

**Effect of Aqueous Extract of *Momordica charantia* on Survival, Locomotive Behaviour and Antioxidant Status of *Drosophila melanogaster***Opeyemi C. De-Campos^{1,2*}, Modupe P. Layole¹, Franklyn N. Iheagwam^{1,2}, Solomon O. Rotimi^{1,2}, Shalom N. Chinedu^{1,2}¹Department of Biochemistry, College of Science and Technology, Covenant University, Canaan Land, PMB 1023 Ota, Ogun State, Nigeria²Covenant University Public Health and Wellbeing Research Cluster (CUPHERC), Covenant University, Canaan Land, PMB 1023 Ota, Ogun State, Nigeria

ARTICLE INFO

ABSTRACT

Article history:

Received 10 October 2020

Revised 18 November 2020

Accepted 28 January 2021

Published online 03 February 2021

Copyright: © 2021 De-Campos *et al.* This is an open-access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Momordica charantia, commonly known as bitter melon, is a therapeutic plant popularly known for its antidiabetic potential in folklore medicine. This study investigated the effect of aqueous leaf extract of *M. charantia* (AMC) on survival rate, locomotive behaviour and antioxidant response in *Drosophila melanogaster*. Adult flies were fed with different concentrations of AMC (0-100 mg/mL) for 12 days, and their locomotive behaviour and whole-body antioxidant status were assessed at 0, 2, 4 and 8 mg/mL of AMC. Results showed a significant decrease ($p < 0.05$) in the survival rate and locomotive behaviour of flies at 8, 16 and 50 mg/mL of AMC compared to the control. There was no significant difference in malondialdehyde content, GSH level and SOD activity of flies exposed to 2, 4, and 8 mg/mL of AMC compared to the control group. Glutathione-s-transferase activity at 8 mg/mL of AMC increased significantly ($p < 0.05$) when compared to the control group. Acetylcholinesterase activity also increased in a dose-dependent manner with a significant increase at 4 and 8 mg/mL of AMC. The evidence from this study suggests that low to moderate doses of aqueous extract of *Momordica charantia* slightly improved survival rate of flies. It also increased the activities of acetylcholinesterase and antioxidant enzymes.

Keywords: *M. charantia*, Survival rate, *D. melanogaster*, Locomotive Behaviour, Antioxidant.

Introduction

Plants rich in phytochemicals and phytonutrients have played an essential role in the overall health and wellness of several individuals in the world for ages.^{1,2} In developing countries, they still serve as the primary source of health care among some communities while in some developed countries, they serve as alternative and complementary medicines.^{3,4} *Momordica charantia* L. is one of the numerous therapeutic plants known today. It is used in ethnomedicine to manage and treat ulcers, jaundice, type 2 diabetes, viral and bacterial diseases.⁵ It is commonly called bitter melon or bitter melon because of its different parts' bitter taste.⁵ Locally, it is known as "Ejirin" among the Yorubas, Alo-ose among the Igbos and Kakayi among the Hausas in Nigeria.⁶ Apart from its therapeutic value, it is also known for its nutritional properties. Unlike most bitter vegetables, the bitter taste of the fruit of *M. charantia* is considered suitable for consumption. It is often added to other vegetables and food to give a bitter taste and flavour.^{5,7} The whole herb can also be boiled and taken as a tea. The leaves, fruits, stems, and seeds contain proteins, carbohydrates, lipids, minerals and vitamins.^{5,8} *Drosophila melanogaster*, commonly called fruit fly, is used as a model organism for studying different human diseases.⁹ About 75% of the genome of *D. melanogaster* are functionally related to that of humans. The fruit fly also produces a large number of offspring within

the shortest time. It is small, not costly to maintain in the laboratory and has a short lifespan, making it a viable model organism.⁹⁻¹¹ Despite the extensive use of *M. charantia* in folklore medicine and the various scientific studies validating its therapeutic potential, its toxicity has not been fully explored, particularly in *D. melanogaster*. This study examined the effect of aqueous extract of *M. charantia* (AMC) on survival, locomotive behaviour and antioxidant status in *D. melanogaster*.

