1</sup>

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Furthermore, this plant is utilized as an astringent and antiseptic as well.<sup>11</sup> This plant has been known to possess pharmacological properties: it can be used as anti-inflammation,<sup>12</sup> antiseptic,<sup>13</sup> anticancer,<sup>14</sup> and anticholesterol.<sup>15</sup> Previous researches reported that phenolics in *U*. gambir extract such as catechin, quercetin, quinic acid have contributed to antioxidant activities.<sup>16</sup> Gambirin A1, A2, B1, B2 have also been isolated from methanol extract of *U.gambir*.<sup>11</sup>

Free radical scavenging activity of U gambir using DPPH method has been reported. Ethyl acetate extract of U. gambir pollen has DPPH radical scavenging activity with IC<sub>50</sub> value of 25.55 µg/mL.<sup>17</sup> In addition, methanol extract of U. gambir leaves and shoots has DPPH radical scavenging activity with IC<sub>50</sub> value of 18.27 µg/mL.<sup>18</sup> Aqueous extract of U gambir leaves and stems has DPPH radical scavenging activity which ranged from 92 to 93.1%.<sup>19</sup> Recently, numerous researchers reported about the antioxidant which was derived from leaves, roots and shoots of this plant. However, reports about stem extract of U. gambir are limited in number. Additionally, there are many reports on free radical scavenging activity of U. gambir using DPPH method, whereas reports on free radical scavenging acvitity using ABTS methods is nonexistent so far.

The study aims mainly to find out the antioxidant activity of three solvents extracts of *U. gambir* stem: ethyl acetate, hexane and methanol extracts.

## **Materials and Methods**

## Plant material

*Uncaria gambir* stems were obtained from West Kalimantan, Indonesia, in May 2021. They were identified at Balai Materia Medika in Batu, Malang and were authenticated with voucher specimen number 074/495/102.20-A/2021.

## Materials and instrumentation

High grade organic solvents including methanol, ethanol, ethyl acetate, n-hexane and dimethylsulfoxide (DMSO) were purchased from Merck. DPPH (2,2-diphenyl-1-picrylhydrazyl), ABTS (2,2'-azinobis (3-ethyl-benzothiazoline-6-sulfonic acid), and  $K_2S_2O_8$  (potassium peroxydisulfate) were purchased from the same source. Trolox (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid; Sigma Aldrich) was used as antioxidant standard. Incubator EYELA SLI-400 was used to incubate the samples. The reaction was monitored by spectrophotometer UV (HITACHI U-2900, Japan). Rotary Evaporator (IKA RV 8 Basic, Germany) was used to obtain crude extracts.

#### Preparation of extracts

Dried gambir stem (25 g each) was extracted separately with methanol, ethyl acetate, and hexane (250 mL for 24 h) at room temperature. The extracts were filtered using filter paper then concentrated with a rotary evaporator under reduced pressure. U. *gambir* crude extracts were obtained from this treatment.

#### DPPH assay

DPPH activity of the crude extracts were assayed using a method described by Brand Williams,<sup>20</sup> modified by Dudonne *et al.*<sup>21</sup> Each of the crude extracts (10 mg) was dissolved in 1 mL methanol. The mixture consisted of 1 mL DPPH solution 6 x 10<sup>-5</sup> M and 33  $\mu$ L of methanol solution of crude extract. After 20 minutes of incubation at 37°C, the absorbance of the reaction mixture was measured at 515 nm by UV-VIS spectrophotometer to get the Absorbance value (As). Methanol was used as a blank and measured at same wavelength (Ab). The experiment was carried out in triplicate and DPPH activity was calculated using the following formula: <sup>21</sup>

Antioxidant activity (%) = 
$$\frac{A_b - A_s}{A_b} \times 100 \%$$
 (1)

Note: Ab = absorbance of solution without sample As = absorbance of sample solution

Furthermore, the extract which have a good DPPH inhibition (%) was measured for 50% inhibitory concentration (IC<sub>50</sub>) value ( $\mu$ g/mL). The value of IC<sub>50</sub> was represented as the concentration of the extracts which is needed to reduce the intensity of 50% DPPH concentration.<sup>22</sup>

