

**Drug Development for the Management of Corona Viruses: Insights from Natural Antiviral Agents**Taiwo O. Elufioye^{1*} and Solomon Habtemariam²¹Department of Pharmacognosy, Faculty of Pharmacy, University of Ibadan, Nigeria²Pharmacognosy Research Laboratories & Herbal Analysis Services UK, University of Greenwich, Chatham-Maritime, Kent ME4 4TB, UK

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ABSTRACT

Infection by human Coronaviruses (CoVs) such as HCoV-229E, HCoV-OC43, HCoV-NL63, HCoV-HKU1 generally results in moderate to severe respiratory and intestinal infections in humans. The deadly human CoVs emerging in the last two decades however became the cause of great global concern. The severe acute respiratory syndrome (SARS) SARS-CoV, Middle East respiratory syndrome (MERS) (MERS-CoV) and the current SARS-CoV-2 that causes COVID-19 are related respiratory infections with high degree of mortality.

With currently no available vaccine or approved therapy for COVID-19, the development of directly acting antiviral drugs becomes a sensible strategy. Viral infections, like many other disease conditions have been frequently managed using traditional medicine. COVID-19 and SARS-CoV-2 infections have also been treated with preparations from traditional Chinese medicine although their efficacy has not yet been well documented. Several antiviral agents have also been reported from natural sources and these could provide good opportunity for developing products and therapies that might be applicable in managing COVID-19.

In this review, we discuss natural antiviral products that target the various infection stages of the different viruses' including CoV, which may be useful for direct management of COVID-19 or provide insights for the development of effective therapies.

Keywords: Natural products, Antiviral, Corona viruses, Drug development, Phytochemicals.

Introduction

Coronaviruses (CoVs) are enveloped viruses that cause multiple respiratory and intestinal infections in humans and animals.¹ The term 'coronavirus' refers to the crown-like appearance of CoVs under electron microscope due to spike projections from the membrane.² These viruses were first described in the 1960s from patients with respiratory infections like the common cold.³ Since then, several human coronaviruses (HCoVs) have been identified including the HCoV-Hong Kong University 1 (HKU1), HCoV-NL63; severe acute respiratory syndrome (SARS)-CoV; and Middle East respiratory syndrome (MERS)-CoV.³ CoVs belong to the subfamilies Coronavirinae and Torovirinae of the family Coronaviridae in the order Nidovirales.⁴ Based on phylogeny, they are divided into four genera as α -CoV, β -CoV, γ -CoV and δ -CoV.⁵ The beta-CoV genus is further subdivided into four lineages (A, B, C, and D)¹ (Figure 1). Mammalian coronavirus include HCoV-229E and HCoV-NL63 which belong to the α -coronavirus class as well as HCoV-HKU1, SARS-CoV, MERS-CoV and HCoV-OC43 which are β -coronaviruses; while avian coronavirus is the γ -coronavirus and δ -coronavirus.³ The novel coronavirus, 2019-nCoV, that caused the current global COVID-19 pandemic was first identified in December 2019 in the Wuhan region (Hubei Province) of China. Since then, the Coronavirus Study Group (CSG) of the International Committee on Taxonomy of Viruses has formally designated the virus as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2).⁶

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The various coronaviruses cause divers infections in human but historically given little attention due to their mild phenotypes. Globally, HCoV 229E, NL63, OC43, and HKU1 are endemic, accounting for 10% to 30% of upper respiratory tract infections in adults.⁷ They rarely cause life-threatening pneumonia and bronchiolitis but this can happen in children and immunocompromised patients.^{8,9} HCoVs have also been linked with enteric and neurological disorders.¹⁰⁻¹² The first major alert on severe cases of HCoVs came in 2002 when lethal atypical pneumonia due to SARS-CoV was reported in China.⁷

Better understanding of the biology and the pathophysiology of CoVs infections have resulted from several researches conducted in the last two decades, especially on SARS-CoVs and MERS-CoVs. This has in turn driven the search for appropriate drug therapy and vaccine development.

The use of natural products derived from plants, animals or micro-organisms for medicinal purpose precedes the recorded human history. Over the years, humankind has discovered and utilized several chemical products of natural origin. Perhaps due to their chemical and biological diversity, the application of natural products for medicine and health throughout evolution has been significant.¹³ It is reported that the single most productive source of lead compounds for drug development is natural product.¹⁴ Newman *et al.*, (2002) reported drugs discovered from natural products between 1981 and 2002, with this accounting for 60% and 75% of new drugs respectively in the areas of cancer and infectious disease.¹⁵ Drugs of natural products origin for managing various disorders such as bacterial and fungal infections, cancer, diabetes, dyslipidemia, atopic dermatitis, Alzheimer's disease, tyrosinaemia and Gaucher disease, introduced for use between 2001 and 2005, have also been reported.¹⁶ Other reviews on the importance and re-emergence of natural products in drug discovery are available.¹⁷⁻²¹ In the last few years, new technologies that made screening of natural products easier thereby helping to overcome certain limitations in natural products drug discovery have been developed.¹⁶ Also, natural products have contributed specifically

to antiviral drug discovery.²² In this review, we focus on various antiviral agents from nature, highlighting their possible use in drug development for the currently ongoing SARS- CoV-2 infection.

Coronavirus genome and replication

CoVs are single-stranded positive-sense RNA (+ssRNA) viruses having the largest genome size so far (~30 kb) with 5'-cap structure and 3'-poly-A tail.²³ During CoVs infection cycle, the genomic RNA plays a critical role in the initial RNA synthesis and acts as a messenger RNA (mRNA).³ It is used as template for the translation of polyprotein 1a/1ab (pp1a/pp1ab) to replication-transcription complex (RTC) in a double-membrane vesicles (DMVs).²⁴ The pp1a/pp1ab also encodes nonstructural proteins (nsps).²⁵ The RTC then produces, in a manner of discontinuous transcription, a set of subgenomic RNAs (sgRNAs) that possess common 5'-leader and 3'-terminal sequences.²⁶ This is followed by the termination of the transcription process and subsequent acquisition of a leader RNA, which takes place at the transcription regulatory sequences found in-between open reading frames (ORFs).²⁷

There are at least six ORFs in a typical CoV. The ORF1a/b constitute about two-thirds of the entire genome length and encodes 16 nsps (nsp1-16) while the ORFs on the remaining one-third of the genome near the 3'-terminus encodes four structural proteins: spike protein (S), a type of glycoprotein I; membrane protein (M), that covers the membrane; envelope protein (E), a hydrophobic protein that covers the entire structure; and nucleocapsid (N) proteins a basic RNA-binding protein.^{3,28} The polypeptides ORFs are processed by chymotrypsin-like protease (3CLpro) or main protease (Mpro) and papain-like protease into 16 nsps that are involved in genome transcription and replication.²⁹

Different CoVs can however encode additional special structural and accessory proteins apart from the four main structural proteins. For instance, viruses in lineage A also encode a smaller protein called hemagglutinin esterase (HE) which has similar function as the S protein.¹ Other special proteins include 3a/b protein and 4a/b protein. The structural proteins S, M, E and N, in addition to facilitating virion assembly, also suppresses host immune response to facilitate viral replication.³⁰ Viral replication starts with the binding of the spike protein (S) onto the host's cell-surface. This is an important recognition step that initiate virus entry into the host cells and different CoVs utilizes different receptors to achieve this. For instance, aminopeptidase N is used by HCoV-229E;³¹ 9-O-acetylated

sialic acid by HCoV-OC43 and HCoV-HKU1,^{32,33} angiotensin-converting enzyme 2 (ACE2) by SARS-CoV;³⁴ and HCoV-NL63^{35,36} and dipeptidyl peptidase 4 (DPP4) by MERS-CoV.³⁷ CoVs also use proteases to facilitate entry into host cell. Proteases used include cathepsin L by SARS- and MERS-CoV viruses, transmembrane protease serine 2 (TMPRSS2) and airway trypsin-like protease TMPRSS11D by HCoV-229E and SARS-CoV viruses.³⁸⁻⁴⁰ The replication process begins after successful entry, with the decoding of viral particle and translation of ORF 1a and 1b into polyproteins pp1a and pp1ab, a process facilitated by 3-C-like protease (3CLpro) and papain-like protease (PLpro). The polyproteins are thereafter split into about 15 NSPs, which subsequently form the replication–transcription complex (RTC). Using replicases, the positive genomic RNA is then transcribed to negative-strand template for the synthesis of new genomic RNAs which are further transcribed to form structural and accessory proteins. Chemical compounds capable of interrupting any stage in the replication process would be a good candidate for therapeutic development.

Mechanisms against human corona viruses

Mechanisms directed at combating human corona virus infections include elimination of the viruses directly using antiviral agents and modulating host's immune system to be able to fight the viruses.

Antiviral agents can be designed to target different stages of the virus replication process such as cell entry, viral transcription as well as translation and protein processing (Figure 2). The coronavirus S protein is used for binding to the cellular receptor through the receptor binding site (RBS) resulting in fusion with the cellular membrane. Following this binding, the virus can then enter their target cells either by fusion of viral membranes onto the cell surface or through endocytosis and subsequent acidification of the environment.⁴¹ This provides two distinct mechanisms of interrupting virus: inhibition of cell attachment/binding and inhibition of cell-virus fusion. Another therapeutic approach can involve inhibition and downregulation of the various viral receptors such as the ACE receptor.⁴² Though complex, CoVs replication process is also a therapeutic target. Possible therapies that can be developed include polymerase inhibitors, nucleoside analogues, helicase inhibitors and small interfering RNA and antisense phosphorodiamidate morpholino oligomers.⁴³ Translation and protein processing such as by chymotrypsin-like proteinase and papain-like proteinase are also good anti-coronavirus targets.⁴⁴

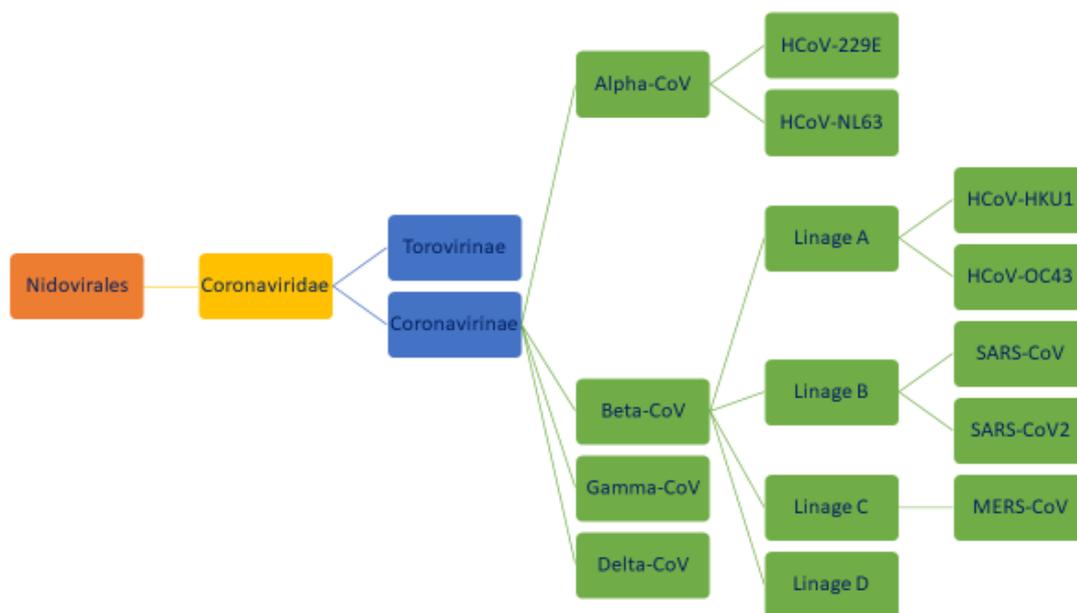


Figure 1: Schematic representation of the human coronaviruses

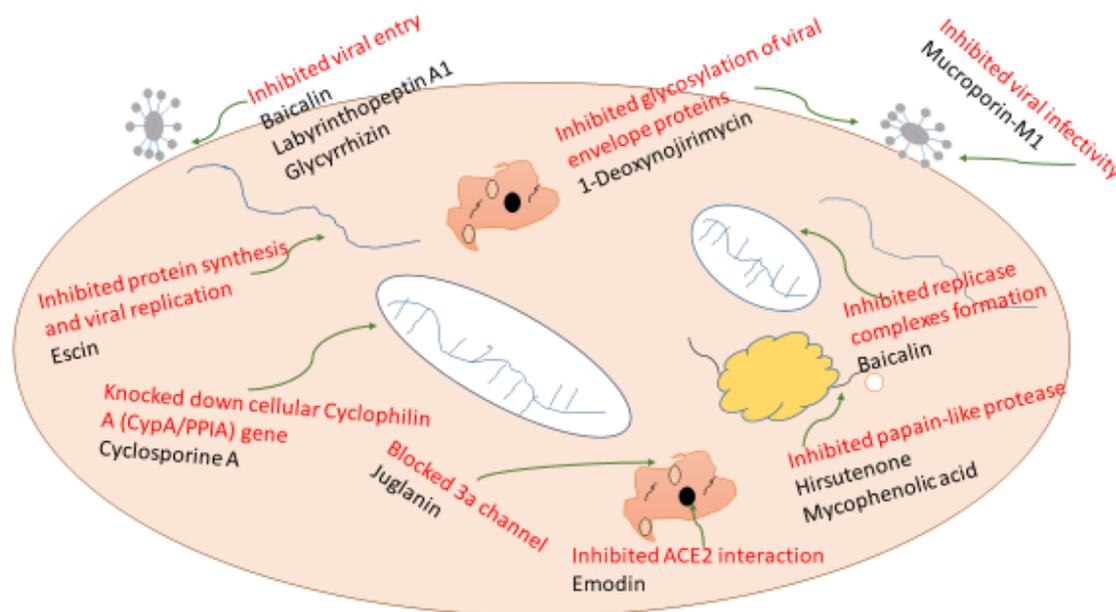


Figure 2: Potential Pharmacologic antiviral targets against SARS-CoV-2

Overview of the Current Pharmacotherapy of Covid 19

There is currently no consensus globally regarding effective therapies for managing SARS-CoV-2 infection. However, several therapies have been used in different parts of the world and many existing drugs are in clinical trial with a view of being repurposed. In the United States, remdesivir over no antiviral treatment and glucocorticoids rather than no glucocorticoids is suggested for use among hospitalized patients with severe COVID-19; while hydroxychloroquine/chloroquine plus azithromycin, lopinavir/ritonavir, tocilizumab, famotidine and COVID-19 convalescent plasma are to be used only in the context of a clinical trial.⁴⁵ In India, hydroxychloroquine is recommended as prophylaxis for asymptomatic healthcare personnel handling COVID-19 cases, frontline workers, and asymptomatic contacts of the confirmed cases, while hydroxychloroquine-azithromycin combination is used for patients with serious sickness requiring ventilator.⁴⁶ In other countries of the world, different therapy combinations have been employed as summarized by Chen *et al.*, (2020)⁴⁷ and guidelines for care by different relevant societies are available here <https://www.uptodate.com/contents/society-guideline-links-coronavirus-disease-2019-covid-19-international-and-government-guidelines-for-general-care>. There are also several review articles on the different therapeutic options.⁴⁸⁻⁵¹

Antiviral drugs of natural origin and their mechanisms of action

Several drugs and bioactive compounds of natural origin have been previously reported active against many viruses. The application of some of these molecules, whose information are as summarized in Table 1, in drug development against coronavirus is hereby discussed.

1-Deoxynojirimycin

1-Deoxynojirimycin (1) is a piperidine alkaloid found in various plants such as *Commelina communis*, and in the *Streptomyces* and *Bacillus* bacteria.^{52,53} It has antihyperglycemic, anti-obesity, and antiviral activities.⁵⁴ It has been reported to be effective against hepatitis,⁵⁵ influenza⁵⁶ and HIV.⁵⁷ 1-Deoxynojirimycin inhibits cellular α -glucosidase I-II activity. It also inhibited the envelope-mediated membrane fusion process at the CXCR4 binding step thus preventing the spread of human

immunodeficiency virus in (HIV)-infected lymphocyte cultures.⁵⁷ In fowl plague virus-infected chicken-embryo cells, *N*-methyl-1-deoxynojirimycin inhibited the trimming of the outermost glucose residue of the *N*-linked precursor-oligosaccharide $\text{Glc}_3\text{Man}_9\text{GlcNAc}_2$, thus inhibiting oligosaccharide processing.⁵⁸ The envelope-mediated membrane fusion inhibitory mechanism of this compound can be explored in designing possible therapy for coronavirus and/or SARS-CoV-2.

Aloe-emodin and Emodin

Emodin (3) is an anthraquinone obtained from the genus *Rheum* and *Polygonum*.⁵⁹ It is known for its larvicidal⁶⁰ and anticancer⁶¹ activities. It was investigated against SARS-CoV and reported to significantly inhibit the S protein and ACE2 interaction in a dose-dependent manner.⁵⁹ Evaluation against herpes simplex virus (HSV) infections revealed that emodin inhibited the nuclease activity of HSV-1 UL12 as well as reduced plaque formation with an EC_{50} of 21.5 ± 4.4 mM.⁶² The effect of emodin on herpes virus was also confirmed by Xiong *et al.* (2011).⁶³

The study by Li *et al.* (2014)⁶⁴ on the effect and inhibitory mechanism of aloe-emodin (2) against influenza A virus revealed that aloe-emodin could reduce virus-induced cytopathic effect dose-dependently with an IC_{50} value less than 0.05 $\mu\text{g}/\text{mL}$. It also up-regulated galectin-3 and thioredoxin as well as down-regulated nucleoside diphosphate kinase A.⁶⁴ It restored nonstructural protein 1 (NS1)-inhibited signal transducer and activator of transcription 1 (STAT1)-mediated antiviral responses in transfected cells, for instance, STAT1 phosphorylation of interferon (IFN) stimulation response element (ISRE)-driven promoter, RNA-dependent protein kinase (PKR) and 2',5'-oligoadenylate synthetase (2',5'-OAS) expression.⁶⁴ Aloe-emodin was identified as a potential interferon (IFN)-inducer when tested against Japanese encephalitis virus (JEV) and enterovirus 71 (EV71).⁶⁵ It activated interferon-stimulated response element (ISRE) and gamma-activated sequence (GAS)-driven cis-reporting systems. It also up-regulated expression of IFN-stimulated genes such as dsRNA-activated protein kinase and 2',5'-oligoadenylate synthase as well as activated nitric oxide production.⁶⁵ The several antiviral mechanisms of action of emodin (3) and aloe-emodin (2) against different viruses makes them a promising source of antiviral therapy. In particular,

emodin inhibited the S protein and ACE2 interaction of SARS-CoV while aloe-emodin restored NS1-inhibited signal transducer both of which can be adapted in developing suitable medication against coronavirus and/or SARS-CoV-2.