Materials and Methods

Preparation of plant material

M. charantia vines were harvested fresh, in October 2018, from local farms in Ota, Ogun State, Nigeria, and the leaves were separated. A botanist, Dr. J. O. Popoola, at the Department of Biological Sciences, Covenant University, Ota Ogun State, identified the plant and voucher specimen (with number; FHI 112033) was kept at the Forestry Research Institute of Nigeria. The leaves were air-dried, ground to powder and extracted with distilled water (400 g in 4 L) for 72 h. The resulting extract was evaporated to dryness under vacuum in a rotary evaporator at 55°C.

Strain and culture of *D. melanogaster*

Harwich strain (wild-type) of *D. melanogaster* was raised at the Department of Biochemistry, Covenant University Ota. Male and female flies were fed brewer's yeast paste on an apple agar medium in a standard egg collecting cage. Eggs (6 hours old) were collected, rinsed in sterile 1x phosphate buffer saline (PBS) and used for further experiment.

Survival assay

D. melanogaster eggs were raised on a regular diet (containing 10 g of agar powder, 15 g of yeast, 50 g of sugar, 50 g of semolina and 1.2 g of paraben) and different concentrations of AMC (0, 1, 2, 4, 16, 32, 50

*Corresponding author. E mail:

opeyemi.decampos@covenantuniversity.edu.ng
Tel: +2347062246775

Citation: De Campos OC, Layole MP, Iheagwam FN, Rotimi SO, Chinedu SN. Effect of Aqueous Extract of *Momordica charantia* on Survival, Locomotive Behaviour and Antioxidant Status of *Drosophila melanogaster*. Trop J Nat Prod Res. 2021; 5(1):178-181. doi.org/10.26538/tjnpr/v5i1.23

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

and 100 mg/mL). The developmental process was monitored until moulting into the adult stage. Adult flies (1-3 days old), 50 per vial, enclosed from embryos raised on different concentration of AMC were further exposed to regular diet and different concentrations (0, 1, 2, 4, 16, 32, 50 and 100 mg/mL) of AMC for 12 days. The mortality and survival rates of the flies were monitored each day for 12 days. The diet mixed with different concentration of AMC was replaced every 72 h. The results were presented as the percentage of life flies.¹²

Negative geotaxis assay

Flies treated with different concentrations of AMC were examined for their locomotive performance, using previously described methods.^{12,13} Data generated were analyzed and represented as a percentage of the mean number of flies above the 6 cm mark.

Whole fly homogenate preparation

After exposure of adult flies to different concentrations of AMC for 12 days, 10 flies were slightly made inactive under mild ice. The flies were homogenized in homogenizing buffer containing 1 mM EDTA, 0.25 M sucrose and 10 mM HEPES NaOH pH 7.4. The homogenate was centrifuged at 4000 rpm for 10 min at 4°C in a refrigerated centrifuge. After centrifugation, the supernatant was stored at -20°C until the time of use.¹⁴

Assessment of antioxidant status and acetylcholinesterase activity

The extent of lipid peroxidation was assayed spectrophotometrically by determining malondialdehyde (MDA) concentration in the fly homogenate.¹⁵ Glutathione-s-transferase (GST) activity was evaluated based on the method of Habig *et al.*¹⁶ while superoxide dismutase (SOD) activity was evaluated based on pyrogallol autoxidation method described previously.¹⁷ Glutathione level in fly homogenates was estimated according to the method described by Moron *et al.*¹⁸ Acetylcholinesterase activity was assessed in fly homogenate based on the hydrolysis of acetylcholine iodide as reported by Ellman *et al.*¹⁹

Total protein determination

The total protein level in the fly homogenates was determined by Lowry's method using bovine serum albumin as a standard.²⁰

Statistical analysis

Data generated in this study were subjected to one-way analysis of variance (ANOVA) using Statistical Package for the Social Sciences (version 20.0, SPSS Inc., Chicago, IL, USA). Duncan's multiple range test was used to compare the level of heterogeneity among groups at $p < 0.05$. The results are presented as mean \pm SEM of three biological replicates