## ABTS assay

ABTS activity of crude extracts were assayed using the method described by Pellegrini *et al.*<sup>23</sup> ABTS was dissolved in water at 7 mM concentration. Then, the ABTS radical cation was generated by reacting 5 mL of 7 mM ABTS with 88 µL of 140 mM potassium peroxydisulfate (K<sub>2</sub>S<sub>2</sub>O<sub>8</sub>). The mixture was allowed to stand at room temperature for 12-16 h to yield a dark blue solution. The ABTS solution was diluted with 99.5% ethanol to give an absorbance of 0.7  $\pm$  0.02 at 734 nm, before starting the assay. 1 mL DMSO was utilized to dissolve each of crude extracts (10 mg). The reaction mixture consisted of 1 mL ABTS solution and 10 µL of crude extract were shaken for 10 sec. After 4 min incubation at 30°C, the absorbance of the reaction mixture was measured at 734 nm by spectrophotometer UV-Vis to provide the  $A_s$  value. Blank sample with 10  $\mu$ L of DMSO in ABTS solution was prepared and measured at same wavelength to find out the Ab value. Trolox was used as a positive control. The ABTS activity was calculated by formula 1 with triplicate experiment. Further, the extract which has a good ABTS activity (%) was measured for its IC<sub>50</sub> value ( $\mu g/mL$ ).

#### Statistical analysis

The antioxidant activity analyses were performed on three varieties of stem extracts: methanol, ethyl acetate, and n-hexane solvent using independent t-test at an error rate of 5%. The tests were conducted to compare the above extracts' antioxidant activity against that of Trolox's as the positive control.

#### **Results and Discussion**

The extraction of *U. gambir* was performed by using methanol, ethyl acetate, and n-hexane which provided varying extraction yield; 1.59, 0.92, and 0.92 g, respectively. The highest yield extraction for various stem extracts was obtained with methanol solvent with the value of 0.0636 % w/w. The result showed different yield extraction for each solvent, depending on the sample's polarity. Then, the stem extract showed the tendency to be polar when extracted with methanol that has the highest polarity. This indicated that the main compounds of *U.gambir* stems are polar solvent soluble. The determination of antioxidant activity of *U. gambir* extracts was carried out using DPPH and ABTS methods. The antioxidant activity of *U. gambir* extracts which is based on DPPH assay at concentration of 39.93  $\mu$ g/mL can

be seen in Figure 1 which presents the percentages of antioxidant activity of stem extracts; methanol, ethyl acetate, and n-hexane were 90.31  $\pm$  0.93%, 86.23  $\pm$  0.146% and 27.5  $\pm$  0.31%, respectively. Trolox as a positive control has antioxidants activity of 95.11  $\pm$  0.086%. Figure 1 shows that the methanol extract of the stem of *U. gambir* has the highest activity in comparison with the other extracts, because these extracts may contain phenolic compounds that contributed to the antioxidant activity.<sup>16</sup> The previous study by Phongpaichit *et al*,<sup>24</sup> showed the strength of antioxidant activity based on DPPH assay: IC<sub>50</sub> > 250 µg/mL, inactive; >100-250 µg/mL, weakly active; <50-100 µg/mL, wery strongly active. Figure 2 shows that, the IC<sub>50</sub> value of *U. gambir* methanol extract is 9.71 µg/mL, while Trolox as positive control has IC<sub>50</sub> value of 4.12 µg/mL. This result indicates that *U. gambir* methanol extract is a strongly active antioxidant (IC<sub>50</sub> < 10 µg/mL).



**Figure 1:** DPPH Scavenging activity of *U. gambir* extracts at a concentration of 39.93  $\mu$ g/mL, MeOH, methanol extract; EA, ethyl acetate extract; HX, hexane extract and T, Trolox (positive control). Each column represents the mean±SD, n = 3



**Figure 2:** DPPH Scavenging activity of *U. gambir* methanol extract

Antioxidant activity of *U. gambir* extracts was also investigated using the ABTS method. The antioxidant activity of three extracts (methanol, ethyl acetate, and n-hexane) at a concentration of 49.5  $\mu$ g/mL are presented in Figure 3. The antioxidant activity percentage of *U. gambir* stem extracts were 94.88  $\pm$  0.405, 75.47  $\pm$  0.234, and 44.42  $\pm$  0.219%, respectively, while Trolox as a standard has

antioxidant activity of 98.04  $\pm$  0.673%. Figure 3 illustrates the ABTS scavenging activity of U.gambir stem extract on several solvents: ethyl acetate, hexane and methanol with Trolox. ABTS scavenging activity of methanol extract of U. gambir is shown in Figure 4. The IC<sub>50</sub> value of methanol extracts was 6.63 µg/mL, while, Trolox which functions as positive control has IC\_{50} value of 2.25  $\mu g/mL.$  The IC\_{50} value obtained from the methanol extract of U. gambir, the antioxidant activity of U. gambir is very strong. The result shows the correlation between IC<sub>50</sub> value and antioxidant activity: the lower the IC<sub>50</sub> value, the higher the antioxidant activity of the compound. The IC50 value of Trolox indicated that it had a higher activity than the methanol extract (the same as scavenging activity with DPPH method). Such result was made possible by the fact that Trolox is a derivative of vitamin E, which consist of aromatic ring with substitution of hydroxy and carboxylate group. The presence of distinct substituents in the backbone structure of phenol compound gives its antioxidant property. Further, it is also based on its structure, especially on several properties: the position, number of hydroxyl groups and the characteristic of the aromatic ring substituents.