Amentoflavone

Amentoflavone (**4**) is a biflavone isolated from the ethanol extract of *Torreya nucifera*.⁶⁶ It is reported to have many biological activities such as anti-inflammatory, anti-oxidative, and anti-diabetic activities.⁶⁷ It has also been reported to be active against SARS-CoV by inhibiting SARS-CoV 3CLpro.⁶⁶ Amentoflavone (**4**) also showed inhibitory activity against human immunodeficiency virus (HIV)⁶⁸ and respiratory syncytial virus (RSV).⁶⁹ Other antiviral activities reported for this compound are anti-dengue,⁷⁰ anti-influenza and anti-herpes.⁷¹ Flavonoids in general are promising leads in drug development.

Apigenin

Apigenin (**5**) is a flavonoid found in many plants such as vegetables (onions; *Allium cepa*), fruits (oranges; *Citrus aurantifolia*), and herbs (basil; *Ocimum basilicum*). It is associated with many health-promoting effects/therapeutic functions.⁷² The antiviral effect of apigenin (**5**) has been reported against enterovirus-71 (EV71),⁷³ hepatitis C virus (HCV),⁷⁴ Foot-and-mouth disease virus (FMD),⁷⁵ African swine fever virus (ASFV),⁷⁶ and herpes virus HSV-2.⁷⁷ It has also been implicated as constituent of herbal products used in the management of corona virus.^{78,79} In an *in silico* study, it was reported to have moderate binding energy on corona virus.⁸⁰ Mechanisms of antiviral action of apigenin include reducing the expression levels of mature MicroRNA122 (miR122) through inhibition of TAR RNA-binding protein (TRBP) phosphorylation,⁷⁴ blocking EV71 RNA association with hnRNP A1 and A2 proteins,⁷³ and inhibition of viral internal ribosome entry site driven translation.⁷⁵ *In silico* binding energy study is a fair prediction of the possible use of a candidate molecule. Moreover, the ability of apigenin to inhibit viral translation is a promising mechanism for drug development against coronavirus and/or SARS-CoV-2.

Baicalin

Baicalin (**6**) a flavone glycoside, is the glucuronide of baicalein and is found in many plant species of the genus *Scutellaria*, such as *S. baicalensis*, *S. lateriflora* and *S. galericulata*.^{81,82} Baicalin is associated with several pharmacological activities including antioxidant, antihypertensive, antifungal and anticancer effects.⁸³ It is also found to be effective in treating cerebral ischemia.⁸⁴ The antiviral activity against dengue virus (DENV),⁸⁵ influenza virus,⁸⁶ HIV-1,^{87,88} enterovirus 71 (EV71),⁸⁹ chikungunya virus (CHIKV),⁹⁰ and SARS-CoV⁹¹ is well established. Liu *et al.*, (2008)⁹² reported that the 4'-OH, 7-OH, C4 keto and C2-C3 double bond functionalities of flavonoids were essential for their anti-influenza effect. Testing of baicalin by viral neutralization assay against 10 strains of SARS-CoV in fRhK4 cell line and against the prototype strains (39849) of SARS-CoV in fRhK4 and Vero-E6 cell lines revealed that it was active. It was also active in the plaque reduction assay with an EC₅₀ value of 11 µg/mL.⁹¹ This, combined with the numerous antiviral activities of baicalin (**6**), makes it a promising candidate for antiviral drug development. Moreover, baicalin capsules (250 mg per capsule) is an approved drug by the state food and drug administration of China as an adjuvant therapy for managing hepatitis.⁹³

Betulinic acid

Betulinic acid (**7**) is a triterpenoid found in several plants such as *Betula pubescens*.⁹⁴ It has anticancer,^{95,96} antimalarial⁹⁷ and anti-inflammatory activities.⁹⁸ The antiviral efficacy against HIV-1,⁹⁹ herpes simplex type I, influenza FPV/Rostock and ECHO 6 viruses,¹⁰⁰ Herpes simplex virus¹⁰¹ have been reported. Several derivatives of betulinic acid (**7**) are potent and highly selective inhibitors of HIV-1 whose mechanism of action include inhibition of HIV fusion and interference with a specific step in HIV-1 maturation.⁹⁹ Synthetic derivatives containing nicotinoyl-, methoxycinnamoyl-, alkyne and aminopropoxy-2-cyanoethyl-moieties of betulin have been produced through structure modifications at positions C-3, C-20 and C-28.

Antiviral activity was reported for 3β,28-di-*O*-nicotinoylbetulin against human papillomavirus type 11 while the 3β,28-Dihydroxy-29-norlup-20(30)-yne derivative was active against HCV replicon and the 28-*O*-Methoxycinnamoylbetulin derivative was active against influenza type A virus.¹⁰² The antiviral effect of betulinic acid against enveloped virus along with its possible structure modification associated with increased antiviral activity can be an added benefit in designing a drug against coronavirus and/or SARS-CoV-2.

Castanospermine

Castanospermine (**8**) is an indolizidine alkaloid obtained from the seeds of *Castanospermum australe*.¹⁰³ It is known to inhibit β-glucosidase and β-glucocerebrosidase.¹⁰⁴ The antiviral effect of castanospermine has been documented against HIV,¹⁰⁵ dengue virus,¹⁰⁶ and influenza.¹⁰⁷ It decreased viral replication by inhibiting syncytium formation induced by the envelope glycoprotein of the human immunodeficiency virus. This is achieved by inhibiting the processing of the envelope precursor protein gp160, with resultant decreased cell surface expression of the mature envelope glycoprotein gp120. Also, castanospermine caused defects in steps involved in membrane fusion after binding of CD4 antigen.¹⁰⁵ Castanospermine inhibited dengue virus infection by preventing secretion and infectivity of viral particles. It also prevented mortality in a mouse model of dengue virus infection.¹⁰⁶ Several synthetic derivatives with antiviral effect are available.¹⁰⁸ This makes it easier to develop new therapeutics that can inhibit the processing of envelope precursor protein gp160 from this molecule, which may be useful in treating coronavirus and/or SARS-CoV-2 infection.

Chebularic acid

Chebularic acid is a tannin obtained from *Terminalia* species like *T. chebula*, *T. citrina* and *T. catappa*.¹⁰⁹ It has been reported as an immunosuppressive,¹¹⁰ hepatoprotective¹¹¹ and α-glucosidase inhibitor.^{112,113} The compound has satisfactory activity against several viruses including herpes simplex virus,¹¹⁴ Human enterovirus 71,¹¹⁵ HIV,¹¹⁶ and Influenza.¹¹⁷ Chebularic acid inhibits Herpes simplex virus 1 (HSV-1) entry in A549 human lung cells by targeting and inactivating HSV-1 viral particles thus preventing binding, penetration, cell-to-cell spread and secondary infection. The inhibitory action targets at HSV-1 glycoproteins since it blocked polykaryocyte formation mediated by expression of recombinant viral glycoproteins involved in attachment and membrane fusion.¹¹⁴ Its broad spectrum of activity was demonstrated against human cytomegalovirus (HCMV), hepatitis C virus (HCV), dengue virus (DENV), measles virus (MV), and respiratory syncytial virus (RSV), where it inhibited viral attachment, penetration, and spread supposedly through host cell Glycosaminoglycans cell entry mechanism.¹¹⁸ Chebularic acid (IC₅₀ = 1.41 ± 0.51 µg/mL) and chebulinic acids (IC₅₀ = 0.06 ± 0.002 µg/mL) showed dose-dependent potent *in vitro* direct anti-viral activity against HSV-2 by preventing the attachment and penetration of the HSV-2 to Vero cells.¹¹⁹ Against influenza A virus, chebularic acid and chebulinic acid inhibited viral replication through inhibiting neuraminidase-mediated viral release.¹¹⁷ Basically, this compound's broad antiviral effect is due to its ability to block viral fusion and entry, two important targets in virus life cycle for drug development against coronavirus and/or SARS-CoV-2.

Cyanovirin-N

Cyanovirin-N (CV-N) is a protein produced by the cyanobacterium *Nostoc ellipsosporium*.¹²⁰ It is an elongated, largely β-sheet protein that displays internal two-fold pseudosymmetry.¹²¹ Multiple antiviral activities have been associated with this compound. Cyanovirin-N inactivated different HIV-1 strains and other lentiviruses due to its irreversible binding to the viral envelope glycoprotein gp120.¹²² Tsai *et al.*,¹²³ reported that recombinant CV-N effectively blocks HIV-1Ba-L infection of human ectocervical explants thus making it a potential candidate for testing in humans as an anti-HIV topical microbicide. It has also been reported to target N-linked high-mannose oligosaccharides found on the viral envelope of HIV-1, providing possible explanation for its broad antiviral activity.¹²⁴ It binds to hepatitis C viral envelope glycoproteins and blocked the interaction

between the envelope protein E2 and CD81.¹²⁵ Against influenza virus, it showed considerable activity through binding with viral hemagglutinin.¹²⁶ It also inhibits infectivity of Ebola virus by binding to the viral surface glycoprotein, GP1,2.¹²⁷ This compound can be explored against coronavirus and/or SARS-CoV-2 due to its ability to bind viral envelope.

Cyclosporine A

Cyclosporine A is a cyclic peptide isolated from the fungus *Hypocladium inflatum* *gams*.¹²⁸ It is known for its immunosuppressant,¹²⁹ antimicrobial,¹³⁰ anti-psoriasis¹³¹ activities. It was officially approved in 1997 by the US Food and Drug Administration for the treatment of plaque psoriasis.¹³² The antiviral activity of cyclosporine A has been reported against HIV. It has been shown to inhibit HIV-1 replication by binding with cyclophilin A and thus disrupting the ability of cyclophilin A to interact with HIV-1 Gag polyprotein.¹³³ It also blocked HIV-1 infectivity by blocking HIV-1 capsid (CA) interaction with target cell cyclophilin A (CypA) as well as decreased gp120 and gp41 incorporation into HIV-1 virions thus impairing the fusion of these virions with susceptible target cells.¹³⁴ Several reports are available on the effects of cyclosporine A against hepatitis C virus (HCV).^{135,136} It also inhibits the replication of Japanese Fulminant Hepatitis (JFH1) full-length genomes much more efficiently than subgenomic replicons by targeting cleavage at the nonstructural 2/nonstructural 3 (NS2/NS3) junction.¹³⁷ It also inhibited hepatitis B replication by interacting with mitochondria, preventing the release of inter-mitochondria calcium, and blocking cytosolic calcium signaling.¹³⁸ Against influenza virus, it also inhibited the replication of influenza A virus through cyclophilin A (CypA)-dependent and -independent pathways.¹³⁹ Furthermore, it inhibits the growth of human coronavirus HCoV-NL63 through cyclophilin A pathway.¹⁴⁰ Other viruses against which cyclosporine has been found effective include *Herpes simplex*,¹⁴¹ stomatitis,¹⁴² vaccinia,¹⁴³ cytomegalovirus¹⁴⁴ and human papilloma virus.¹⁴⁵ Its effectiveness in preventing viral fusion and replication, coupled with the fact that it has also been found effective against human coronavirus HCoV-NL63 makes this molecule a valuable candidate for consideration in developing anti-coronavirus and/or SARS-CoV-2 drug.

Droserone

Droserone (**9**) is a naphthoquinone obtained from dicotyledonous plants.¹⁴⁶ It is one of the pigments in *Drosera whittakeri*.¹⁴⁷ The compound has anti-fungal,¹⁴⁸ bactericidal¹⁴⁹ and weak antimalarial¹⁵⁰ properties. It has been reported to reduce measles virus entry considerably suggesting its interaction with viral particles to reduce infectivity.¹⁴⁶ Some synthetic Bis-Naphthoquinones have also been reported effective against Zika virus¹⁵¹ and HSV-1¹⁵² suggesting possible use of naphthoquinones as antiviral lead compounds. Optimizing synthetic derivatives of this compound as agents that can block viral entry is a possible consideration for anti-coronavirus and/or SARS-CoV-2 drug development.

Escin

Escin is a triterpene saponin mixture from the horse chestnut, *Aesculus hippocastanum*.¹⁵³ The anti-edematous, anti-inflammatory and venotonic properties of β -escin have been linked to induction of cholesterol synthesis followed by decreased actin cytoskeleton integrity leading to reduced responses to tumor necrosis factor alpha (TNF α) stimulation, including reduced migration, alleviated endothelial monolayer permeability, and inhibition of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) signal transduction leading to downregulation of TNF- α -induced effector proteins' expression.¹⁵⁴ It also has anticancer activity through intrinsic-mitochondrial apoptosis pathway by arresting G2/M and ROS generation.¹⁵⁵ The antiviral activity of escin has been reported against porcine epidemic diarrhea virus (PEDV),¹⁵⁶ HIV-1¹⁵⁷ and SARS-CoV.¹⁵⁸ Strong cytotoxicity of escins create a challenge for their drug development.¹⁵⁹ However, SAR studies have shown that acylations at C-21 and C-22 with angeloyl or tigloyl groups were important for their cytotoxic effects.¹⁵⁶ Thus, escin derivatives with strong antiviral activities have been reported.¹⁵⁶ Moreover, it is an

approved, well tolerated anti-edematous, anti-inflammatory drug.¹⁵³ Thus, adapting it as a drug template for repurposing against coronavirus and/or SARS-CoV-2 can be explored.

Genistein

Genistein (**10**) is an isoflavone originally from *Genista tinctoria* but also found to occur in soybeans, *Glycine max*.¹⁶⁰ It has been implicated in the treatment of menopausal vasomotor symptoms,¹⁶¹ cancer,¹⁶² and memory loss¹⁶³ among other uses. Antiviral studies indicated that it inhibits African swine fever virus replication by disrupting viral DNA synthesis.¹⁶⁴ It also prevented plaque formation of Herpes B virus and reduced virus production.¹⁶⁵ It was reported to show some efficacy against H1N1 influenza A virus.¹⁶⁶ Against human immunodeficiency virus type 1 (HIV-1), it caused cell cycle arrest in G2.¹⁶⁷ It was also reported effective against Lassa virus and Ebola virus infections.¹⁶⁸ Other viruses against which it has been found effective include avian leucosis virus¹⁶⁹ and Moloney murine leukemia virus (Mo-MLV).¹⁷⁰ The broad spectrum of antiviral effect of this molecule makes it a good candidate for testing against coronavirus and/or SARS-CoV-2.

Geraniin

Geraniin is a dehydroellagitannin isolated from *Geranium carolinianum*. Pharmacological activities associated with geraniin include antioxidant, liver protective, anticancer properties among others.¹⁷¹ Geraniin inhibits the replication of human immunodeficiency virus-1 (HIV-1) *in vitro* by inhibiting virus uptake and HIV-1 reverse transcriptase.¹⁷² It was also reported to suppress herpes simplex virus (HSV) with better activity on HSV-2 infection than HSV-1.¹⁷³ It had antiviral activity against hepatitis B virus (HBV). It has been shown to inhibit hepatitis B surface antigen (HBsAg) and hepatitis Be antigen (HBeAg) secretion by about 85.8% and 63.7%, respectively, at a concentration of 200 mg/mL.¹⁷⁴ Geraniin also had inhibitory effect on human enterovirus 71 (EV71).¹⁷⁵ The diverse effects against many viruses as well as different mechanisms of action makes it suitable for drug development against coronavirus and/or SARS-CoV-2.

Glycyrrhizin

Glycyrrhizin (**11**) is a triterpenoid saponin isolated from licorice (*Glycyrrhiza glabra*, *Glycyrrhiza inflata* and *G. uralensis*) root.¹⁷⁶ It is reported to have several biological activities including antiviral against varicella-zoster virus (VZV),¹⁷⁷ human immunodeficiency virus type 1 (HIV-1) and herpes simplex virus type 1 (HSV-1),¹⁷⁸ Hepatitis virus (HCV)¹⁷⁹ and coronavirus^{180,181} among others. Glycyrrhizin (**11**) is known to affect many cellular signaling pathways¹⁸²⁻¹⁸⁴ with one other possible mechanism of anti-coronavirus action being stimulation of inducible NO synthase (iNOS) expression and subsequent increase of the nitric oxide (NO) concentration.¹⁸⁵ Nitric oxide (NO) plays diverse physiological functions in the body and there are evidences suggesting that NO and oxygen radicals are involved in the pathogenesis of various infectious diseases.^{186,187} NO biosynthesis through expression of inducible iNOS occurs in different microbial infections and iNOS produce large amount of NO over a long period. The production of NO leads to the generation of a highly reactive nitrogen oxide species, peroxynitrite, through radical coupling reaction of NO with superoxide and the generated peroxynitrite causes oxidative tissue injury through potent oxidation and nitration reactions of various biomolecules.¹⁸⁸ NO is also known to modulate host's immune response¹⁸⁹ thus acting as host response modulator rather than a simple antiviral agent.¹⁸⁸ Glycyrrhizin is an already approved drug for clinical use as an intravenous medication and it is commercially available.¹⁹⁰ Thus it can be easily adapted for use as possible anti-coronavirus and/or SARS-CoV-2 drug.