Results and Discussion

M. charantia is a medicinal plant with both therapeutic and nutritional properties. Despite its wide use in folklore medicine, there are still growing concerns about its toxic effect. Also, little is known on how the plant extract affects the survival rate, locomotive behaviour and antioxidant response in *D. melanogaster*. This current study found that AMC caused the death of all flies at 100 mg/mL. There was no significant difference in the survival rate of adult flies exposed to 1, 2 and 4 mg/ml of AMC when compared to the control. However, the survival rate of flies reduced significant reduction ($p < 0.05$) at 8, 16 and 50 mg/mL of AMC when compared to the control (Figure 1A and 1B). Findings from this study also showed that the locomotive behaviour of flies was significantly ($p < 0.05$) reduced when compared to the control, at 4, 8, 16 and 50 mg/mL of AMC (Figure 1C). Based on the result of the survival rate and negative geotaxis assay, AMC concentrations of 2, 4 and 8 mg/ml were chosen to evaluate the antioxidant status and acetylcholinesterase activity of *D. melanogaster*

that were fed low to moderate doses of AMC. Findings from the antioxidant evaluation showed that there was no significant difference ($p < 0.05$) in MDA content of flies exposed to 2, 4, and 8 mg/mL of AMC when compared to the control group (Table 1). There was also no significant difference in the GSH content of the treated groups when compared to the control (Table 1). Glutathione-s-transferase activity however increased significantly ($p < 0.05$) in a dose-dependent manner among the treated groups when compared to the control group (Table 1). The increase was significant at 8 mg/ml of AMC ($p < 0.05$). Superoxide dismutase and acetylcholinesterase activities also increased in a dose-dependent manner. However, for acetylcholinesterase activity, the increase was significant at 4 and 8 mg/ml of AMC (Figure 2).

The fact that AMC, at 100 mg/ml, caused 100% mortality of flies even before the end of the experimental period indicates a lethal dose which affects the survival and life span of adult flies. Although AMC at 50 mg/ml did not lead to 100% mortality of adult flies, it significantly reduced the survival rate and locomotive performance of the flies. Based on these findings, malondialdehyde content and antioxidant status of flies exposed to 2, 4 and 8 mg/ml of AMC were assessed.

Malondialdehyde is one of the secondary products of lipid peroxidation and a prominent biomarker for assessing the extent at which reactive oxygen species react with lipids. In most biological systems, the effects of reactive oxygen species and other oxidants are neutralized by enzymatic and non-enzymatic antioxidants.²¹ Glutathione-s-transferase and superoxide dismutase are two important enzymatic antioxidants that help to reduce the harmful effects of reactive oxygen species and their conjugated products.^{22,23} The fact that MDA levels among the flies exposed to 2, 4 and 8 mg/mL of AMC compared well with that of the control suggest oxidant production within the ability of endogenous antioxidants defence system. Also, flies exposed to AMC had a dose-dependent increase in superoxide dismutase and glutathione-s-transferase activities. However, for glutathione-s-transferase activity, the increase was significant at 8 mg/mL of AMC. This suggests that AMC induces synthesis of the antioxidant enzyme, glutathione-s-transferase, and thus have antioxidant activity in *D. melanogaster*. Also, the fact that AMC increased acetylcholinesterase activity in a dose-dependent manner suggests that at low to moderate doses, AMC could be an excellent activator of acetylcholinesterase. Acetylcholinesterase is involved in the metabolism of acetylcholine, an important neurotransmitter that plays a crucial role in the nervous system.²⁴ Inhibition of acetylcholinesterase causes acetylcholine levels to increase and leads to muscular weakness and paralysis. Though a decrease in acetylcholinesterase activity is often associated with locomotive defects^{12,13} in *D. melanogaster*, there are contradictory reports about this. One study showed that, though hydroalcoholic extract of *Croton Campestris* affected the locomotive performance of flies at 0.1- 50 mg/mL, it did not cause any observable difference in acetylcholinesterase activity.²⁵ Findings from this study corroborate previous studies that showed that medicinal plants are toxic at moderate to high doses in *D. melanogaster*.^{13,26,27} The findings observed in this study also mirror those of the previous studies that have examined the toxic effect of *M. charantia* studies in other models. A recent study showed that the LD₅₀ of seed extract of *M. charantia* was 50 μ g/mL. In comparison, the fruit extract did not cause any toxic effect at a dose of 200 μ g/mL in Zebrafish embryos.²⁸ Another study showed that *M. charantia*, at a dose of 1000 mg/kg body weight was not toxic to male and female Sprague-Dawley rats.²⁹

Conclusion

The evidence from this study suggests that low to moderate doses of aqueous extract of *Momordica charantia* slightly improved survival rate of flies. It also increased the activities of acetylcholinesterase and antioxidant enzymes.