**Figure 3:** ABTS Scavenging activity of *U. gambir* extracts at a concentration of 49.5  $\mu$ g/mL, MeOH, methanol extract; EA, ethyl acetate extract; HX, hexane extract and T, Trolox (positive control). Each column represents the mean±SD, n = 3



**Figure 4**: ABTS Scavenging activity of *U. gambir* methanol extract

 Table 1: DPPH and ABTS radical scavenging activity of U.

 gambir extracts

Extraction solvent	DPPH	ABTS
	$IC_{50} \left(\mu g/mL\right)$	$IC_{50}\left(\mu g/mL ight)$
Hexane	$107.11\pm0.58$	$73.07\pm0.54$
Ethyl acetate	$23.39\pm0.32$	$22.45\pm0.18$
Methanol	$9.71\pm0.45$	$6.63\pm0.36$
Trolox	$4.12\pm1.13$	$2.25\pm0.77$

Value are the Mean  $\pm$  SD for three parallel assessment (n=3)

Table 1 shows various extracts of U. gambir that were tested for antioxidant activity using DPPH and ABTS methods. The table also indicates that methanol extract of U. gambir has the highest antioxidant activity on both methods. The extract has an IC50 value of 9.71  $\mu$ g/mL, whereas Trolox has IC<sub>50</sub> value of 4.12  $\mu$ g/mL using DPPH method. Furthermore, the methanol extract has IC<sub>50</sub> value of 6.63 µg/mL, while Trolox has IC<sub>50</sub> value of 2.25 µg/mL using ABTS method. There was no meaningful difference of the results based on ttest between the antioxidant activity of the U. gambir extracts and Trolox as a positive control since the t-tailed value was > 0.05. Furthermore, the result shows that the scavenging activity with ABTS method provides higher antioxidant activity than DPPH method. Such result was made possible by the fact that the ABTS method test was performed with the absorbance reading of the ABTS++ measured at 734 nm. The wavelength was chosen to give the minimal interference from the other absorbing components and turbidity of sample. Another reason for this condition is the presence of phenolic which is contained in the extract of U. gambir. The phenolic interferences resulted in the near visible region rather than the far ones. DPPH posed serious problem of interference since it was analyzed at 515 nm, while ABTS scavenging method give less serious problem since the chromogen provides the absorbance peaks at 734 nm.<sup>2</sup>

## Conclusion

The antioxidant activity of *U.gambir* stems extract were confirmed by DPPH and ABTS methods on several solvent namely, methanol, ethyl acetate, and n-hexane. The methanol extract showed the highest antioxidant activity on both methods, DPPH with  $IC_{50}$  of 9.71 µg/mL and ABTS with  $IC_{50}$  of 6.63 µg/mL. Based on this result, methanol extract of *U. gambir* could be utilized as a potential source of antioxidant.

## **Conflict of Interest**

The authors declare no conflict of interest.

#### **Authors' Declaration**

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

#### References

- Ngo TC, Mai TVT, Pham TT, Jeremic S, Markovic Z, Huynh LK, Dao DQ. Natural acridones and coumarins as free radical scavengers: Mechanistic and kinetic studies. Chem Phys Lett. 2020; 746(9):137312.
- Munné-Bosch S and Pintó-Marijuan M. Free Radicals, Oxidative Stress and Antioxidants. Encycl Appl Plant Sci. 2016; 1:16–19.
- Poprac P, Jomova K, Simunkova M, Kollar V, Rhodes CJ, Valko M. Targeting Free Radicals in Oxidative Stress-Related Human Diseases. Trends Pharmacol Sci. 2017; 38(7):592–607.