Guggulsterone

Guggulsterone (**12**) is a phytosteroid found in the resin of the guggul plant, *Commiphora mukul*.¹⁹¹ It has anti-inflammatory¹⁹² and anticancer¹⁹³ activities. Guggulsterone (**12**) isolated from *Commiphora gileadensis* was reported to be responsible for the antiviral activity of the plant against herpes simplex virus type 2 (HSV-2), respiratory

syncytial virus type B (RSV-B), coxsackie virus B type 3, and adenovirus type 5.¹⁹⁴ High serum level of bile acids is known to be responsible for antiviral therapy failure in patients with hepatitis C (HCV) infection. Also, bile acids are important for the replication of the porcine enteric calicivirus. Free, and not conjugated bile acids up-regulated genotype 1 HCV RNA replication suggesting that this effect was mediated by a nuclear receptor. Guggulsterone inhibited basal level of HCV replication as well as blocked bile acids-induced up-regulation of genotype 1 HCV RNA replication through Farnesoid X receptor (FXR) silencing and FXR antagonism.¹⁹⁵ Enveloped viruses such as influenza viruses, alphaviruses and coronaviruses, that uses host cells endocytic pathway, have fusion protein that is activated at low pH made possible in the presence of bile acids.¹⁹⁶ Thus, guggulsterone might be a promising molecule in drug development for coronavirus and/or SARS-CoV-2 through the blockage of bile acid-induced viral replication.

Hirsutenone

Hirsutenone (**13**) is a diarylheptanoids obtained from the stem bark of *Alnus japonica*.¹⁹⁷ It has been reported for its anticancer,¹⁹⁸ anti-dermatitis¹⁹⁹ and regulation of osteoclastogenesis.²⁰⁰ It was found active against SARS-CoV by inhibiting papain-like protease of SARS-CoV.¹⁹⁶ Diarylheptanoids are a broad class of plant structurally divergent phenolics with diverse therapeutic applications.²⁰¹ Park *et al.*¹⁹⁷ correlated good antiviral activity with the α,β -unsaturated carbonyl group and catechol moiety while observing that monohydroxyl substitution and glycosidation led to reduced activity. Assessment against recombinant SARS-CoV 3CLpro also showed that Hirsutenone had good selectivity towards the coronaviral proteases. Thus, Hirsutenone can be a potential drug target for the treatment of SARS disease.

Juglanin

Juglanin (**14**) is a flavonol isolated from *Juglans mandshurica*²⁰² and has also been identified in *Polygonum aviculare*.²⁰³ It has been reported to possess anti-inflammatory²⁰⁴ and anticancer²⁰⁵ activities. The antiviral effect against some virus infections are also available in literature. It was evaluated for its ability to block the 3a channel of SARS coronavirus. Juglanin (**14**) was effective with an IC₅₀ value of 2.3 μ M for inhibition of the 3a-mediated current.²⁰⁶ In an *in silico* experiment against influenza virus, it was shown to have binding specificities to hemagglutinin (H7) and neuraminidase (N9).²⁰⁷ The effectiveness of this compound to block 3a channel of SARS coronavirus can be used in developing effective therapy against coronavirus and/or SARS-CoV-2 infection.

Labyrinthopeptin

Labyrinthopeptins are peptides produced by actinomycete *Actinomyces namibiensis*.²⁰⁸ Labyrinthopeptin A1 showed broad anti-HIV activity and anti-HSV (herpes simplex virus) activity. It inhibited viral cell-cell transmission between persistently HIV-infected T cells and uninfected CD4+ T cells as well as the transmission of HIV captured by DC-SIGN+ cells to uninfected CD4+ T cells.²⁰⁹ Labyrinthopeptins A1 and A2, were found to inhibit the proliferation of many enveloped viruses including dengue virus, Zika virus, West Nile virus, hepatitis C virus, chikungunya virus, Kaposi's sarcoma-associated herpes virus, cytomegalovirus, and herpes simplex virus with mechanism of action showing that it induced vireolytic effect through binding to the viral membrane lipid phosphatidylethanolamine (PE).²¹⁰ Several derivatives are currently available for treating different viral infections,²¹¹ which makes it a promising compound for assessment against coronavirus and/or SARS-CoV-2 infection.

Lectins

The high glycosylation of the S protein makes it a good target for compounds like plant lectins, that can bind to sugar moieties thus forming a coat around the protein and blocking possible interaction with the receptor.⁴³ Lectins are known inhibitors of glycosylated viruses like HIV-type 1, cytomegalovirus and human T-cell leukemia virus.²¹²⁻²¹⁴ *Galanthus nivalis* (Common Snowdrop), *Narcissus tazetta*, *Hippeastrum hybrid* (Amaryllis) and *Allium porrum* (leek) are sources

of lectins that inhibited the replication of SARS-CoV, syncytial virus and feline coronavirus by blocking the S protein-receptor interaction.²¹⁵⁻²¹⁸ They can be explored in the ongoing search for good antiviral therapy for SARS-CoV-2.

Luteolin

Luteolin (**15**) is a flavone obtained from several plant species. It has been reported to show neuropharmacological,²¹⁹ anticancer²²⁰ and anti-inflammatory²²¹ activities, among others. The antiviral effect against HBV²²² and Herpes virus HSV-2²⁷ has also been documented. In HepG2.2.15 cells, luteolin inhibited the expression of hepatocyte nuclear factor 4 α (HNF4 α) and its binding to the HBV promoters and in a HBV replication mouse model, it decreased the levels of HBsAg, HBeAg, HBV DNA replication intermediates, and the HBsAg and HBcAg expression.²²² In a time-of-addition assay, it was shown to interfere with viral replication at the early stage of infection as well as suppressed coat protein I complex expression.²²³ Against Japanese encephalitis virus (JEV), luteolin inhibited viral replication in A549 cells with IC₅₀ = 4.56 μ g/mL and had extracellular virucidal activity on JEV.²²⁴ Luteolin (**15**) inhibited dengue fever virus replication of all four serotypes. It reduced infectious virus particle formation, but not viral RNA synthesis in Huh-7 cells. It also prevented virion maturation process by inhibiting the protease furin responsible for cleaving the pr moiety from prM protein of immature virus particles in the *trans*-Golgi network in the process of producing mature virions.²²⁵ Through a two-step screening platform consisting of reporter virus-based assays and cell viability-based assays, luteolin was shown to exhibit potent inhibition of viral infection against enterovirus 71 and coxsackie virus A16. In the same study, cell viability assay and plaque reduction assay showed that it had an EC₅₀ values of about 10 μ M. The mechanism of action was described as targeting the post-attachment stage of EV71 and CA16 infection by inhibiting viral RNA replication.²²⁶ In another study, luteolin was reported to profoundly reduce HIV-1 infection in reporter cells and primary lymphocytes with the inhibition by luteolin independent of viral entry. It also showed antiviral effect in a latent HIV-1 reactivation model and removed both clade-B- and -C-Tat-driven LTR transactivation in reporter assays.²²⁷ Luteolin inhibited protein expression from Epstein-Barr virus (EBV) lytic genes in EBV-positive epithelial and B cell lines, reduced the numbers of EBV-reactivating cells detected by immunofluorescence analysis as well as reduced virion production.²²⁸ The study concluded that luteolin inhibited EBV reactivation by repressing the promoter activities of Zta (Zp) and Rta (Rp) genes.²²⁸ The diverse mechanisms of anti-viral action of luteolin, particularly inhibition of viral RNA replication and virion maturation makes it a promising candidate for drug development against coronavirus and/or SARS-CoV-2.

Mucroporin-M1

Mucroporin is a cationic host defense peptide isolated from the venom of *Lychas mucronatus* and mucroporin-M1 was designed by amino acid substitution based on its molecular template.²²⁹ Mucroporin-M1 has been evaluated for a variety of activities including antibacterial effect.²²⁹ This compound has also been shown to be effective against several viral infections. The antiviral activities of mucroporin-M1 against measles, SARS-CoV and influenza H5N1 viruses were reported with an EC₅₀ of 7.15 μ g/mL (3.52 μ M) and a CC₅₀ of 70.46 μ g/mL (34.70 μ M) against measles virus, an EC₅₀ of 14.46 μ g/mL (7.12 μ M) against SARS-CoV and an EC₅₀ of 2.10 μ g/mL (1.03 μ M) against H5N1 and the mechanism of action proposed to be inhibition of virus infectivity.²³⁰ Using both *in vitro* and *in vivo* studies, Zhao *et al.*²³¹ established that mucroporin-M1 inhibited hepatitis B virus replication by activating the mitogen-activated protein kinase (MAPK) pathway and down-regulating HNF4 α . The therapeutic potentials of peptides have gained attention lately and investigation into their mechanisms of action with respect to establishing antiviral prospect is on the rise as a result of the global threat posed by viruses. Mucroporin M1 is proposed to directly interact with the virus envelope leading to decreased infectivity. Thus, rational modification of mucroporin would be a good approach to

developing antiviral agents with broad spectrum of activity against RNA viruses particularly SARS-CoVs.

Mycophenolic acid

Mycophenolic acid (**16**) which belongs to the class of organic compounds known as phthalides was first isolated from the fungus *Penicillium stoloniferum* and is an inhibitor of nucleic acid synthesis.²³² It is known as an immunosuppressant²³³ and is an FDA approved immunosuppressive drug. It also has anti-fungal²³⁴ and antitumor²³⁵ effects. The antiviral effect against some unrelated viruses was reported by Planterose (1969).²³⁶ Its inhibitory effect on Dengue virus was attributed to its ability to prevent the synthesis and accumulation of viral RNA.²³⁷ Smee *et al.*²³⁸ reported its effect against orthopoxviruses. It inhibited hepatitis E virus (HEV) replication through nucleotide depletion.²³⁹ It also inhibited HIV replication by depleting the substrate (guanosine nucleotides) for reverse transcriptase and depletion of the pool of activated CD4+ T lymphocytes.²⁴⁰ Hepatitis C virus (HCV) replication was shown to be inhibited by mycophenolic acid at 1.0–6.0 µg/mL to approximately 75% through a mechanism independent of guanosine depletion.²⁴¹ Mycophenolic acid (**16**) significantly inhibited Japanese encephalitis virus both *in vitro* and *in vivo*,²⁴² and Avian reoviruses (ARV).²⁴³ Mycophenolic acid inhibited papain-like protease (PLpro) of MERS-CoV.²⁴⁴

Niranthin

Niranthin (**17**) is a lignan isolated from plants of the genus *Phyllanthus* like *P. niruri* and *P. amarus*. It has been reported with anti-inflammatory activity.²⁴⁵ Initial screening revealed that it had anti-hepatitis activity.²⁴⁶ The anti-hepatitis B virus activity of niranthin (**17**) again evaluated both *in vitro* and *in vivo* showed that it significantly decreased the secretion of HBV surface antigen (HBsAg) and HBVe antigen (HBeAg) with IC₅₀ values of 15.6 µM for HBsAg and IC₅₀ values of 25.1 µM for HBeAg. It also reduced the serum duck hepatitis B virus (DHBV) DNA, HBsAg, HBeAg, ALT and AST *in vivo*²⁴⁷ suggesting that niranthin (**17**) acts as an anti-HBV agent through at least two or more targets. Anti-virucidal evaluation against the white spot syndrome virus showed it inactivated the virus.²⁴⁸ It is thus a potential inhibitor that can be evaluated against SARS-CoV antigen.

Podophyllotoxin

Podophyllotoxin (**18**) is an aryltetralin-type lignan obtained from the resin of *Podophyllum peltatum* L. or *P. emodi*.²⁴⁹ It has several activities including insecticidal²⁵⁰ and anticancer²⁵¹ activities. Antiviral evaluation on herpes simplex virus type I (HSV/CV-1) and vesicular stomatitis virus infecting fibroblasts of hamster kidney (VSV/DHK) showed activity at concentration below 1 µM.²⁵² Several analogues also showed against herpes simplex virus type II.²⁵³ Chen *et al.*²⁵⁴ also reported the anti-HIV effect of podophyllotoxin derivatives. Podophyllotoxin and its derivatives could be exploited in the search for antiviral therapy in general and against coronavirus and/or SARS-CoV-2 in particular.

Quercetin

Quercetin (**19**) is a flavonoid found in many plant species. Numerous biological activities are reported for quercetin. This include neuronal protective,²⁵⁵ hypoglycemic,²⁵⁶ antioxidant²⁵⁷ and antidiabetic²⁵⁸ effects. Different flavonoids including quercetin (**19**) were tested for their effect on infectivity and replication of herpes simplex virus type 1 (HSV-I), polio-virus type 1, parainfluenza virus type 3 (Pf-3), and respiratory syncytial virus (RSV). Quercetin (**19**) caused a dose-dependent reduction in the infectivity and intracellular replication of each virus.²⁵⁹ It also, inhibited HIV-infection by preventing binding of gp120 to CD4.²⁶⁰ Again, the 4'-OH, 7-OH, C4 keto, and C2-C3 double bond group of flavonoids⁹¹ would come to play here and the broad spectrum of activity of quercetin (**19**) will be a major advantage in it prospect as a good antiviral drug template against coronavirus and/or SARS-CoV-2.

Reserpine

Reserpine (**20**) is an indole alkaloid obtained from the roots of *Rauwolfia serpentina* and *Rauwolfia vomitoria*. The compound has anti-arrhythmic,²⁶¹ antibacterial,²⁶² anti-parkinson,²⁶³ anticancer²⁶⁴ and anti-hypertensive²⁶⁵ activities, among others. It is an approved drug for the management of high blood pressure and has been reported active against SARS-CoVs.¹⁵⁸ It inhibited viral replication with IC₅₀ of 3.4 µM. Six other compounds related to reserpine were also shown to have activities toward SARS-CoV at <100 µM.¹⁵⁸

Yatein

Yatein, (**21**) a lignan was isolated from *Chamaecyparis obtusa* and tested for its activity against herpes simplex virus type 1 (HSV-1). It significantly suppressed HSV-1 multiplication by suppressing the levels of glycoprotein B (gB) and gC mRNA expression, arresting the replication of HSV-1 DNA and decreasing ICP0 and ICP4 gene expression.²⁶⁶ Lignans are distributed widely in the plant kingdom and more than 200 classical lignans and 100 neolignans with vast structural diversity have been identified till date.²⁶⁷ Yatein is a dibenzylbutyrolactone that inhibited HSV-1 alpha gene expression by arresting HSV-1 DNA synthesis and structural protein expression in HeLa cells.²⁶⁸ This compound and its derivatives can be assessed for effectiveness against coronaviruses.

Others

Other natural products with reported antiviral activity include sorbifolin (**22**) and pedalitin (**23**), two flavonoids from *Pterogyne nitens* proven effective against Hepatitis C virus (HCV).²⁶⁹ Herbacetin (**24**), rhoifolin and pectolinarin blocked the enzymatic activity of SARS-CoV 3CLpro.²⁷⁰ Herbacetin, isobavachalcone (**25**), quercetin 3-β-d-glucoside, and helichrysetin (**26**) blocked the enzymatic activity of MERS-CoV/3CLpro.²⁷¹ Robustaflavone (**27**) inhibited influenza A and influenza B viruses with EC₅₀ values of 2.0 µg/mL and 0.2 µg/mL, respectively.⁷¹ Ferruginol, dehydroabieta-7-one, suginol, 8β-hydroxyabieta-9(11),13-dien-12-one, 6,7-dehydroroyleanone, pinusolidic acid, α-cadinol, hinokinin, and savinin purified from the ethyl acetate extracts of *Chamaecyparis obtusa*, 3β,12-diacetoxyabieta-6,8,11,13-tetraene, cedrane-3β,12-diol, and betulonic acid isolated from the ethyl acetate extracts of *Juniperus formosana* along with cryptojaponol and 7β-hydroxydeoxycryptojaponol isolated from *Cryptomeria japonica* were evaluated for activity against SARS-CoV. All the compounds inhibited SAR-CoV at concentrations between 3.3 and 10 µM.²⁷² Gomisin M₁ is a ligand from *Schisandra rubriflora* that had potent anti-HIV activity with EC₅₀ value of <0.65 µM.²⁷³ Rubrifloralignan A, from the fruits of *Schisandra rebriflora* is an anti-HIV-1 product with effect on early stage of HIV-1 replication.²⁷⁴ 1α-hydroxybrussonol, a diterpenoid from *Perovskia atriplicifolia* suppressed the replication of hepatitis B virus DNA with selectivity index of 137.7.²⁷⁵ The anti-influenza virus activity of six stilbenoids from *Gnetum pendulum* showed that isorhapontigenin, gnetupendin B, shegansu B, and gnetin D had significant anti-influenza virus activity in MDCK cells, with IC₅₀ values ranging from 0.67 to 11.99 µg/mL when compared to the positive controls oseltamivir and ribavirin with IC₅₀ values of 0.040 and 5.54 µg/mL, respectively.²⁷⁶ Tellimagrandin I, a tannin from *Cornus canadensis* was found effective against herpes simplex virus type 1 (HSV-1) with an EC₅₀ of 2.6 µM for the direct mode and 5.0 µM for the absorption mode of the plaque reduction assay.²⁷⁷ There are several literatures documenting antiviral agents from other natural sources such as those from mushrooms,^{278, 279} and from marine natural products.²⁷⁵⁻²⁸³