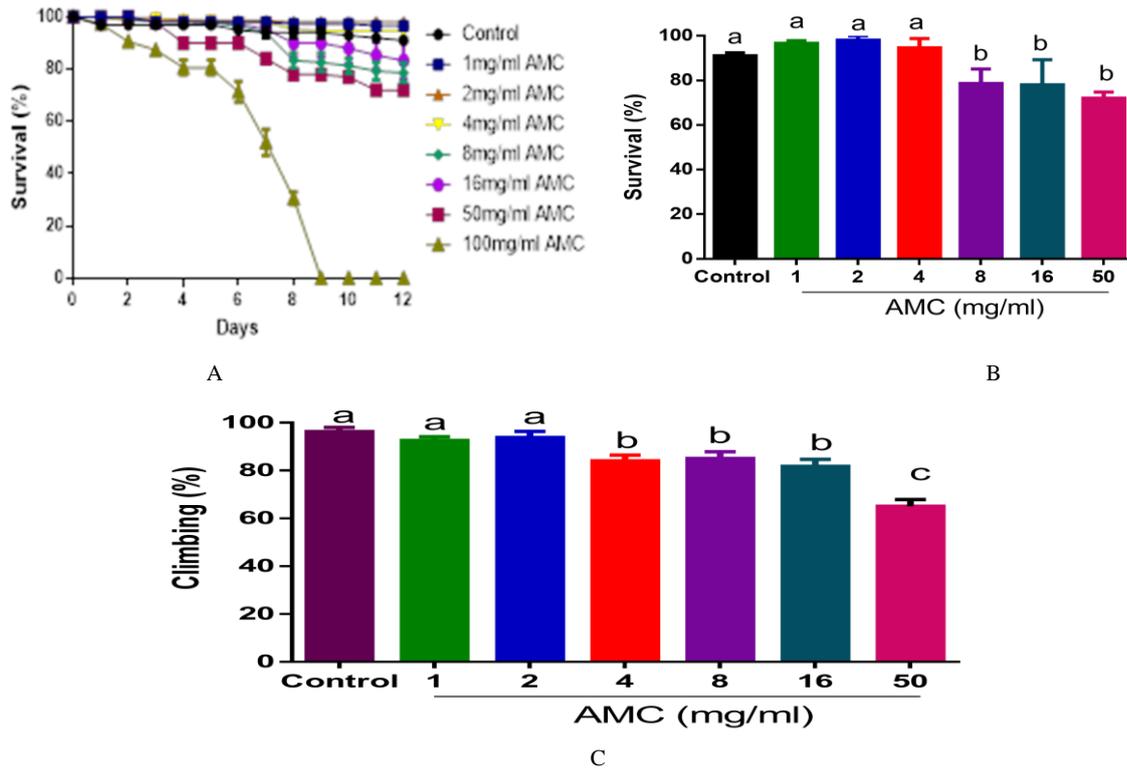


Figure 1: Effect of AMC on survival rate and locomotive behaviour in *D. melanogaster*

The results represent the mean \pm SEM of three biological replicates. Bars with different letters are significantly different. ($p < 0.05$). (A) survival rate curve; (B) survival rate (%) (C) Locomotive behaviour after 12 days exposure of *Drosophila melanogaster* to AMC.

Table 1: Effect of AMC on MDA content and antioxidant status

	MDA content (nM)	SOD activity (U/mg protein)	GST activity (U/mg protein)	GSH (μ M)
Control	72.61 \pm 2.14 ^a	0.01 \pm 0.00 ^a	0.07 \pm 0.01 ^a	5.44 \pm 1.64 ^a
2 mg/ml AMC	87.61 \pm 9.1 ^a	0.03 \pm 0.00 ^a	0.13 \pm 0.01 ^a	3.69 \pm 0.59 ^a
4 mg/ml AMC	86.54 \pm 9.61 ^a	0.04 \pm 0.01 ^a	0.16 \pm 0.02 ^a	3.11 \pm 0.35 ^a
8 mg/ml AMC	86.54 \pm 3.21 ^a	0.04 \pm 0.01 ^a	0.29 \pm 0.07 ^b	3.17 \pm 0.89 ^a

The results represent the mean \pm SEM of three biological replicates. Values on the same columns with different letters are significantly different ($p < 0.05$).