- Pisoschi AM, Pop A, Iordache F, Stanca L, Predoi G, Serban AI. Oxidative stress mitigation by antioxidants - An overview on their chemistry and influences on health status. Eur J Med Chem. 2021; 209(1):112891.
- Igwe CU, Onyeagoro EE, Morah AC, Nwaogu LA, Iheme CI, Onwuliri VA. Phenolic Profile and In Vitro Antioxidant Potential of Four Medicinal Plants. Trop J Nat Prod Res. 2021; 5(11):20372042.
- Martelli G and Giacomini D. Antibacterial and antioxidant activities for natural and synthetic dual-active compounds. Eur J Med Chem. 2018; 158(16):91–105.
- Gulcin İ. Antioxidants and antioxidant methods: an updated overview. Arch Toxicol. 2020; 94(3):651-715.
- Lemmens J, Wulijarni N, Wageningen P. Plant Resources of South-East Asia. 1991; (3):125-127.
- Dian H, Dharsono A, Wibisono L, Hayati AT, Apriyanti E, Satari MH, Kurnia D. Mode action prediction of catechin from Uncaria gambir Roxb against UDP-Nacetylenolpyruvyl-glucosamine reductase (MurB enzyme) of Streptococcus mutans : In silico study. J Adv Pharm Technol Res. 2022; 13(3):197-201.
- Yunarto N, Intan PR, Kurniatri AA, Sulistyowati I, Aini N. Anti-Inflammatory Activities of Ethyl Acetate Fraction From Uncaria Gambir Leaves Through the Inhibition of Edema, COX-2 and iNOS Expression. Adv Health Sci Res. 2020; 22(Ishr 2019):108–112.
- 11. Taniguchi S, Kuroda K, Doi KI, Tanabe M, Shibata T, Yoshida T, Hatano T. Revised structures of gambiriins A1, A2, B1, and B2, chalcane-flavan dimers from gambir (Uncaria gambir extract). Chem Pharm Bull. 2007; 55(2):268-272.
- 12. Yimam M, Lee YC, Kim TW, Moore B, Jiao P, Hong M, Kim HJ, Nam JB, Kim MR, Oh JS, Cleveland S, Hyun EJ, Chu M, Jia Q. Analgesic and anti-inflammatory effect of UP3005, a botanical composition Containing two standardized extracts of *Uncaria gambir* and *Morus alba*. Pharmacogn Res. 2015; 7(5s):s39–46.
- 13. Dewi SRP and Pratiwi A, Teodorus. The effect of Gambier extracts (Uncaria gambir RoxB.) as antiseptic on gingival wound in rats. ODONTO Dent J. 2018; 5(1):80–87.
- 14. Syarifah S, Widyawati T, Rita Anggraini D, Sari Wahyuni A, Indah Sari M. Anticancer activity of uncaria gambir roxb on T47D breast cancer cells. J Physics: Conf Ser. 2019; 1317(1):012106.
- 15. Yunarto N, Sulistyaningrum N, Kurniatri AA, Elya B. Gambir

(*Uncaria gambir* Roxb.) as A Potential Alternative Treatment for Hyperlipidemia. Media Penelitian dan Pengembangan Kesehatan. 2021; 31(3):183-192.

- M. Jain Kassim, M. Hazwan Hussin, A. Achmad, N. Hazwani Dahon TKS and HSH. Determination of total phenol, condensed tannin and flavonoid contents and antioxidant activity of Uncaria gambir extracts. Maj Farm Indones. 2011;22(1):50–9.
- Melia S, Novia D, Juliyarsi I. Antioxidant and antimicrobial activities of gambir (Uncaria gambir Roxb) extracts and their application in rendang. Pak J Nutr. 2015;14(12):938–941.
- Amir M, Mujeeb M, Khan A, Ashraf K, Sharma D, Aqil M. Phytochemical analysis and in vitro antioxidant activity of Uncaria gambir. Int J Green Pharm. 2012; 6(1):67–72.
- Anggraini T, Tai A, Yoshino T, Itani T. Antioxidative activity and catechin content of four kinds of Uncaria gambir extracts from West Sumatra, Indonesia. Afr J Biochem Res. 2011; 5(1):33–38.
- Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. LWT - Food Sci Technol. 1995; 28(1):25–30.
- 21. Dudonné S, Vitrac X, Coutiere P, Woillez M, Merillon JM. Comparative Study of Antioxidant Properties and Total Phenolic Content of 30 Plant Extracts of Industrial Interest Using DPPH, ABTS, FRAP, SOD, and ORAC Assays. J Agric Food Chem. 2009; 57(5):1768-1774.
- 22. Kristanti AN, Aminah NS, Zahroh FF, Hudaniah KB, Budiman MA, Renaldy R, Indrawan, Alfatsyah R, Wardana AP, Takaya Y. Phytochemistry of *Syzygium polycephalum*. Trop J Nat Prod Res. 2022; 6(5):728–731.
- 23. Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free Radic Biol Med. 1999; 26(9–10):1231–1237.
- Phongpaichit S, Nikom J, Rungjindamai N, Sakayaroj J, Hutadilok-Towatana N, Rukachaisirikul V, Kirtikara, K. Biological activities of extracts from endophytic fungi isolated from Garcinia plants. FEMS Immunol Med Microbiol. 2007; 51(3):517–525.
- 25. Arnao MB. Some methodological problems in the determination of antioxidant activity using chromogen radicals: A practical case. Trends Food Sci Technol. 2000; 11(11):419– 421.