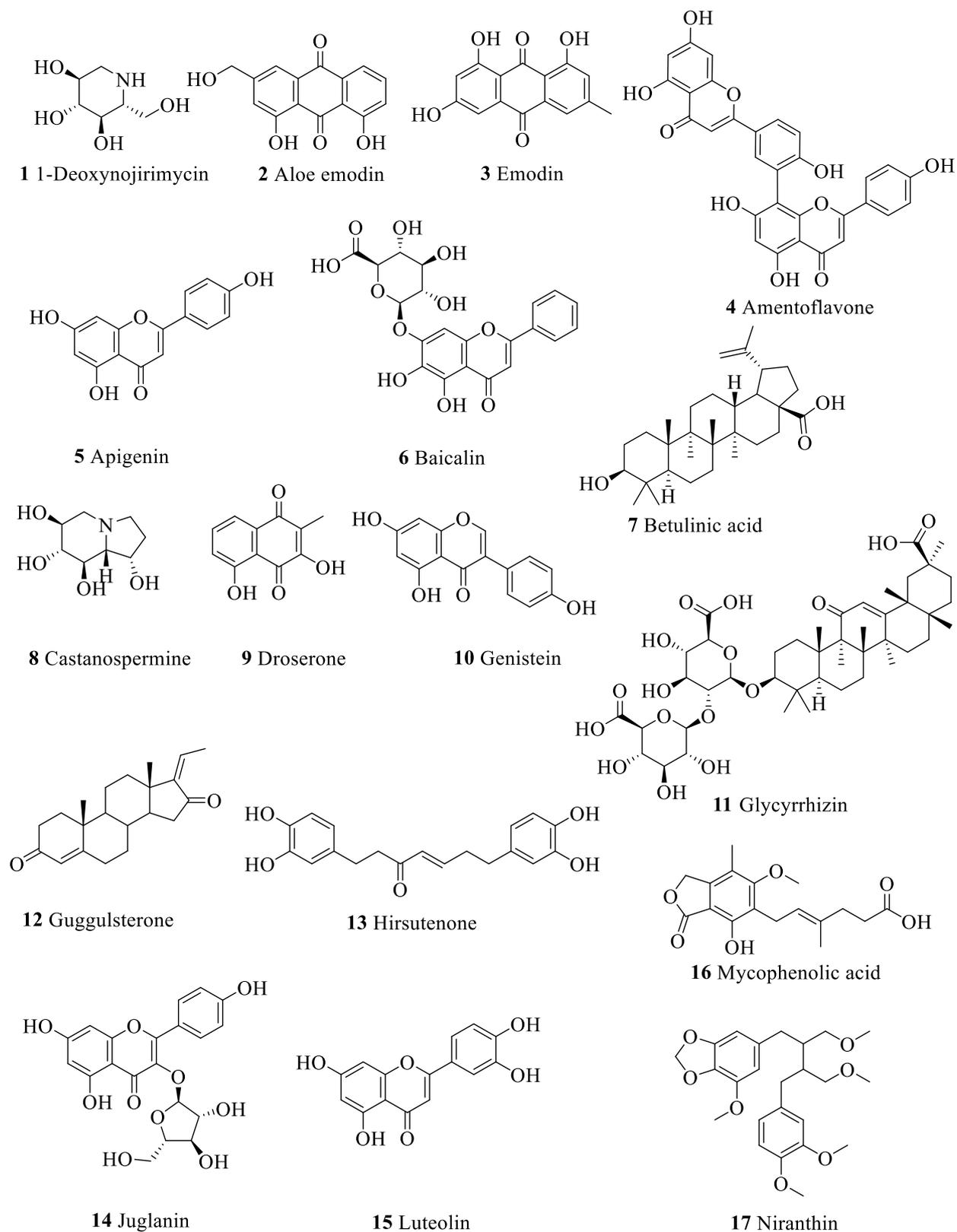


Figure 3: Chemical structures of some promising lead compounds

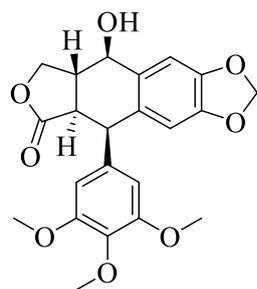
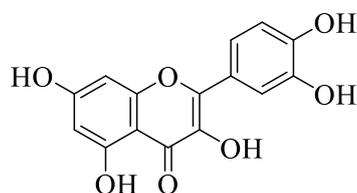
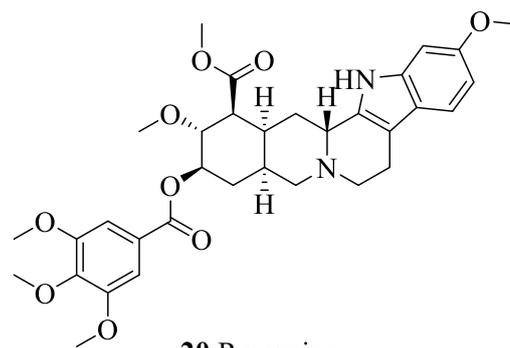
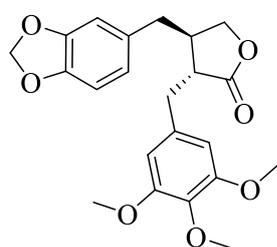
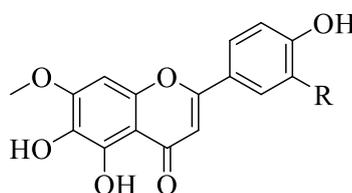
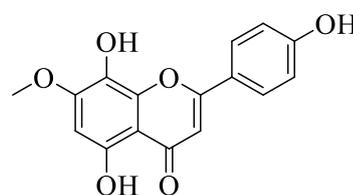
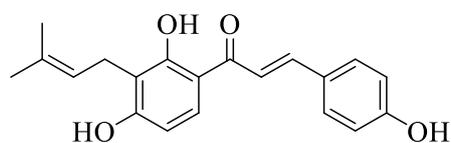
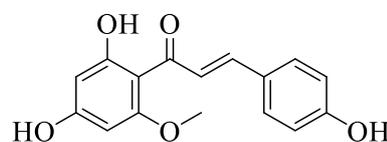
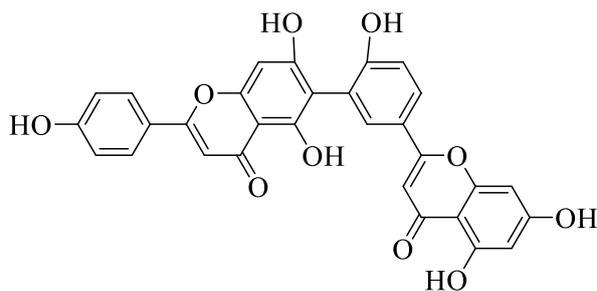
**18** Podophyllotoxin**19** Quercetin**20** Reserpine**21** Yatein**22** R=H Sorbifolin**23** R=OH Padalitin**24** Herbacetin**25** Isobavachalcone**26** Helichrysetin**27** Robustaflavone**Figure 3 Cont'd:** Chemical structures of some promising lead compounds

Table 1: Selected natural compounds and their antiviral mechanisms of action

Compounds	Class	Source	Virus	Mechanisms of action	Dose/IC ₅₀	References
1-Deoxynojirimycin	Alkaloid	<i>Commelina communis</i>	Hepatitis	Inhibited glycosylation of viral envelope proteins	9.92 µM.	55
Aloe-emodin	Anthraquinone	<i>Aloe vera</i>	Influenza A virus	Up-regulated galectin-3 and thioredoxin	0.05 µg/mL	64
			Enterovirus 71 (EV71)	Induced interferon (IFN)	0.14 µg/mL to 0.52 µg/mL	65
			Japanese encephalitis virus (JEV)	Induced interferon (IFN)	0.50 µg/mL to 1.51 µg/mL	65
Amentoflavone	Biflavone	<i>Torreya nucifera</i>	SARS-CoV	3CL ^{pro} inhibition	8.3 µM	66
			Herpes viruses (HSV-1, HSV-2)		17.9 and 48.0 mg/mL	71
			Human immunodeficiency virus (HIV)	Inhibited HIV-1 RT	119 µM	68
			Respiratory syncytial virus (RSV)		5.5 µg/mL	69
			Dengue virus	Inhibited dengue virus NS5 RNA-dependent RNA polymerase	1.3 µM	70
Apigenin	Flavonoids	<i>Arisaema tortuosum</i>	Herpes virus HSV-2	Inhibited both early and late events of the HSV-2 replication	0.05 µg/mL	77
			African swine fever virus (ASFV)	Inhibited ASFV-specific protein synthesis and viral factory formation		76
			Hepatitis C virus (HCV)	Inhibited maturation of miRNAs and HCV replication Decreased the expression levels of mature miR122		74
			Foot-and-mouth disease virus(FMD)	Inhibited FMDV-mediated cytopathogenic effect and FMDV replication	8.593 µg/mL	75
			Enterovirus-71 (EV71)	Disrupted viral RNA association with hnRNP A1 and A2 proteins	10.3 µM	73
Baicalin	Flavone glycoside	<i>Scutellaria baicalensis</i>	Dengue virus (DENV)	Inhibited virus replication, extracellular particles and showed anti-adsorption effect	13.5 ± 0.08, 8.74 ± 0.08 and 18.07 ± 0.2 µg/mL respectively	85
			Influenza virus	Inhibited neuraminidases	43.3 µg/mL	86

			HIV-1	Inhibited both T cell tropic (X4) and monocyte tropic (R5) HIV-1 Env protein mediated fusion Inhibited the activity of HIV-1 reverse transcriptase	0.5 µg/mL	87,88
			Enterovirus 71 (EV71)	Inhibited EV71/3D polymerase expression and Fas/FasL signaling pathways	4.96 µg/mL	89
			Chikungunya virus (CHIKV)	Inhibited viral entry, viral particle attachment, and replicase complexes formation	7 µM,	90
			SARS-CoV		12.5 to 25 µg/mL	91
Betulinic acid	Triterpenoid	<i>Betula pubescens</i>	<i>Herpes simplex</i> virus	Reduced viral cytopathic effect	30 µg/mL	101
			HIV-1	Inhibited HIV fusion and interfere with a specific step in HIV-1 maturation		99
Castanospermine	Alkaloid	<i>Castanospermum austral</i>	HIV	Modified glycoprotein		105
			Dengue virus	Inhibited infectivity of viral particles		107
			Influenza	Inhibited glucosidase 1		107
			Influenza	Inhibited glycoprotein		56
			HIV	Inhibited virus entry at the Env/coreceptor interaction step		57
Chebulagic acid	Tannin	<i>Terminalia chebula</i>	Herpes simplex virus	Inactivated HSV-1 viral particles	17.02±2.82 µM	114
			Human enterovirus 71	Inhibition of viral replication	12.5 µg/mL	115
			HIV	Inhibited HIV reverse transcriptase		116
			Influenza	Inhibited neuraminidase		117
Cyanovirin-N	Protein	<i>Nostoc ellipsosporium</i>	HIV-1	Blocked binding of gp120 to cell-associated CD4		122
			Hepatitis C	Binds to viral envelope glycoproteins and blocked the interaction between the envelope protein E2 and CD81	1.6 ± 0.1 nM	125
			Influenza	Bound viral hemagglutinin	0.004 to 0.04 µg/mL	126
			Ebola virus	Binds to the viral surface glycoprotein		127

Cyclosporine A	Peptide	<i>Hypocladium inflatum gams</i>	HIV-1	Disrupted Gag–cyclophilin A interaction		133
			HIV-1	Blocked incorporation of HIV-1 envelope glycoprotein into virions		134
			Hepatitis C virus (HCV)	Cleavage at NS2/NS3 junction	0.15 g/mL	137
			Hepatitis B	Inhibited viral replication by blocking cytosolic calcium signaling		138
			Influenza virus	Inhibited the replication through CypA-dependent pathway		139
			Coronavirus HCoV-NL63	Knockdown of cellular Cyclophilin A (CypA/PPIA) gene	0.9-2.0 µM	140
Droserone	Naphthoquinone	<i>Drosera whittakeri</i>	Measles virus	Reduced viral entry	2µM	146
Emodin	Anthraquinone	<i>Rheum Palmatum</i>	SARS-CoV	Inhibited ACE2 interaction		59
			Herpes simplex virus (HSV)	Inhibited nuclease activity of HSV-1 UL12		62
Escin	Triterpene saponin	<i>Aesculus hippocastanum</i>	HIV-1	Anti-HIV-1 protease	35 µM	157
			Porcine epidemic diarrhea virus (PEDV)	Inhibited nucleocapsid protein synthesis and viral replication	20 µM	156
			SARS-CoV	Inhibited viral replication	6.0 µM	158
<i>Galanthus nivalis</i> agglutinin (GNA)	Lectins	<i>Galanthus nivalis</i>	Mouse hepatitis virus	Inhibition of viral fusion	50 mg/L	284
Genistein	Isoflavones	<i>Genista tinctoria</i>	African swine fever virus	Disrupted viral DNA synthesis		164
			Herpes B virus	Prevented plaque formation of Herpes B virus and reduced virus production	33 and 46 µM	165
Geraniin	Dehydroellagitannin	<i>Geranium carolinianum</i>	Hepatitis B virus (HBV)	Inhibited HBsAg and HBeAg		174
			HSV-1 and HSV-2	suppressed both HSV-1 and HSV-2	35.0 µM and 18.4 µM	173
			HIV-1	Inhibited virus uptake and HIV-1 reverse transcriptase	0.24 µM	172
Glycyrrhizin	Triterpenoid saponin	<i>Glycyrrhiza glabra</i>	Varicella-zoster virus	Inhibited replication	0.71 mM	177

			Hepatitis virus (HCV)	Suppressed of viral entry and replication, Inhibited the release of infectious HCV particles due to its inhibitory effect on phospholipase A2 group 1B (PLA2G1B)	16.5 μ M	179
			Coronavirus	Inhibited virus adsorption, penetration and replication	300 mg/L	180
Gomisin M ₁	Ligand	<i>Schisandra rubriflora</i>	HIV		<0.65 μ M	273
<i>Hippeastrum</i> hybrid agglutinin (HHA)	Lectins	<i>Hippeastrum hybrid</i>		Inhibition of viral fusion	50 mg/L	284
Hirsutenone	Diarylheptanoids	<i>Alnus japonica</i>	SARS-CoV.	Inhibited papain-like protease of SARS-CoV.	4.1 μ M	197
Juglanin	Flavonol	<i>Juglans mandshurica</i>	SARS coronavirus	Blocked 3a channel	2.3 μ M	206
Labyrinthopeptin A1	Peptide	<i>Actinomadura namibiensis</i>	HIV	Inhibited viral cell-cell transmission	0.70–3.3 μ M	209
			Herpes	Inhibited viral cell-cell transmission	0.29–2.8 μ M	209
			Human respiratory syncytial virus (hRSV)	Inhibited cell entry		285
Luteolin	Flavonoids	<i>Arisaema tortuosum</i>	Herpes virus HSV-2	Inhibited both early and late events of the HSV-2 replication	0.41 μ g/mL	77
			HBV	Reduced HBV DNA replication and inhibited hepatocyte nuclear factor 4 α (HNF4 α) expression		222
Mucroporin-M1	Peptide	<i>Lychas mucronatus</i>	Measles virus	Inhibited viral infectivity	3.52 μ M	230
			SARS-CoV	Inhibited viral infectivity	7.12 μ M	230
			Influenza A (H5N1)	Inhibited viral infectivity	1.03 μ M	230
			Hepatitis B virus	Activated MAPK pathway and down-regulated HNF4 α		231
Mycophenolic acid	Phthalides	<i>Penicillium stoloniferum</i> <i>Fungi</i>	Dengue virus	Prevented the synthesis and accumulation of viral RNA	0.3 μ M	237
			Hepatitis E virus	Nucleotide depletion		239
			HIV	Depleted guanosine nucleotides		240
			Japanese encephalitis		3.1 g/ml	242
			MERS-CoV	Inhibited papain-like protease (PL ^{pro})	222.5–	244

Niranthin	Lignan	<i>Phyllanthus niruri</i>	Hepatitis B virus	Decreased the secretion of HBsAg and HBeAg	247.6 μ M 15.6 μ M and 25.1 μ M	247
Quercetin	Flavonoid	Many plant species	Herpes simplex virus type 1 (HSV-1), polio-virus type 1, parainfluenza virus type 3 (Pf-3), and respiratory syncytial virus (RSV)	Reduced infectivity and intracellular replication		259
			HIV-1	inhibited HIV-infection by preventing binding of gp120 to CD4	20 μ g/mL	260
Reserpine	Indole alkaloid	<i>Rauwolfia serpentine</i>	SARS-CoVs	Inhibited viral replication	3.4 μ M	158
Rubrifloralignan A	Ligand	<i>Schisandra rebriflora</i>	HIV	Inhibited early stage of HIV-1 replication		274
<i>Urtica dioica</i> agglutinin (UDA)	Lectins	<i>Urtica dioica</i>		Inhibition of viral fusion	50 mg/L	284
Yatein,	Lignan	<i>Chamaecyparis obtuse</i>	Herpes simplex virus type 1 (HSV-1).	Inhibited HSV-1 α gene expression	30.6 \pm 5.5 μ M	266
			EV71	Viral inhibition	10 μ g/mL	175

Concluding remarks

The cellular components of the host are used by corona viruses for different physiological processes in their life cycle. These include viral entry, genomic replication and the assembly and budding of virions resulting in considerable pathological damages to the host cells. Thus, any chemical compound that can interrupt any stage of the viral lifecycle would offer potential therapeutic option for the development of antiviral therapies. In this review, we have reported many compounds of natural origin that have demonstrated *in vitro* or *in vivo* potential against other viruses that are similar to SARS-CoV-2. Our hope is to promote the continuing research of diverse molecules with antiviral potentials. All the discussed compounds could be exploited either directly as antiviral agents or form templates for such. Furthermore, structurally related compounds could also be tested for their antiviral potentials. These compounds could also be optimized for drug discovery and development as they can potentially assist in the rational design of novel antiviral therapeutics. Modes of efficient delivery and safety profiles in humans could be further investigated.