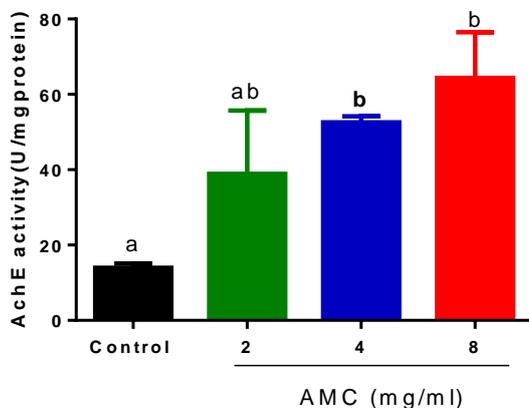


Figure 2: Effect of AMC on acetylcholinesterase activity. The result represents the mean \pm SEM of three biological replicates. Bars with different letters are significantly different ($p < 0.05$).

Conflict of interest

The authors declare no conflict of interest.

Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

Acknowledgements

The authors appreciate Covenant University for paying the article processing charge of this article. The authors also appreciate Dr. Amos O. Abolaji of the Department of Biochemistry, the University of Ibadan for supplying the *Drosophila melanogaster* strain used in this study.

References

1. Yakubu OF, Adebayo AH, Famakinwa TO, Adegbite OS, Ishola TA, Imonikhe LO, Adeyemi OA, Awotoye OA, Iweala EEJ. Antimicrobial and toxicological studies of *Ricnodendron Heudelotii* (Baill.). *Asian J Pharm Clin Res.* 2018; 11:299-305.
2. Iheagwam NF, Okeke CO, Decampos OC, Okere DU, Ogunlana OO, Chinedu SN. Safety evaluation of *Terminalia catappa* Linn (Combretaceae) aqueous leaf extract: Sub-acute cardio-toxicopathological studies in albino Wistar rats. *J Phys Conf Ser.* 2019; 1299 012109:1-6.
3. Mazzari ALDA and Prieto JM. Herbal medicines in Brazil: Pharmacokinetic profile and potential herb-drug interactions. *Front Pharmacol.* 2014; 5:162.
4. Amaeze OU, Aderemi-Williams RI, Ayo-Vaughan MA, Ogundemuren DA, Ogunmola DS, Anyika EN. Herbal medicine use among Type 2 diabetes mellitus patients in Nigeria: understanding the magnitude and predictors of use. *Int J Clin Pharm.* 2018; 40(3):580-588.
5. Dandawate PR, Subramaniam D, Padhye SB, Anant S. Bitter melon: A panacea for inflammation and cancer, *Chin J Nat Med.* 2016; 14(2):81-100.
6. Akinsiku AA, Ajanaku KO, Adebisi AA, Edobor-Osoh A, Aladesuyi O, Samson TO, Dare EO. *Momordica charantia* stem extract mediated biogenic synthesis of silver nanoparticles: Optical and antimicrobial efficacy. *IOP Conf Ser Mater Sci Eng.* 2019; 509(1):012018.
7. Farooqi AA, Khalid S, Tahir F, Sabitaliyevich UY, Yaylim I, Attarf R, Xu, B. Bitter gourd (*Momordica charantia*) as a rich source of bioactive components to combat cancer naturally: Are we on the right track to fully unlock its potential as inhibitor of deregulated signalling pathways. *Food Chem Toxicol.* 2018; 119:98-105.
8. Joseph B and Jini D. Antidiabetic effects of *Momordica charantia* (bitter melon) and its medicinal potency. *Asian Pacific J Trop Dis.* 2013; 3(2):93-102.
9. Yamaguchi M and Yoshida H. *Drosophila* as a model organism. *Adv Exp Med Biol.* 2018; 1076:1-10.
10. Aryal B and Lee Y. Disease model organism for Parkinson disease: *Drosophila melanogaster*. *BMB Rep.* 2019; 52(4):250-258.
11. Baenas N and Wagner AE. *Drosophila melanogaster* as an alternative model organism in nutrigenomics. *Genes Nutr.* 2019; 14(1):1-11
12. Abolaji OA, Kamdem PJ, Lugokenski TH, Nascimento KT, Waczuk PE, Farombi, OE Loreto DEL, Rocha TJB. Involvement of oxidative stress in 4-vinylcyclohexene-induced toxicity in *Drosophila melanogaster*, *Free Radic Biol Med.* 2014; 71:99-108.