Conflict of interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

References

1. Su S, Wong G, Shi W, Liu J, Lai AC, Zhou J, Liu W, Bi Y, Gao GF. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. *Trends Microbiol.* 2016; 24:490-502.
2. Jaimes JA and Whittaker GR. Feline coronavirus: insights into viral pathogenesis based on the spike protein structure and function. *Virology*. 2018; 517:108-121.
3. Pillaiyar T, Meenakshisundaram S, Manickam M. Recent discovery and development of inhibitors targeting coronaviruses. *Drug Discov Today*. 2020; 25(4):668-688.
4. Fehr AR and Perlman S. Coronaviruses: an overview of their replication and pathogenesis. In *Coronaviruses*, Humana Press, New York, NY. 2015. 1-23 p.
5. Saif LJ, Wang Q, Vlasova AN, Jung K, Xiao S. Coronaviruses. *Dis Swine*. 2019; 3:488-523.
6. Gorbalenya AE. Severe acute respiratory syndrome-related coronavirus—The species and its viruses, a statement of the Coronavirus Study Group. *BioRxiv*. 2020. <https://doi.org/10.1101/2020.02.07.93786> (Accessed May 11, 2020).

7. Paules CI, Marston HD, Fauci AS. Coronavirus infections—more than just the common cold. *JAMA*. 2020; 323(8):707-708.
8. Gorse GJ, O'Connor TZ, Hall SL, Vitale JN, Nichol KL. Human coronavirus and acute respiratory illness in older adults with chronic obstructive pulmonary disease. *J Infect Dis*. 2009; 199:847-857.
9. Walsh EE, Shin JH, Falsey AR. Clinical impact of human coronaviruses 229E and OC43 infection in diverse adult populations. *J Infect Dis*. 2013; 208:1634-1642.
10. Talbot PJ, Jacomy H, Desforges M. Pathogenesis of human coronaviruses other than severe acute respiratory syndrome coronavirus. In *Nidoviruses*. Am Soc Microbiol. 2008. 313-324 p.
11. Desforges M, Le Coupanec A, Stodola JK, Meessen-Pinard M, Talbot PJ. Human coronaviruses: viral and cellular factors involved in neuroinvasiveness and neuropathogenesis. *Virus Res*. 2014; 194:145-158.
12. Wu Y, Xu X, Chen Z, Duan J, Hashimoto K, Yang L, Liu C, Yang C. Nervous system involvement after infection with COVID-19 and other coronaviruses. *Brain Behav Immun*. 2020; 87:18-22.
13. Ji HF, Li XJ, Zhang HY. Natural products and drug discovery. *EMBO Rep*. 2009; 10:194-200.
14. Harvey AL. Natural products in drug discovery. *Drug Discov Today*. 2008; 13(19-20):894-901.
15. Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981– 2002. *J Nat Prod*. 2003; 66:1022-1037.
16. Lam KS. New aspects of natural products in drug discovery. *Trends Microbiol*. 2007; 15:279-289.
17. Lahlou M. The success of natural products in drug discovery. *Pharmacol Pharm*. 2013; 4:17-31.
18. Harvey AL, Edrada-Ebel R, Quinn RJ. The re-emergence of natural products for drug discovery in the genomics era. *Nat Rev Drug Discov*. 2015; 14:111-129.
19. Rodrigues T, Reker D, Schneider P, Schneider G. Counting on natural products for drug design. *Nat Chem*. 2016; 8:531.
20. Thomford NE, Senthelane DA, Rowe A, Munro D, Seele P, Maroyi A, Dzobo K. Natural products for drug discovery in the 21st century: innovations for novel drug discovery. *Int J Mol Sci*. 2018; 19:1578.
21. Avato P. Editorial to the Special Issue–Natural Products and Drug Discovery. *Mol*. 2020, 25:1128.
22. Martinez JP, Sasse F, Brönstrup M, Diez J, Meyerhans A. Antiviral drug discovery: broad-spectrum drugs from nature. *Nat Prod Rep*. 2015; 32:29-48.
23. Chen Y, Liu Q, Guo D. Emerging coronaviruses: genome structure, replication, and pathogenesis. *J Med Virol*. 2020; 92:418-423.
24. Snijder EJ, Van Der Meer Y, Zevenhoven-Dobbe J, Onderwater JJ, van der Meulen J, Koerten HK, Mommaas AM. Ultrastructure and origin of membrane vesicles associated with the severe acute respiratory syndrome coronavirus replication complex. *J Virol*. 2006; 80:5927-5940.
25. Rasool A, Ishfaq S, Uqab B. Novel Coronavirus (2019-nCoV) Outbreak in China: From Local Epidemics to Global Pandemics. Available at SSRN 3559461. 2020. https://papers.ssrn.com/sol3/papers.cfm?abstract_id=3559461
26. Sola I, Almazan F, Zuniga S, Enjuanes L. Continuous and discontinuous RNA synthesis in coronaviruses. *Ann Rev Virol*. 2015; 2:265-288.
27. Enjuanes L, Sola I, Almazan F, Ortego J, Izeta A, Gonzalez JM, Alonso S, Sanchez JM, Escors D, Calvo E, Riquelme C. Coronavirus derived expression systems. *J Biotech*. 2001; 88:183-204.
28. Masters PS. The molecular biology of coronaviruses. *Adv Virus Res*. 2006; 66:193-292.
29. Lei J and Hilgenfeld R. RNA-virus proteases counteracting host innate immunity. *FEBS Lett*. 2017; 591:3190-3210.
30. Kamitani W, Huang C, Narayanan K, Lokugamage KG, Makino S. A two-pronged strategy to suppress host protein synthesis by SARS coronavirus Nsp1 protein. *Nat Struct Mol Biol*. 2009; 16:1134.
31. Yeager CL, Ashmun RA, Williams RK, Cardellicchio CB, Shapiro LH, Look AT, Holmes KV. Human aminopeptidase N is a receptor for human coronavirus 229E. *Nature* 1992; 357:420-422.
32. Huang X, Dong W, Milewska A, Golda A, Qi Y, Zhu QK, Marasco WA, Baric RS, Sims AC, Pyrc K, Li W. Human coronavirus HKU1 spike protein uses O-acetylated sialic acid as an attachment receptor determinant and employs hemagglutinin-esterase protein as a receptor-destroying enzyme. *J Virol*. 2015; 89:7202-7213.
33. Butler N, Pewe L, Trandem K, Perlman S. Murine encephalitis caused by HCoV-OC43, a human coronavirus with broad species specificity, is partly immune-mediated. *J Virol*. 2006; 347:410-421.
34. Li W, Moore MJ, Vasilieva N, Sui J, Wong SK, Berne MA, Somasundaran M, Sullivan JL, Luzuriaga K, Greenough TC, Choe H. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature* 2003; 426:450-454.
35. Li W, Sui J, Huang IC, Kuhn JH, Radoshitzky SR, Marasco WA, Choe H, Farzan M. The S proteins of human coronavirus NL63 and severe acute respiratory syndrome coronavirus bind overlapping regions of ACE2. *J Virol*. 2007; 367:367-374.
36. Wu K, Li W, Peng G, Li F. Crystal structure of NL63 respiratory coronavirus receptor-binding domain complexed with its human receptor. *P Nat Acad Sci*. 2009; 106:19970-19974.
37. van Doremalen N, Miazgowiec KL, Milne-Price S, Bushmaker T, Robertson S, Scott D, Kinne J, McLellan JS, Zhu J, Munster VJ. Host species restriction of Middle East respiratory syndrome coronavirus through its receptor, dipeptidyl peptidase 4. *J Virol*. 2014; 88:9220-9232.
38. Bertram S, Glowacka I, Müller MA, Lavender H, Gnirss K, Nehlmeier I, Niemeyer D, He Y, Simmons G, Drosten C, Soilleux EJ. Cleavage and activation of the severe acute respiratory syndrome coronavirus spike protein by human airway trypsin-like protease. *J Virol*. 2011; 85:13363-13372.
39. Bertram S, Dijkman R, Habjan M, Heurich A, Gierer S, Glowacka I, Welsch K, Winkler M, Schneider H, Hofmann-Winkler H, Thiel V. TMPRSS2 activates the human coronavirus 229E for cathepsin-independent host cell entry and is expressed in viral target cells in the respiratory epithelium. *J Virol*. 2013; 87:6150-6160.
40. Cheng VC, Lau SK, Woo PC, Yuen KY. Severe acute respiratory syndrome coronavirus as an agent of emerging and reemerging infection. *Clin Microbiol Rev*. 2007; 20:660-694.
41. Bosch BJ, van der Zee R, de Haan CA, Rottier PJ. The coronavirus spike protein is a class I virus fusion protein: structural and functional characterization of the fusion core complex. *J Virol*. 2003; 77:8801-88011.
42. Zhang H, Penninger JM, Li Y, Zhong N, Slutsky AS. Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target. *Intens Care Med*. 2020; 46:586-590.
43. Pyrc K, Berkhout B, van der Hoek L. Antiviral strategies against human coronaviruses. *Infect Disord-Drug Targets (Formerly Current Drug Targets-Infectious Disorders)*. 2007; 7:59-66.
44. Mothay D, Ramesh KV. Binding site analysis of potential protease inhibitors of COVID-19 using AutoDock. *Virus Dis*. 2020; 31:194-199.

45. Bhimraj A, Morgan RL, Shumaker AH, Lavergne V, Baden L, Cheng VC, Edwards KM, Gandhi R, Muller WJ, O'Horo JC, Shoham S. Infectious diseases Society of America guidelines on the treatment and management of patients with COVID-19. *Clin Infect Dis.* 2020. DOI: 10.1093/cid/ciaa478
46. Sharma S, Basu S, Shetti NP, Aminabhavi TM. Current treatment protocol for COVID-19 in India. *Sensors Int.* 2020; 1: 1-3 100013.
47. Chen ZR, Zhou Y, Liu J, Peng HW, Zhou J, Zhong HL, Liu LL, Lai MF, Wei XH, Wen JH. The Pharmacotherapies Advice of Guidelines for COVID-19. *Front Pharmacol.* 2020; 11:950.
48. Barlow A, Landolf KM, Barlow B, Yeung SY, Heavner JJ, Claassen CW, Heavner MS. Review of emerging pharmacotherapy for the treatment of coronavirus disease 2019. *Pharmacotherapy: J Hum Pharmacol Drug Ther.* 2020; 40:416-437.
49. Guo YR, Cao QD, Hong ZS, Tan YY, Chen SD, Jin HJ, Tan KS, Wang DY, Yan Y. The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak—an update on the status. *Mil Med Res.* 2020; 7:1-10.
50. Salvi R, Patankar P. Emerging pharmacotherapies for COVID-19. *Biomed Pharmacother.* 2020; 128: 110267.
51. Richards G, Mer M, Schleicher G, Stacey S. COVID-19 and the rationale for pharmacotherapy: A South African perspective. *Wits J Clin Med* 2020; 2(SI):11.
52. Kang KD, Cho YS, Song JH, Park YS, Lee JY, Hwang KY, Rhee SK, Chung JH, Kwon O, Seong SI. Identification of the genes involved in 1-deoxynojirimycin synthesis in *Bacillus subtilis* MORI 3K-85. *J Microbiol.* 2011; 49:431-440.
53. Gomollón-Bel F, Delso I, Tejero T, Merino P. Biosynthetic pathways to glycosidase inhibitors. *Curr Chem Bio.* 2014; 8:10-16.
54. Gao K, Zheng C, Wang T, Zhao H, Wang J, Wang Z, Zhai X, Jia Z, Chen J, Zhou Y, Wang W. 1-deoxynojirimycin: occurrence, extraction, chemistry, oral pharmacokinetics, biological activities and *in silico* target fishing. *Mol.* 2016; 21:1600.
55. Jacob JR, Mansfield K, You JE, Tennant BC, Kim YH. Natural iminosugar derivatives of 1-deoxynojirimycin inhibit glycosylation of hepatitis viral envelope proteins. *J Microbiol.* 2007; 45:431-440.
56. Tanaka Y, Kato J, Kohara M, Galinski MS. Antiviral effects of glycosylation and glucose trimming inhibitors on human parainfluenza virus type 3. *Antiviral Res.* 2006; 72:1-9.
57. Papandréou MJ, Barbouche R, Guieu R, Kiény MP, Fenouillet E. The α -glucosidase inhibitor 1-deoxynojirimycin blocks human immunodeficiency virus envelope glycoprotein-mediated membrane fusion at the CXCR4 binding step. *Mol Pharmacol.* 2002; 61:186-193.
58. Romero PA, Datema R, Schwarz RT. N-methyl-1-deoxynojirimycin, a novel inhibitor of glycoprotein processing, and its effect on fowl plague virus maturation. *Viol.* 1983; 130:238-242.
59. Ho TY, Wu SL, Chen JC, Li CC, Hsiang CY. Emodin blocks the SARS coronavirus spike protein and angiotensin-converting enzyme 2 interaction. *Antiviral Res.* 2007; 74:92-101.
60. Yang YC, Lim MY, Lee HS. Emodin isolated from *Cassia obtusifolia* (Leguminosae) seed shows larvicidal activity against three mosquito species. *J Agric Food Chem.* 2003; 51:7629-7631.
61. Chun-Guang W, Jun-Qing Y, Bei-Zhong L, Dan-Ting J, Chong W, Liang Z, Dan Z, Yan W. Anti-tumor activity of emodin against human chronic myelocytic leukemia K562 cell lines *in vitro* and *in vivo*. *Eur J Pharmacol.* 2010; 627:33-41.
62. Hsiang CY and Ho TY. Emodin is a novel alkaline nuclease inhibitor that suppresses herpes simplex virus type 1 yields in cell cultures. *Brit J Pharmacol.* 2008; 155:227-235.
63. Xiong HR, Luo J, Hou W, Xiao H, Yang ZQ. The effect of emodin, an anthraquinone derivative extracted from the roots of *Rheum tanguticum*, against herpes simplex virus *in vitro* and *in vivo*. *J Ethnopharmacol.* 2011; 133:718-723.
64. Li SW, Yang TC, Lai CC, Huang SH, Liao JM, Wan L, Lin YJ, Lin CW. Antiviral activity of aloe-emodin against influenza A virus via galectin-3 up-regulation. *Eur J Pharmacol.* 2014; 738:125-132.
65. Lin CW, Wu CF, Hsiao NW, Chang CY, Li SW, Wan L, Lin YJ, Lin WY. Aloe-emodin is an interferon-inducing agent with antiviral activity against Japanese encephalitis virus and enterovirus 71. *Int J Antimicrob Agents* 2008; 32:355-359.
66. Ryu YB, Jeong HJ, Kim JH, Kim YM, Park JY, Kim D, Nguyen TT, Park SJ, Chang JS, Park KH, Rho MC. Biflavonoids from *Torreya nucifera* displaying SARS-CoV 3CLpro inhibition. *Bioorg Med Chem.* 2010; 18:7940-7947.
67. Yu S, Yan H, Zhang L, Shan M, Chen P, Ding A, Li SF. A review on the phytochemistry, pharmacology, and pharmacokinetics of amentoflavone, a naturally-occurring biflavonoid. *Mol.* 2017; 22:299.
68. Lin YM, Anderson H, Flavin MT, Pai YH, Mata-Greenwood E, Pengsuparp T, Pezzuto JM, Schinazi RF, Hughes SH, Chen FC. In vitro anti-HIV activity of biflavonoids isolated from *Rhus succedanea* and *Garcinia multiflora*. *J Nat Prod.* 1997; 60:884-888.
69. But PP, Ooi VE, He YH, Lee SH, Lee SF, Lin RC. Antiviral amentoflavone from *Selaginella sinensis*. *Bio Pharma Bull.* 2001; 24:311-312.
70. Coulerie P, Nour M, Maciuk A, Eydoux C, Guillemot JC, Lebouvier N, Hnawia E, Leblanc K, Lewin G, Canard B, Figadère B. Structure-activity relationship study of biflavonoids on the Dengue virus polymerase DENV-NS5 RdRp. *Planta Med.* 2013; 79:1313-1318.
71. Lin YM, Flavin MT, Schure R, Chen FC, Sidwell R, Barnard DI, Huffmann JH, Kern ER. Antiviral activities of biflavonoids. *Planta Med.* 1999; 65:120-125.
72. Salehi B, Venditti A, Sharifi-Rad M, Kęrgiel D, Sharifi-Rad J, Durazzo A, Lucarini M, Santini A, Souto EB, Novellino E, Antolak H. The therapeutic potential of apigenin. *Int J Mol Sci.* 2019; 20:1305.
73. Zhang W, Qiao H, Lv Y, Wang J, Chen X, Hou Y, Tan R, Li E. Apigenin inhibits enterovirus-71 infection by disrupting viral RNA association with trans-acting factors. *PloS one.* 2014; 9(10):e110429.
74. Shibata C, Ohno M, Otsuka M, Kishikawa T, Goto K, Muroyama R, Kato N, Yoshikawa T, Takata A, Koike K. The flavonoid apigenin inhibits hepatitis C virus replication by decreasing mature microRNA122 levels. *Viol.* 2014; 462:42-48.
75. Qian S, Fan W, Qian P, Zhang D, Wei Y, Chen H, Li X. Apigenin restricts FMDV infection and inhibits viral IRES driven translational activity. *Viruses.* 2015; 7: 1613-1626.
76. Hakobyan A, Arabyan E, Avetisyan A, Abroyan L, Hakobyan L, Zakaryan H. Apigenin inhibits African swine fever virus infection *in vitro*. *Arch Virol.* 2016; 161:3445-3453.
77. Rittà M, Marengo A, Civra A, Lembo D, Cagliero C, Kant K, Lal UR, Rubiolo P, Ghosh M, Donalisio M. Antiviral Activity of a *Arisaema Tortuosum* Leaf Extract and Some of its Constituents against Herpes Simplex Virus Type 2. *Planta Med.* 2020; 86:267-275.
78. Rosa-Calatrava M, Terrier O, Proust A, Moules V, inventors; Centre National de la Recherche Scientifique CNRS, Université Claude Bernard Lyon 1 (UCBL), Institut National de la Santé et de la Recherche Médicale INSERM, assignee. Antiviral compositions for the treatment of

- infections linked to coronaviruses. United States patent application US 16/340,346. 2019.
79. Pitchiah KM, Sundaram KM, Ramasamy MS. Coronavirus Spike (S) Glycoprotein (2019-Ncov) Targeted Siddha Medicines Kabasura Kudineer and Thonthasura Kudineer–*In silico* Evidence for Corona Viral Drug. *Asian J Pharm Res Health Care*. 2019; 11:1-9.
 80. Mishra A, Pathak Y, Tripathi V. Natural compounds as potential inhibitors of novel coronavirus (COVID-19) main protease: An *in silico* study. *Res Square* 2020. DOI: <https://doi.org/10.21203/rs.3.rs-22839/v2>
 81. Shang X, He X, He X, Li M, Zhang R, Fan P, Zhang Q, Jia Z. The genus *Scutellaria* an ethnopharmacological and phytochemical review. *J Ethnopharmacol*. 2010; 128:279-313.
 82. Grzegorzczak-Karolak I, Wiktoerek-Smagur A, Hnatuszko-Konka K. An untapped resource in the spotlight of medicinal biotechnology: The genus *Scutellaria*. *Curr Pharm Biotech*. 2018; 19:358-371.
 83. Wen M, Li X, Fu ST. New research progress in pharmacological activities of baicalin [J]. *J Shenyang Pharm Uni* 2008; 2:1-14
 84. Liang W, Huang X, Chen W. The effects of baicalin and baicalein on cerebral ischemia: a review. *Aging Dis*. 2017; 8:850.
 85. Moghaddam E, Teoh BT, Sam SS, Lani R, Hassandarvish P, Chik Z, Yueh A, Abubakar S, Zandi K. Baicalin, a metabolite of baicalein with antiviral activity against dengue virus. *Sci Rep*. 2014; 4:5452.
 86. Ding Y, Dou J, Teng Z, Yu J, Wang T, Lu N, Wang H, Zhou C. Antiviral activity of baicalin against influenza A (H1N1/H3N2) virus in cell culture and in mice and its inhibition of neuraminidase. *Arch Virol*. 2014; 159:3269-3278.
 87. Kitamura K, Honda M, Yoshizaki H, Yamamoto S, Nakane H, Fukushima M, Ono K, Tokunaga T. Baicalin, an inhibitor of HIV-1 production *in vitro*. *Antiviral Res*. 1998; 37: 131-140.
 88. Li BQ, Fu T, Dongyan Y, Mikovits JA, Ruscetti FW, Wang JM. Flavonoid baicalin inhibits HIV-1 infection at the level of viral entry. *Biochem Biophys Res Comm*. 2000; 276:534-538.
 89. Li X, Liu Y, Wu T, Jin Y, Cheng J, Wan C, Qian W, Xing F, Shi W. The antiviral effect of baicalin on enterovirus 71 *in vitro*. *Viruses* 2015; 7:4756-4771.
 90. Oo A, Rausalu K, Merits A, Higgs S, Vanlandingham D, Bakar SA, Zandi K. Deciphering the potential of baicalin as an antiviral agent for Chikungunya virus infection. *Antiviral Res*. 2018; 150:101-111.
 91. Chen F, Chan KH, Jiang Y, Kao RY, Lu HT, Fan KW, Cheng VC, Tsui WH, Hung IF, Lee TS, Guan Y. In vitro susceptibility of 10 clinical isolates of SARS coronavirus to selected antiviral compounds. *J Clin Virol*. 2004; 31:69-75.
 92. Liu AL, Wang HD, Lee SM, Wang YT, Du GH. Structure–activity relationship of flavonoids as influenza virus neuraminidase inhibitors and their *in vitro* anti-viral activities. *Bioorg Med Chem*. 2008; 16:7141-7147.
 93. Liu R, Li X, Wei J, Liu S, Chang Y, Zhang J, Zhang J, Zhang X, Fuhr U, Taubert M, Tian X. A single dose of baicalin has no clinically significant effect on the pharmacokinetics of cyclosporine A in healthy chinese volunteers. *Front Pharmacol*. 2019; 10:518.
 94. Tan Y, Yu R, Pezzuto JM. Betulinic acid-induced programmed cell death in human melanoma cells involves mitogen-activated protein kinase activation. *Clin Cancer Res*. 2003; 9:2866-2875.
 95. Fulda S. Betulinic acid for cancer treatment and prevention. *Int J Mol Sci*. 2008; 9:1096-1107.
 96. Chintharlapalli S, Papineni S, Lei P, Pathi S, Safe S. Betulinic acid inhibits colon cancer cell and tumor growth and induces proteasome-dependent and-independent downregulation of specificity proteins (Sp) transcription factors. *BMC cancer*. 2011; 11:371.
 97. Innocente AM, Silva GN, Cruz LN, Moraes MS, Nakabashi M, Sonnet P, Gosmann G, Garcia CR, Gnoatto SC. Synthesis and antiplasmodial activity of betulinic acid and ursolic acid analogues. *Molecules*. 2012; 17:12003-12014.
 98. Tsai JC, Peng WH, Chiu TH, Lai SC, Lee CY. Anti-inflammatory effects of *Scoparia dulcis* L. and betulinic acid. *Am J Chinese Med*. 2011; 39:943-956.
 99. Aiken C, Chen CH. Betulinic acid derivatives as HIV-1 antivirals. *Trends Mol Med*. 2005; 11:31-36.
 100. Pavlova NI, Savinova OV, Nikolaeva SN, Boreko EI, Flekhter OB. Antiviral activity of betulin, betulinic and betulonic acids against some enveloped and non-enveloped viruses. *Fitoter*. 2003; 74:489-492.
 101. Ryu SY, Lee CK, Lee CO, Kim HS, Zee OP. Antiviral triterpenes from *Prunella vulgaris*. *Arch Pharm Res*. 1992; 15:242-245.
 102. Kazakova OB, Gul'nara VG, Yamansarov EY, Tolstikov GA. Betulin and ursolic acid synthetic derivatives as inhibitors of Papilloma virus. *Bioorg Med Chem Lett*. 2010; 20:4088-4090.
 103. Nash RJ, Fellows LE, Dring JV, Stirton CH, Carter D, Hegarty MP, Bell EA. Castanospermine in Alexa species. *Phytochem*. 1988; 27:1403-1404.
 104. Saul R, Chambers JP, Molyneux RJ, Elbein AD. Castanospermine, a tetrahydroxylated alkaloid that inhibits beta-glucosidase and beta-glucocerebrosidase [isolated from the seeds of the Australian legume, *Castanospermum australe*]. *Arch Biochem Biophys*. 1983; 221:593-597.
 105. Walker BD, Kowalski M, Goh WC, Kozarsky K, Krieger M, Rosen C, Rohrschneider L, Haseltine WA, Sodroski J. Inhibition of human immunodeficiency virus syncytium formation and virus replication by castanospermine. *Proc Nat Acad Sci*. 1987; 84:8120-8124.
 106. Whitby K, Pierson TC, Geiss B, Lane K, Engle M, Zhou Y, Doms RW, Diamond MS. Castanospermine, a potent inhibitor of dengue virus infection *in vitro* and *in vivo*. *J Virol*. 2005; 79:8698-8706.
 107. Pan YT, Hori H, Saul R, Sanford BA, Molyneux RJ, Elbein AD. Castanospermine inhibits the processing of the oligosaccharide portion of the influenza viral hemagglutinin. *Biochem*. 1983; 22:3975-3984.
 108. Liu PS, Sunkara SP, Bowlin TL, inventors; Aventis Inc, assignee. Anti-retroviral castanospermine esters. United States patent US 5,004,746. 1991.
 109. Chen PS, Li JH. Chemopreventive effect of punicalagin, a novel tannin component isolated from *Terminalia catappa*, on H-ras-transformed NIH3T3 cells. *Toxicol Lett*. 2006; 163:44-53.
 110. Hamada SI, Kataoka T, Woo JT, Yamada A, Yoshida T, Nishimura T, Otake N, Nagai K. Immunosuppressive effects of gallic acid and chebulagic acid on CTL-mediated cytotoxicity. *Bio Pharm Bull*. 1997; 20:1017-1019.
 111. Kinoshita S, Inoue Y, Nakama S, Ichiba T, Aniya Y. Antioxidant and hepatoprotective actions of medicinal herb, *Terminalia catappa* L. from Okinawa Island and its tannin corilagin. *Phytomed*. 2007; 14:755-762.
 112. Sasidharan I, Sundaresan A, Nisha VM, Kirishna MS, Raghu KG, Jayamurthy P. Inhibitory effect of *Terminalia chebula* Retz. fruit extracts on digestive enzyme related to diabetes and oxidative stress. *J Enz Inhib Med Chem*. 2012; 27:578-586.
 113. Pham AT, Malterud KE, Paulsen BS, Diallo D, Wangenstein H. α -Glucosidase inhibition, 15-lipoxygenase inhibition, and brine shrimp toxicity of extracts and isolated compounds from *Terminalia macroptera* leaves. *Pharm Bio*. 2014; 52:1166-1169.
 114. Lin LT, Chen TY, Chung CY, Noyce RS, Grindley TB, McCormick C, Lin TC, Wang GH, Lin CC, Richardson CD.

- Hydrolyzable tannins (chebulagic acid and punicalagin) target viral glycoprotein-glycosaminoglycan interactions to inhibit herpes simplex virus 1 entry and cell-to-cell spread. *J Virol.* 2011; 85:4386-4398.
115. Yang Y, Xiu J, Liu J, Zhang L, Li X, Xu Y, Qin C, Zhang L. Chebulagic acid, a hydrolyzable tannin, exhibited antiviral activity *in vitro* and *in vivo* against human enterovirus 71. *Int J Mol Sci.* 2013; 14:9618-9627.
 116. Nonaka GI, Nishioka I, Nishizawa M, Yamagishi T, Kashiwada Y, Dutschman GE, Bodner AJ, Kilkuskie RE, Cheng YC, Lee KH. Anti-AIDS agents, 2: inhibitory effect of tannins on HIV reverse transcriptase and HIV replication in H9 lymphocyte cells. *J Nat Prod.* 1990; 53:587-595.
 117. Li P, Du R, Wang Y, Hou X, Wang L, Zhao X, Zhan P, Liu X, Rong L, Cui Q. Identification of Chebulinic Acid and Chebulagic Acid as Novel Influenza Viral Neuraminidase Inhibitors. *Front Microbiol.* 2020; 11:182.
 118. Lin LT, Chen TY, Lin SC, Chung CY, Lin TC, Wang GH, Anderson R, Lin CC, Richardson CD. Broad-spectrum antiviral activity of chebulagic acid and punicalagin against viruses that use glycosaminoglycans for entry. *BMC Microbiol.* 2013; 13:187.
 119. Kesharwani A, Polachira SK, Nair R, Agarwal A, Mishra NN, Gupta SK. Anti-HSV-2 activity of *Terminalia chebula* Retz extract and its constituents, chebulagic and chebulinic acids. *BMC Compl Altern Med.* 2017; 17:110.
 120. Dittmann E, Neilan B, Börner T. Molecular biology of peptide and polyketide biosynthesis in cyanobacteria. *App Microbiol Biotech.* 2001; 57:467-473.
 121. Bewley CA, Gustafson KR, Boyd MR, Covell DG, Bax A, Clore GM, Gronenborn AM. Solution structure of cyanovirin-N, a potent HIV-inactivating protein. *Nat Stru Bio.* 1998; 5:571-578.
 122. Dey B, Lerner DL, Lusso P, Boyd MR, Elder JH, Berger EA. Multiple antiviral activities of cyanovirin-N: blocking of human immunodeficiency virus type 1 gp120 interaction with CD4 and coreceptor and inhibition of diverse enveloped viruses. *J Virol.* 2000; 74:4562-4569.
 123. Tsai CC, Emau P, Jiang Y, Agy MB, Shattock RJ, Schmidt A, Morton WR, Gustafson KR, Boyd MR. Cyanovirin-N inhibits AIDS virus infections in vaginal transmission models. *AIDS Res Hum Retroviruses.* 2004; 20:11-18.
 124. Bolmstedt AJ, O'Keefe BR, Shenoy SR, McMahon JB, Boyd MR. Cyanovirin-N defines a new class of antiviral agent targeting N-linked, high-mannose glycans in an oligosaccharide-specific manner. *Mol Pharmacol.* 2001; 59:949-954.
 125. Helle F, Wychowski C, Vu-Dac N, Gustafson KR, Voisset C, Dubuisson J. Cyanovirin-N inhibits hepatitis C virus entry by binding to envelope protein glycans. *J Bio Chem.* 2006; 281:25177-25183.
 126. O'Keefe BR, Smee DF, Turpin JA, Saucedo CJ, Gustafson KR, Mori T, Blakeslee D, Buckheit R, Boyd MR. Potent anti-influenza activity of cyanovirin-N and interactions with viral hemagglutinin. *Antimicrob Agents Chemother.* 2003; 47:2518-2525.
 127. Barrientos LG, O'Keefe BR, Bray M, Sanchez A, Gronenborn AM, Boyd MR. Cyanovirin-N binds to the viral surface glycoprotein, GP1, 2 and inhibits infectivity of Ebola virus. *Antiviral Res.* 2003; 58:47-56.
 128. Lee TY, Park YK, Jang BC. Anti-adipogenic Effect and Mechanism in 3T3-L1 Preadipocytes by Cyclosporin A, an Immunosuppressant. *Quant Bio-Sci.* 2018; 37:57-63.
 129. Kanitakis J, Thivolet J. Cyclosporine: an immunosuppressant affecting epithelial cell proliferation. *Arch Derm.* 1990; 126:369-375.
 130. High KP. The antimicrobial activities of cyclosporine, FK506, and rapamycin. *Transplant.* 1994; 57:1689-1700.
 131. Rosmarin DM, Lebwohl M, Elewski BE, Gottlieb AB. Cyclosporine and psoriasis: 2008 national psoriasis foundation consensus conference. *J Am Acad Derm.* 2010; 62:838-853.
 132. Vidhi V. Shah, BA, Shivani P. Reddy, BS, Elaine J. Lin, MD, Jashin J. Wu, MD. Cyclosporine: In Therapy for Severe Psoriasis E-Book. Ed: Wu JJ, Feldman SR, Lebwohl MG Expert Consult. Elsevier Health Sci. 2016. 63-73 p.
 133. Franke EK, Luban J. Inhibition of HIV-1 replication by cyclosporine A or related compounds correlates with the ability to disrupt the Gag-cyclophilin A interaction. *Viol.* 1996; 222:279-282.
 134. Sokolskaja E, Olivari S, Zufferey M, Strambio-De-Castillia C, Pizzato M, Luban J. Cyclosporine blocks incorporation of HIV-1 envelope glycoprotein into virions. *J Virol.* 2010; 84:4851-4855.
 135. Firpi RJ, Zhu H, Morelli G, Abdelmalek MF, Soldevila-Pico C, Machicao VI, Cabrera R, Reed AI, Liu C, Nelson DR. Cyclosporine suppresses hepatitis C virus *in vitro* and increases the chance of a sustained virological response after liver transplantation. *Liver Transplant.* 2006; 12:51-57.
 136. Fernandes F, Israr-ul HA, Striker R. Cyclosporine inhibits a direct interaction between cyclophilins and hepatitis C NS5A. *PLoS One.* 2010; 5(3):e9815.
 137. Ciesek S, Steinmann E, Wedemeyer H, Manns MP, Neyts J, Tautz N, Madan V, Bartenschlager R, von Hahn T, Pietschmann T. Cyclosporine A inhibits hepatitis C virus nonstructural protein 2 through cyclophilin A. *Hepatology.* 2009; 50:1638-1645.
 138. Xia WL, Shen Y, Zheng SS. Inhibitory effect of cyclosporine A on hepatitis B virus replication *in vitro* and its possible. *Hepatobil Pancreat Dis Int.* 2005; 4:18-22.
 139. Liu X, Zhao Z, Li Z, Xu C, Sun L, Chen J, Liu W. Cyclosporin A inhibits the influenza virus replication through cyclophilin A-dependent and-independent pathways. *PLoS One.* 2012; 7(5):e37277
 140. Carbajo-Lozoya J, Ma-Lauer Y, Malešević M, Theuerkorn M, Kahlert V, Prell E, von Brunn B, Muth D, Baumert TF, Drosten C, Fischer G. Human coronavirus NL63 replication is cyclophilin A-dependent and inhibited by non-immunosuppressive cyclosporine A-derivatives including Alisporivir. *Virus Res.* 2014; 184:44-53.
 141. McKenzie RC, Epanand RM, Johnson DC. Cyclosporine A inhibits herpes simplex virus-induced cell fusion but not virus penetration into cells. *Viol.* 1987; 159:1-9.
 142. Lommer MJ. Efficacy of cyclosporine for chronic, refractory stomatitis in cats: a randomized, placebo-controlled, double-blinded clinical study. *J Vet Den.* 2013; 30:8-17.
 143. Castro AP, Carvalho TM, Moussatché N, Damaso CR. Redistribution of cyclophilin A to viral factories during vaccinia virus infection and its incorporation into mature particles. *J Virol.* 2003; 77:9052-9068.
 144. Kawasaki H, Mocarski ES, Kosugi I, Tsutsui Y. Cyclosporine inhibits mouse cytomegalovirus infection via a cyclophilin-dependent pathway specifically in neural stem/progenitor cells. *J Virol.* 2007; 81:9013-9023.
 145. Bienkowska-Haba M, Williams C, Kim SM, Garcea RL, Sapp M. Cyclophilins facilitate dissociation of the human papillomavirus type 16 capsid protein L1 from the L2/DNA complex following virus entry. *J Virol.* 2012; 86:9875-9887.
 146. Lieberherr C, Zhang G, Grafen A, Singethan K, Kendl S, Vogt V, Maier J, Bringmann G, Schneider-Schaulies J. The plant-derived naphthoquinone Droserone inhibits *in vitro* measles virus infection. *Planta Med.* 2017; 83:232-238.
 147. Cooke RG and Segal W. Colouring matters of Australian plants. I. The structure of droserone. *Austr J Chem.* 1950; 3:628-634.