13. Pinho FVSD-A, Da Silva GF, Macedo GE, Muller KR, Martins IK, Ternes APL, da Costa JGM, Athayde ML, Boligon AA, Kamdem JP, Franco JL, de Menezes IRA, and Posser T. Phytochemical constituents and toxicity of *Duguetia furfuracea* hydroalcoholic extract in *Drosophila melanogaster*. *Evidence-based Compl Altern Med.* 2014; 2014:838101.
14. Rotimi SO, Bankole GE, Adelani IB, Rotimi OA. Hesperidin prevents lipopolysaccharide-induced endotoxicity in rats. *Immunopharmacol Immunotoxicol.* 2016; 38(5):364-371.
15. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem.* 1979; 95(2):351-358.
16. Habig WH, Pabst MJ, Jakoby WB. Glutathione S transferases. The first enzymatic step in mercapturic acid formation. *J Biol Chem.* 1974; 249(22):7130-7139.
17. Marklund S and Marklund G. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *Eur J Biochem.* 1974; 47(3):469-474.
18. Moron MS, Depierre JW, Mannervik B. Levels of glutathione, glutathione reductase and glutathione S-transferase activities in rat lung and liver. *BBA - Gen Subj.* 1979; 582(1):67-78.
19. Ellman GL, Courtney KD, Andres V, Featherstone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol.* 1961; 7(2):88-95.
20. Lowry OH, Rosebrough NJ, Farr AI, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem.* 1951; 193(1):265-275.
21. Mirończuk-Chodakowska I, Witkowska AM, Zujko ME. Endogenous non-enzymatic antioxidants in the human body. *Adv Med Sci.* 2018; 63(1):68-78.
22. Robaczewska J, Kedziora-Kornatowska K, Kozakiewicz M, Zary-Sikorska E, Pawluk H, Pawlitzak W, Kedziora J. Role of glutathione metabolism and glutathione-related antioxidant defense systems in hypertension. *J Physiol Pharmacol.* 2016; 67(3):331-337.
23. Wang Y, Branicky R, Noë A, Hekimi S. Superoxide dismutases: Dual roles in controlling ROS damage and regulating ROS signalling. *J Cell Biol.* 2018; 217(6):1915-1928.
24. Thapa S, Lv M, Xu H. Acetylcholinesterase: A primary target for drugs and insecticides. *Mini-Rev Med Chem.* 2017; 17(17):1665-1676.
25. Francisco EBJ, Echeverria MG, Paula ZA, da Silva FG, da Cruz CL, Aline AB, de Menezes IRA., Jeferson LF Thaís P. Oxidant effects and toxicity of *Croton campestris* in *Drosophila melanogaster*. *Pharm Biol.* 2016; 54:3068-3077.
26. Ventrella E, Adamski Z, Chudzińska E, Miądowicz-Kobielska M, Marciniak P, Büyükgüzel E, Büyükgüzel K, Erdem M, Falabella P, Scranò L, Bufo SA. *Solanum tuberosum* and *Lycopersicon esculentum* leaf extracts and single metabolites affect development and reproduction of *Drosophila melanogaster*. *PLoS One* 2016; 11(5):e0155958.
27. Riaz B, Zahoor MK, Zahoor MA, Majeed HN, Javed I, Ahmad A, Jabeen F, Zulhussnain M, Sultana K. Toxicity, phytochemical composition, and enzyme inhibitory activities of some indigenous weed plant extracts in fruit fly, *Drosophila melanogaster*. *Evidence-based Compl Altern Med.* 2018; 2018:2325659.
28. Khan MF, Abutaha N, Nasr FA, Alqahtani AS, Noman OM, Wadaan MAM. Bitter gourd (*Momordica charantia*) possess developmental toxicity as revealed by screening the seeds and fruit extracts in zebrafish embryos. *BMC Compl Altern Med.* 2019; 19(1):184.
29. Deshmukh NS. Safety assessment of McB-E60 (extract of a *Momordica sp.*): Subchronic toxicity study in rats. *Toxicol Rep.* 2016; 3:481-489.