148. Higa M, Ogihara K, Yogi S. Bioactive naphthoquinone derivatives from *Diospyros maritima* Blume. Chem Pharm Bull. 1998; 46:1189-1193.
149. Krychowiak M, Kawiak A, Narajczyk M, Borowik A, Królicka A. Silver nanoparticles combined with naphthoquinones as an effective synergistic strategy against *Staphylococcus aureus*. Front Pharmacol. 2018; 9:816.
150. Likhitwitayawuid K, Kaewamatawong R, Ruangrungsri N, Krungkrai J. Antimalarial naphthoquinones from *Nepenthes thorelii*. Planta med. 1998; 64:237-241.
151. Gonzaga DT, Gomes RS, Marra RK, Silva FC, Gomes MW, Ferreira DF, Santos R, Pinto A, Ratcliffe NA, Carne-Santos CC, Barros CS. Inhibition of Zika Virus Replication by Synthetic Bis-Naphthoquinones. J Braz Chem Soc. 2019; 30:1697-1706.
152. Polonik SG, Krylova NV, Kompanets GG, Iunikhina OV, Sabutski YE. Synthesis and Screening of Anti-HSV-1 Activity of Thioglucoside Derivatives of Natural Polyhydroxy-1, 4-Naphthoquinones. Nat Prod Comm. 2019; 14:1934578X19860672.
153. Gallelli L. Escin: a review of its anti-edematous, anti-inflammatory, and venotonic properties. Drug Des Dev Ther. 2019; 13:3425.
154. Domanski D, Zegrocka-Stendel O, Perzanowska A, Dutkiewicz M, Kowalewska M, Grabowska I, Maciejko D, Fogtman A, Dadlez M, Kozia K. Molecular mechanism for cellular response to β -escin and its therapeutic implications. PloS one. 2016; 11(10):e0158765
155. Yuan SY, Cheng CL, Wang SS, Ho HC, Chiu KY, Chen CS, Chen CC, Shiau MY, Ou YC. Escin induces apoptosis in human renal cancer cells through G2/M arrest and reactive oxygen species-modulated mitochondrial pathways. Oncol Rep. 2017; 37:1002-1010.
156. Kim JW, Cho H, Kim E, Shim SH, Yang JL, Oh WK. Antiviral escin derivatives from the seeds of *Aesculus turbinata* Blume (Japanese horse chestnut). Bioorg Med Chem Lett. 2017; 27:3019-3025.
157. Yang XW, Zhao J, Cui YX, Liu XH, Ma CM, Hattori M, Zhang LH. Anti-HIV-1 protease triterpenoid saponins from the seeds of *Aesculus chinensis*. J Nat Prod. 1999; 62:1510-1513.
158. Wu CY, Jan JT, Ma SH, Kuo CJ, Juan HF, Cheng YS, Hsu HH, Huang HC, Wu D, Brik A, Liang FS. Small molecules targeting severe acute respiratory syndrome human coronavirus. Pro Natl Acad Sci. 2004; 101:10012-10017.
159. Piao S, Kang M, Lee YJ, Choi WS, Chun YS, Kwak C, Kim HH. Cytotoxic effects of escin on human castration-resistant prostate cancer cells through the induction of apoptosis and G2/M cell cycle arrest. Urol. 2014; 84:982.e1-982.e7.
160. Walter ED. Genistin (an isoflavone glucoside) and its aglucone, genistein, from soybeans. J Am Chem Soc. 1941; 63:3273-3276.
161. Bitto A, Arcoraci V, Alibrandi A, D'Anna R, Corrado F, Atteritano M, Minutoli L, Altavilla D, Squadrito F. Visfatin correlates with hot flashes in postmenopausal women with metabolic syndrome: effects of genistein. Endocr. 2017; 55:899-906.
162. Banerjee S, Li Y, Wang Z, Sarkar FH. Multi-targeted therapy of cancer by genistein. Cancer Lett. 2008; 269:226-242.
163. Shahmohammadi A, Roustae AM, Azadi MR, Fahanik-Babaei J, Baluchnejadmojarad T, Roghani M. Soy isoflavone genistein attenuates lipopolysaccharide-induced cognitive impairments in the rat via exerting anti-oxidative and anti-inflammatory effects. Cytokine 2018; 104:151-159.
164. Arabyan E, Hakobyan A, Kotsinyan A, Karalyan Z, Arakelov V, Arakelov G, Nazaryan K, Simonyan A, Aroutiounian R, Ferreira F, Zakaryan H. Genistein inhibits African swine fever virus replication *in vitro* by disrupting viral DNA synthesis. Antiviral Res. 2018; 156:128-137.
165. LeCher JC, Diep N, Krug PW, Hilliard JK. Genistein has antiviral activity against herpes B virus and acts synergistically with antiviral treatments to reduce effective dose. Viruses 2019; 11:499.
166. Wei B, Cha SY, Kang M, Kim YJ, Cho CW, Rhee YK, Hong HD, Jang HK. Antiviral activity of Chongkukjang extracts against influenza A virus *in vitro* and *in vivo*. J Ethnic Foods. 2015; 2:47-51.
167. Gozlan J, Lathey JL, Spector SA. Human immunodeficiency virus type 1 induction mediated by genistein is linked to cell cycle arrest in G2. J Virol. 1998; 72:8174-8180.
168. Kolokoltsov AA, Adhikary S, Garver J, Johnson L, Davey RA, Vela EM. Inhibition of Lassa virus and Ebola virus infection in host cells treated with the kinase inhibitors genistein and tyrphostin. Arch Virol. 2012; 157:121-127.
169. Qian K, Gao AJ, Zhu MY, Shao HX, Jin WJ, Ye JQ, Qin AJ. Genistein inhibits the replication of avian leucosis virus subgroup J in DF-1 cells. Virus Res. 2014; 192:114-120.
170. Kubo Y, Ishimoto A, Amanuma H. Genistein, a protein tyrosine kinase inhibitor, suppresses the fusogenicity of Moloney murine leukemia virus envelope protein in XC cells. Arch Virol. 2003; 148:1899-1914.
171. Perera A, Ton SH, Palanisamy UD. Perspectives on geraniin, a multifunctional natural bioactive compound. Trends Food Sci Tech. 2015; 44:243-257.
172. Notka F, Meier GR, Wagner R. Inhibition of wild-type human immunodeficiency virus and reverse transcriptase inhibitor-resistant variants by *Phyllanthus amarus*. Antiviral Res. 2003; 58:175-186.
173. Yang CM, Cheng HY, Lin TC, Chiang LC, Lin CC. The *in vitro* activity of geraniin and 1, 3, 4, 6-tetra-O-galloyl- β -D-glucose isolated from *Phyllanthus urinaria* against herpes simplex virus type 1 and type 2 infection. J Ethnopharmacol. 2007; 110:555-558.
174. Li J, Huang H, Zhou W, Feng M, Zhou P. Anti-hepatitis B virus activities of *Geranium carolinianum* L. extracts and identification of the active components. Bio Pharm Bull. 2008; 31:743-747.
175. Yang Y, Zhang L, Fan X, Qin C, Liu J. Antiviral effect of geraniin on human enterovirus 71 *in vitro* and *in vivo*. Bioorg Med Chem Lett. 2012; 22:2209-2211.
176. Xie J, Zhang Y, Wang W. HPLC analysis of glycyrrhizin and licochalcone a in *Glycyrrhiza inflata* from Xinjiang (China). Chem Nat Comp. 2010; 46:148-151.
177. Baba M, Shigeta S. Antiviral activity of glycyrrhizin against varicella-zoster virus *in vitro*. Antiviral Res. 1987; 7:99-107.
178. Hirabayashi K, Iwata S, Matsumoto H, Mori T, Shibata S, Baba M, Ito M, Shigeta S, Nakashima H, Yamamoto N. Antiviral activities of glycyrrhizin and its modified compounds against human immunodeficiency virus type 1 (HIV-1) and herpes simplex virus type 1 (HSV-1) *in vitro*. Chem Pharm Bull. 1991; 39:112-115.
179. Matsumoto Y, Matsuura T, Aoyagi H, Matsuda M, Hmwe SS, Date T, Watanabe N, Watahi K, Suzuki R, Ichinose S, Wake K. Antiviral activity of glycyrrhizin against hepatitis C virus *in vitro*. PloS one. 2013; 8:e68992
180. Cinatl J, Morgenstern B, Bauer G, Chandra P, Rabenau H, Doerr HW. Glycyrrhizin, an active component of liquorice roots, and replication of SARS-associated coronavirus. Lancet. 2003; 361:2045-2046.
181. Hoever G, Baltina L, Michaelis M, Kondratenko R, Baltina L, Tolstikov GA, Doerr HW, Cinatl J. Antiviral Activity of Glycyrrhizic Acid Derivatives against SARS- Coronavirus. J Med Chem. 2005; 48:1256-1259.
182. O'Brian CA, Ward NE, Vogel VG. Inhibition of protein kinase C by the 12-O-tetradecanoylphorbol-13-acetate antagonist glycyrrhetic acid. Cancer Lett. 1990; 49:9-12.

183. Harada S, Karino A, Shimoyama Y, Shamsa F, Ohtsuki K. Identification of glycyrrhizin-binding protein kinase as casein kinase II and characterization of its associated phosphate acceptors in mouse liver. *Biochem Biophys Res Comm.* 1996; 227:102-109.
184. Lee J, Jung E, Park J, Jung K, Park E, Kim J, Hong S, Park J, Park S, Lee S, Park D. Glycyrrhizin induces melanogenesis by elevating a cAMP level in b16 melanoma cells. *J Inv Derm.* 2005; 124:405-411.
185. Yi H, Nakashima I, Isobe KI. Enhancement of nitric oxide production from activated macrophages by glycyrrhizin. *Am J Chinese Med.* 1996; 24:271-278.
186. Akaike T. Role of free radicals in viral pathogenesis and mutation. *Rev Med Virol.* 2001; 11:87-101.
187. Zaki MH, Akuta T, Akaike T. Nitric oxide-induced nitrate stress involved in microbial pathogenesis. *J Pharmacol Sci.* 2005; 98:117-129.
188. Akaike T, Maeda H. Nitric oxide and virus infection. *Immunol.* 2000; 101:300-308.
189. Bogdan C. Nitric oxide synthase in innate and adaptive immunity: an update. *Trends Immunol.* 2015; 36:161-178.
190. Miyake K, Tango T, Ota Y, Mitamura K, Yoshihara M, Kako M, Hayashi S, Ikeda Y, Hayashida N, Iwabuchi S, Sato Y. Efficacy of Stronger Neo-Minophagen C compared between two doses administered three times a week on patients with chronic viral hepatitis. *J Gastroenterol Hepatol.* 2002; 17:1198-1204.
191. Urizar NL, Moore DD. GUGULIPID: a natural cholesterol-lowering agent. *Annu Rev Nut.* 2003; 23:303-313.
192. Gebhard C, Stämpfli SF, Gebhard CE, Akhmedov A, Breitenstein A, Camici GG, Holy EW, Lüscher TF, Tanner FC. Guggulsterone, an anti-inflammatory phytosterol, inhibits tissue factor and arterial thrombosis. *Basic Res Cardiol.* 2009; 104:285-294.
193. Almazari I and Surh YJ. Cancer chemopreventive and therapeutic potential of guggulsterone. In *Natural Products in Cancer Prevention and Therapy*, Springer, Berlin, Heidelberg. 2012. 35-60 p.
194. Bouslama L, Kouidhi B, Alqurashi YM, Chaieb K, Papetti A. Virucidal Effect of Guggulsterone Isolated from *Commiphora gileadensis*. *Planta Med.* 2019; 85:1225-1232.
195. Scholtes C, Diaz O, Icard V, Kaul A, Bartenschlager R, Lotteau V, André P. Enhancement of genotype 1 hepatitis C virus replication by bile acids through FXR. *J Hepatol.* 2008; 48:192-199.
196. Shivanna V, Kim Y, Chang KO. The crucial role of bile acids in the entry of porcine enteric calicivirus. *Virology.* 2014; 456:268-278.
197. Park JY, Jeong HJ, Kim JH, Kim YM, Park SJ, Kim D, Park KH, Lee WS, Ryu YB. Diarylheptanoids from *Alnus japonica* inhibit papain-like protease of severe acute respiratory syndrome coronavirus. *Bio Pharm Bull.* 2012; 60:612-619.
198. Farrand L, Kim JY, Byun S, Im-aram A, Lee J, Suh JY, Lee KW, Lee HJ, Tsang BK. The diarylheptanoid hirsutenone sensitizes chemoresistant ovarian cancer cells to cisplatin via modulation of apoptosis-inducing factor and X-linked inhibitor of apoptosis. *J Bio Chem.* 2014; 289:1723-1731.
199. Joo SS, Kim SG, Choi SE, Kim YB, Park HY, Seo SJ, Choi YW, Lee MW, Lee DI. Suppression of T cell activation by hirsutenone, isolated from the bark of *Alnus japonica*, and its therapeutic advantages for atopic dermatitis. *Eur J Pharmacol.* 2009; 614:98-105.
200. Lee DI, Jang SK, Da WP, Kim ST, Park JS, Jo BR, Park JY, Park HY, Joo SS. Diarylheptanoid hirsutenone attenuates osteoclastogenesis by suppressing IFN γ and NF- κ B signaling in Th1 and preosteoclastic cells. *Bio Pharm Bull.* 2017; 65:630-637.
201. Ganapathy G, Preethi R, Moses JA, Anandharamakrishnan C. Diarylheptanoids as nutraceutical: A review. *Biocatal Agric Biotech.* 2019; 19:101109.
202. Dong ZW, Yuan YF. Juglanin suppresses fibrosis and inflammation response caused by LPS in acute lung injury. *Int J Mol Med.* 2018; 41:3353-3365.
203. Xu F, Guan H, Li G, Liu H. LC method for analysis of three flavonols in rat plasma and urine after oral administration of *Polygonum aviculare* extract. *Chromatogr.* 2009; 69:1251.
204. Zhang FX and Xu RS. Juglanin ameliorates LPS-induced neuroinflammation in animal models of Parkinson's disease and cell culture via inactivating TLR4/NF- κ B pathway. *Biomed Pharmacother.* 2018; 97:1011-1019.
205. Sun ZL, Dong JL, Wu J. Juglanin induces apoptosis and autophagy in human breast cancer progression via ROS/JNK promotion. *Biomed Pharmacother.* 2017; 85:303-312.
206. Schwarz S, Sauter D, Wang K, Zhang R, Sun B, Karioti A, Bilia AR, Efferth T, Schwarz W. Kaempferol derivatives as antiviral drugs against the 3a channel protein of coronavirus. *Planta Med.* 2014; 80:177-182.
207. Yang Y, Yang Z, Liu J, Wu F, Yuan X. Screening of potential anti-influenza agents from *Juglans mandshurica* Maxim. by docking and MD simulations. *Digest J Nanomat Biostruc (DJNB).* 2015; 10:43-57.
208. Krawczyk JM, Völler GH, Krawczyk B, Kretz J, Brönstrup M, Süßmuth RD. Heterologous expression and engineering studies of labyrinthopeptins, class III lantibiotics from *Actinomadura namibiensis*. *Chem Bio.* 2013; 20:111-122.
209. Féris G, Petrova MI, Andrei G, Huskens D, Hoorelbeke B, Snoeck R, Vanderleyden J, Balzarini J, Bartoschek S, Brönstrup M, Süßmuth RD. The lantibiotic peptide labyrinthopeptin A1 demonstrates broad anti-HIV and anti-HSV activity with potential for microbicidal applications. *PLoS one.* 2013; 8:e64010.
210. Prochnow H, Rox K, Birudukota NS, Weichert L, Hotop SK, Klahn P, Mohr K, Franz S, Banda DH, Blockus S, Schreiber J. Labyrinthopeptins exert broad-spectrum antiviral activity through lipid-binding-mediated virolysis. *J Virol.* 2020; 94:e01471-19.
211. Brönstrup M, Prochnow HP, Birudukota NS, Schulz T, Messerle M, Pietschmann T, Haid S, Blockus S, Laqmani-Goffinet C, Franz S, Banda DH, inventors. Labyrinthopeptins as anti-viral agents. United States patent application US 16/665,956. 2020.
212. Hoorelbeke B, Huskens D, Féris G, François KO, Takahashi A, Van Laethem K, Schols D, Tanaka H, Balzarini J. Actinohivin, a broadly neutralizing prokaryotic lectin, inhibits HIV-1 infection by specifically targeting high-mannose-type glycans on the gp120 envelope. *Antimicrobial Agents Chemother.* 2010; 54:3287-3301.
213. Favacho AR, Cintra EA, Coelho LC, Linhares MI. In vitro activity evaluation of *Parkia pendula* seed lectin against human cytomegalovirus and herpes virus 6. *Biologicals* 2007; 35:189-194.
214. Yang DW, Haraguchi Y, Iwai H, Handa A, Shimizu N, Hoshino H. Inhibition of adsorption of human T-cell-leukemia virus type 1 by a plant lectin, wheat-germ agglutinin. *Int J Cancer* 1994; 56:100-105.
215. Vijgen L, Keyaerts E, Van Damme E, Peumans W, De Clercq E, Balzarini J, Van Ranst M. Antiviral effect of plant compounds of the Alliaceae family against the SARS coronavirus. *Antiviral Res.* 2004; 62:A76.
216. Keyaerts E, Vijgen L, Pannecouque C, Van Damme E, Peumans W, Egberink H, Balzarini J, Van Ranst M. Plant lectins are potent inhibitors of coronaviruses by interfering with two targets in the viral replication cycle. *Antiviral Res.* 2007; 75:179-187.
217. Ooi LS, Ho WS, Ngai KL, Tian L, Chan PK, Sun SS, Ooi VE. Narcissus tazetta lectin shows strong inhibitory effects against respiratory syncytial virus, influenza A (H1N1, H3N2, H5N1) and B viruses. *J Biosci.* 2010; 35: 95-103.

218. François KO and Balzarini J. Potential of carbohydrate-binding agents as therapeutics against enveloped viruses. *Med Res Rev.* 2012; 32:349-387.
219. Coleta M, Campos MG, Cotrim MD, de Lima TC, da Cunha AP. Assessment of luteolin (3', 4', 5, 7-tetrahydroxyflavone) neuropharmacological activity. *Behav Brain Res.* 2008; 189:75-82.
220. Leung HW, Kuo CL, Yang WH, Lin CH, Lee HZ. Antioxidant enzymes activity involvement in luteolin-induced human lung squamous carcinoma CH27 cell apoptosis. *Eur J Pharmacol.* 2006; 534:12-18.
221. Funakoshi-Tago M, Nakamura K, Tago K, Mashino T, Kasahara T. Anti-inflammatory activity of structurally related flavonoids, Apigenin, Luteolin and Fisetin. *Int Immunopharmacol.* 2011; 11:1150-1159.
222. Bai L, Nong Y, Shi Y, Liu M, Yan L, Shang J, Huang F, Lin Y, Tang H. Luteolin inhibits hepatitis B virus replication through extracellular signal-regulated kinase-mediated down-regulation of hepatocyte nuclear factor 4 α expression. *Mol Pharm.* 2016; 13:568-577.
223. Yan H, Ma L, Wang H, Wu S, Huang H, Gu Z, Jiang J, Li Y. Luteolin decreases the yield of influenza A virus *in vitro* by interfering with the coat protein I complex expression. *J Nat Med.* 2019; 73:487-496.
224. Fan W, Qian S, Qian P, Li X. Antiviral activity of luteolin against Japanese encephalitis virus. *Virus Res.* 2016; 220:112-116.
225. Peng M, Watanabe S, Chan KW, He Q, Zhao Y, Zhang Z, Lai X, Luo D, Vasudevan SG, Li G. Luteolin restricts dengue virus replication through inhibition of the proprotein convertase furin. *Antiviral Res.* 2017; 143:176-185.
226. Xu L, Su W, Jin J, Chen J, Li X, Zhang X, Sun M, Sun S, Fan P, An D, Zhang H. Identification of luteolin as enterovirus 71 and coxsackievirus A16 inhibitors through reporter viruses and cell viability-based screening. *Viruses.* 2014; 6:2778-2295.
227. Mehla R, Bivalkar-Mehla S, Chauhan A. A flavonoid, luteolin, cripples HIV-1 by abrogation of tat function. *Plos one.* 2011; 6:e27915.
228. Wu CC, Fang CY, Hsu HY, Chen YJ, Chou SP, Huang SY, Cheng YJ, Lin SF, Chang Y, Tsai CH, Chen JY. Luteolin inhibits Epstein-Barr virus lytic reactivation by repressing the promoter activities of immediate-early genes. *Antiviral Res.* 2016; 132:99-110.
229. Dai C, Ma Y, Zhao Z, Zhao R, Wang Q, Wu Y, Cao Z, Li W. Mucroporin, the first cationic host defense peptide from the venom of *Lychas mucronatus*. *Antimicrob Agents Chemother.* 2008; 52:3967-3972.
230. Li Q, Zhao Z, Zhou D, Chen Y, Hong W, Cao L, Yang J, Zhang Y, Shi W, Cao Z, Wu Y. Virucidal activity of a scorpion venom peptide variant mucroporin-M1 against measles, SARS-CoV and influenza H5N1 viruses. *Peptides.* 2011; 32:1518-1525.
231. Zhao Z, Hong W, Zeng Z, Wu Y, Hu K, Tian X, Li W, Cao Z. Mucroporin-M1 inhibits hepatitis B virus replication by activating the mitogen-activated protein kinase (MAPK) pathway and down-regulating HNF4 α *in vitro* and *in vivo*. *J Bio Chem.* 2012; 287:30181-30190.
232. Franklin TJ and Cook JM. The inhibition of nucleic acid synthesis by mycophenolic acid. *Biochem J.* 1969; 113:515-524.
233. Mitsui A and Suzuki S. Immunosuppressive effect of mycophenolic acid. *J Antibiotics* 1969; 22: 358-363.
234. Noto T, Sawada M, Ando K, Koyama K. Some biological properties of mycophenolic acid. *J Antibiotics.* 1969; 22:165-169.
235. Domhan S, Muschal S, Schwager C, Morath C, Wirkner U, Ansoorge W, Maercker C, Zeier M, Huber PE, Abdollahi A. Molecular mechanisms of the antiangiogenic and antitumor effects of mycophenolic acid. *Mol Cancer Ther.* 2008; 7:1656-1668.
236. Planterose DN. Antiviral and cytotoxic effects of mycophenolic acid. *J Gen Virol.* 1969; 4:629-630.
237. Diamond MS, Zachariah M, Harris E. Mycophenolic acid inhibits dengue virus infection by preventing replication of viral RNA. *Virol.* 2002; 304:211-221.
238. Smee DF, Bray M, Huggins JW. Antiviral activity and mode of action studies of ribavirin and mycophenolic acid against orthopoxviruses *in vitro*. *Antiviral Chem Chemother.* 2001; 12:327-335.
239. Wang Y, Zhou X, Debing Y, Chen K, Van Der Laan LJ, Neyts J, Janssen HL, Metselaar HJ, Peppelenbosch MP, Pan Q. Calcineurin inhibitors stimulate and mycophenolic acid inhibits replication of hepatitis E virus. *Gastroenterol.* 2014; 146:1775-1783.
240. Chapuis AG, Rizzardi GP, D'agostino C, Attinger A, Knabenhans C, Fleury S, Acha-Orbea H, Pantaleo G. Effects of mycophenolic acid on human immunodeficiency virus infection *in vitro* and *in vivo*. *Nat Med.* 2000; 6:762-768.
241. Henry SD, Metselaar HJ, Lonsdale RC, Kok A, Haagsmans BL, Tilanus HW, Van der Laan LJ. Mycophenolic acid inhibits hepatitis C virus replication and acts in synergy with cyclosporin A and interferon- α . *Gastroenterol.* 2006; 131:1452-1462.
242. Sebastian L, Madhusudana SN, Ravi V, Desai A. Mycophenolic acid inhibits replication of Japanese encephalitis virus. *Chemother.* 2011; 57:56-61.
243. Robertson CM, Hermann LL, Coombs KM. Mycophenolic acid inhibits avian reovirus replication. *Antiviral Res.* 2004; 64:55-61.
244. Cheng KW, Cheng SC, Chen WY, Lin MH, Chuang SJ, Cheng IH, Sun CY, Chou CY. Thiopurine analogs and mycophenolic acid synergistically inhibit the papain-like protease of Middle East respiratory syndrome coronavirus. *Antiviral Res.* 2015; 115:9-16.
245. Kassuya CA, Silvestre A, Menezes-de-Lima Jr O, Marotta DM, Rehder VL, Calixto JB. Anti-inflammatory and anti-platelet actions of the lignan niranthin isolated from *Phyllanthus amarus*: evidence for interaction with platelet activating factor receptor. *Eur J Pharmacol.* 2006; 546:182-188.
246. Huang RL, Huang YL, Ou JC, Chen CC, Hsu FL, Chang C. Screening of 25 compounds isolated from *Phyllanthus* species for anti-human hepatitis B virus *in vitro*. *Phytother Res.* 2003; 17:449-453.
247. Liu S, Wei W, Shi K, Cao X, Zhou M, Liu Z. *In vitro* and *in vivo* anti-hepatitis B virus activities of the lignan niranthin isolated from *Phyllanthus niruri* L. *J Ethnopharmacol.* 2014; 155:1061-1067.
248. Loan LT, Uyen NH, Phuong VH, Cuong DV, Anh PV, Hanh NN, Anh LT. Herbal Extract Effects on White Spot Syndrome Virus (WSSV) in Shrimp (*Penaeus monodon*). *The Israeli Journal of Aquaculture -Bamidgeh,* 2009; 61:1-3.
249. Canel C, Moraes RM, Dayan FE, Ferreira D. Podophyllotoxin. *Phytochemistry.* 2000; 54: 115-120.
250. Inamori Y, Kubo M, Tsujibo H, Ogawa M, Baba K, Kozawa M, Fujita E. The biological activities of podophyllotoxin compounds. *Chem Pharm Bull* 1986; 34:3928-3932.
251. Abubacker MN, Vasantha S. Antibacterial activity of ethanolic leaf extract of *Rauwolfia tetraphylla* (Apocyanaceae) and its bioactive compound reserpine. *Drug Invent Today.* 2011; 3:16-17.
252. Gordaliza M, Castro MA, García-Grávalos MD, Ruiz P, Del Corral JM, Feliciano AS. Antineoplastic and antiviral activities of podophyllotoxin related lignans. *Archiv der Pharmazie.* 1994; 327:175-179.
253. Castro MA, del Corral JM, Gordaliza M, Gomez-Zurita MA, de La Puente ML, Betancur-Galvis LA, Sierra J, San Feliciano A. Synthesis, cytotoxicity and antiviral activity of

- podophyllotoxin analogues modified in the E-ring. Eur J Med Chem. 2003; 38:899-911.
254. Chen SW, Wang YH, Jin Y, Tian X, Zheng YT, Luo DQ, Tu YQ. Synthesis and anti-HIV-1 activities of novel podophyllotoxin derivatives. Bioorg Med Chem Lett. 2007; 17:2091-2095.
255. Ternaux JP and Portalier P. Effect of quercetine on survival and morphological properties of cultured embryonic rat spinal motoneurons. Neurosci Lett. 2002; 332:33-36.
256. Pitoyo FL and Fatmawati H. The effect of quercetine to reduced triglyceride and blood glucose level in animal model diet-induced obesity. J Med Planta. 2012; 1:36-46.
257. Tanir HM, Sener T, Inal M, Akyuz F, Uzuner K, Sivri E. Effect of quercetine and glutathione on the level of superoxide dismutase, catalase, malonyldialdehyde, blood pressure and neonatal outcome in a rat model of pre-eclampsia induced by NG-nitro-L-arginine-methyl ester. Eur J Obst Gyn Rep Bio. 2005; 118:190-195.
258. Abdelmoaty MA, Ibrahim MA, Ahmed NS, Abdelaziz MA. Confirmatory studies on the antioxidant and antidiabetic effect of quercetin in rats. Ind J Clin Biochem. 2010; 25:188-192.
259. Kaul TN, Middleton Jr E, Ogra PL. Antiviral effect of flavonoids on human viruses. J Med Virol. 1985; 15:71-79.
260. Mahmood N, Piacente S, Pizza C, Burke A, Khan AI, Hay AJ. The Anti-HIV Activity and Mechanisms of Action of Pure Compounds Isolated from *Rosa damascena*. Biochem Biophys Res Comm. 1996; 229:73-79.
261. Ciofalo F, Levitt B, Roberts J. Some aspects of the anti-arrhythmic activity of reserpine. Br J Pharmacol Chemother. 1966; 28:44.
262. Abubacker MN and Vasantha S. Antibacterial activity of ethanolic leaf extract of *Rauwolfia tetraphylla* (Apocyanaceae) and its bioactive compound reserpine. Drug Invent Today. 2011; 3:16-17.
263. Fernandes VS, Santos JR, Leão AH, Medeiros AM, Melo TG, Izídio GS, Cabral A, Ribeiro RA, Abílio VC, Ribeiro AM, Silva RH. Repeated treatment with a low dose of reserpine as a progressive model of Parkinson's disease. Behav Brain Res. 2012; 23:154-163.
264. Kesharwani A, Polachira SK, Nair R, Agarwal A, Mishra NN, Gupta SK. Anti-HSV-2 activity of *Terminalia chebula* Retz extract and its constituents, chebulagic and chebulinic acids. BMC Compl Alt Med. 2017; 17:110.
265. Lewis JJ. Rauwolfia derivatives. Physiol Pharmacol. 2017; 1:479-536.
266. Kuo YC, Kuo YH, Lin YL, Tsai WJ. Yatein from *Chamaecyparis obtusa* suppresses herpes simplex virus type 1 replication in HeLa cells by interruption the immediate-early gene expression. Antiviral Res. 2006; 70:112-120.
267. Cui Q, Du R, Liu M, Rong L. Lignans and their derivatives from plants as antivirals. Mol. 2020; 25:183.
268. Wang Y, Wang X, Xiong Y, Kaushik AC, Muhammad J, Khan A, Dai H, Wei DQ. New strategy for identifying potential natural HIV-1 non-nucleoside reverse transcriptase inhibitors against drug-resistance: An *in silico* study. J Biomol Struc Dynam. 2019; 30:1-5.
269. Shimizu JF, Lima CS, Pereira CM, Bittar C, Batista MN, Nazaré AC, Polaquini CR, Zothner C, Harris M, Rahal P, Regasini LO. Flavonoids from *Pterogyne nitens* inhibit hepatitis C virus entry. Sci Rep. 2017; 7:1-9.
270. Jo S, Kim S, Shin DH, Kim MS. Inhibition of SARS-CoV 3CL protease by flavonoids. J Enz Inhib Med Chem. 2020; 35:145-151.
271. Jo S, Kim H, Kim S, Shin DH, Kim MS. Characteristics of flavonoids as potent MERS-CoV 3C-like protease inhibitors. Chem Bio Drug Des. 2019; 94:2023-2030.
272. Wen CC, Kuo YH, Jan JT, Liang PH, Wang SY, Liu HG, Lee CK, Chang ST, Kuo CJ, Lee SS, Hou CC. Specific plant terpenoids and lignoids possess potent antiviral activities against severe acute respiratory syndrome coronavirus. J Med Chem. 2007; 50:4087-4895.
273. Chen M, Kilgore N, Lee KH, Chen DF. Rubrisandrins A and B, lignans and related anti-HIV compounds from *Schisandra rubriflora*. J Nat Prod. 2006; 69:1697-1701.
274. Tian RR, Xiao WL, Yang LM, Wang RR, Sun HD, Liu NF, Zheng YT. The isolation of Rubrifloralignan A and its anti-HIV-1 activities. Chin J Nat Med. 2006; 4:40-45.
275. Jiang ZY, Yu YJ, Huang CG, Huang XZ, Hu QF, Yang GY, Wang HB, Zhang XY, Li GP. Icetexane diterpenoids from *Perovskia atriplicifolia*. Planta Med. 2015; 81:241-246.
276. Liu AL, Yang F, Zhu M, Zhou D, Lin M, Lee SM, Wang YT, Du GH. *In vitro* anti-influenza viral activities of stilbenoids from the lianas of *Gnetum pendulum*. Planta Med. 2010; 76:1874-1876.
277. Lavoie S, Côté I, Pichette A, Gauthier C, Ouellet M, Nagau-Lavoie F, Mshvildadze V, Legault J. Chemical composition and anti-herpes simplex virus type 1 (HSV-1) activity of extracts from *Cornus canadensis*. BMC Compl Altern Med. 2017; 17:123.
278. Brandt CR, Piraino F. Mushroom antivirals. Recent Res Dev Antimicrob Agents Chemother. 2000; 4:11-26.
279. Piraino FF. Emerging antiviral drugs from medicinal mushrooms. Int J Med Mushrooms 2006; 8:101-114.
280. Yasuhara-Bell J, Lu Y. Marine compounds and their antiviral activities. Antiviral Res. 2010; 86:231-340.
281. Wang W, Wang SX, Guan HS. The antiviral activities and mechanisms of marine polysaccharides: an overview. Mar Drugs. 2012; 10:2795-2816.
282. Dang VT, Benkendorff K, Green T, Speck P. Marine snails and slugs: a great place to look for antiviral drugs. J Virol. 2015; 89:8114-8118.
283. Abdelmohsen UR, Balasubramanian S, Oelschlaeger TA, Grkovic T, Pham NB, Quinn RJ, Hentschel U. Potential of marine natural products against drug-resistant fungal, viral, and parasitic infections. Lancet Infect Dis. 2017; 17:e30-41.
284. Van der Meer FJ, de Haan CA, Schuurman NM, Haijema BJ, Verheije MH, Bosch BJ, Balzarini J, Egberink HF. The carbohydrate-binding plant lectins and the non-peptidic antibiotic pradimicin A target the glycans of the coronavirus envelope glycoproteins. J Antimicrob Chemother. 2007; 60:741-749.
285. Haid S, Blockus S, Wiechert SM, Wetzke M, Prochnow H, Dijkman R, Wiegmann B, Rameix-Welti MA, Eleouet JF, Duprex P, Thiel V. Labyrinthopeptin A1 and A2 efficiently inhibit cell entry of hRSV isolates. Eur Res. J 2017 50:PA4124.